

### **Research Article**

# Journal of Marine Science Research and Oceanography

# Preparation and Characterization of Magnetic Iron Oxide (Fe<sub>3</sub>O<sub>4</sub>) Nanoparticles with Different Polymer Coating Agents

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Submitted: 31 Jan 2023; Accepted: 06 Feb 2023 Published: 15 Feb 2023

**Citation:** Karimova, A., Shirinova, H., Nargiz, G., Nuriyeva, S., Gahramanli, L. (2023). Preparation and Characterization of Magnetic Iron Oxide (Fe<sub>3</sub>O<sub>4</sub>) Nanoparticles with Different Polymer Coating Agents. *J Mari Scie Res Ocean*, 6(1), 19-24.

### **Abstract**

In the present study, the influence of the organic stabilizers on the formation of magnetic iron-oxide nanoparticles was investigated. Polyethylene glycol (PEG), dextran (DEX), and chitosan was chosen as biocompatible surface modification agents for obtained magnetic nanoparticles. The structure of the coated  $Fe_3O_4$  nanoparticles was learned by the X-ray diffraction and Fourier-transform infrared spectroscopic methods. It was explored that the PEG coated magnetic nanoparticles have relatively larger crystallite sizes, which indicate a more ordered crystal structure of these nanoparticles. Furthermore, FT-IR analysis showed that the  $Fe_3O_4$ -PEG system possesses a stronger nanoparticle-stabilizer interaction at the supramolecular level. This study emphasizes the significance of optimizing the surface properties of magnetic nanoparticles when using them in biomedical applications.

Keywords: Magnetic Iron Oxide Nanoparticles, Surface Modification, Biocompatible Coating Agents, Polymer Stabilizers

## Introduction

Magnetite nanoparticles (MNPs) have attracted significant attention for different biomedical applications, including heat mediators in hyperthermia, nanocarriers for drug delivery or biochemical molecules, and contrast agents in magnetic resonance imaging (MRI) [1-5]. In these applications, control of particle size, size distribution, shape as well as surface modification are critical steps required for effectively implementing the utilization of MNPs [6-10]. It was established that the unmodified MNPs have high surface energy which makes them thermodynamically unstable and grounds their agglomeration [11]. It is caused by the interaction of MNPs due to their strong magnetic dipole-dipole attraction and steric effects coming from van der Waals forces [12]. Aggregated MNPs are recognized by the reticuloendothelial system (RES) and are accordingly eliminated from the blood circulation [13]. Therefore, some coating procedures with various materials, for instance, long-chain-like polymer materials are required to improve their stability in highly ionic solutions [14-17]. For the obtaining of coated MNPs, polymers emerge as a priority among other materials due to their effortless and straightforward preparation and cost-effectiveness [18, 19]. Moreover, with an excellent hydrophilic feature, uncharged or weakly charged biocompatible polymers, such as polyethylenimine (PEI), polyethylene glycol (PEG), poly(lactic-co-glycolic acid) (PLGA), polyvinylpyrrolidone (PVP), polyvinyl alcohol (PVA), chitosan, starch, and dextran (DEX) have been widely used as protective shells to decrease the agglomeration of MNPs in the presence of serum proteins [20-29]. In addition, the temperature and pH-sensitive characteristics of polymer coating materials could allow for the management of the drug release from the MNPs, for instance, in targeted drug delivery systems [19, 30].

In this study, magnetic iron-oxide nanoparticles were synthesized by the chemical co-precipitation synthesis method, and the samples were produced by using PEG, DEX, and chitosan as biocompatible coating agents. The samples' structure characterization is accomplished by Fourier-transform Infrared Spectroscopy (FT-IR) and X-ray powder diffraction (XRD) spectra, as well as, Scanning Electron Microscopy (SEM) images.

# Materials and methods *Materials*

Iron (III) chloride tetrahydrate (FeCl<sub>2</sub>•4H<sub>2</sub>O), iron (III) chloride hexahydrate (FeCl<sub>3</sub>•6H<sub>2</sub>O), polyethylene glycol (PEG), dextran (DEX), chitosan (degree of deacetylation <90%), ammonium hydroxide (NH<sub>4</sub>OH, 23-25%) and acetic acid (CH<sub>3</sub>COOH, 99%) were purchased from Karma Lab (Izmir, Turkey). All chemicals were of analytical grade.

# Preparation of magnetite nanoparticles vith various coating materials

Magnetic iron oxide nanoparticles were obtained via the co-precipitation method. In this synthesis, FeCl<sub>3</sub>•6H<sub>2</sub>O, and FeCl<sub>2</sub>•4H<sub>2</sub>O, with a 2:1 mole ratio were each dissolved in 50 mL of deionized water. The two solutions were mixed into a 250 mL beaker glass and stirred and then heated until 70°C for 30 min under the nitrogen atmosphere. For coating with PEG appropriate amount of polymer was slowly added to the solution, and stirring was continuing about 1h. After it, 10-15 mL NH<sub>4</sub>OH 23-25% was added drop-wise until the pH was 10 and the temperature was increased until 90°C and solution was vigorously stirred for about 1h [31-33]. Further liquid and precipitate separated using NdFeB magnet and washed with deionized water and ethanol several times. Finally, PEG coated magnetic iron oxide nanoparticles were put in an oven and subsequently dried for 4h at 80°C.

To coat with dextran, 8 g polymer was added to the solution, and the whole process was repeated in the same way. Coating with chitosan was implemented, first, with the preparation of the chitosan solution. For this 0.25 g of fine chitosan powder was dissolved in the mixture of the 25 mL of deionized water and 1.25 mL of acetic acid. Chitosan coated magnetic nanoparticles were obtained in the above-described way.

# Characterization of coated magnetite nanoparticles

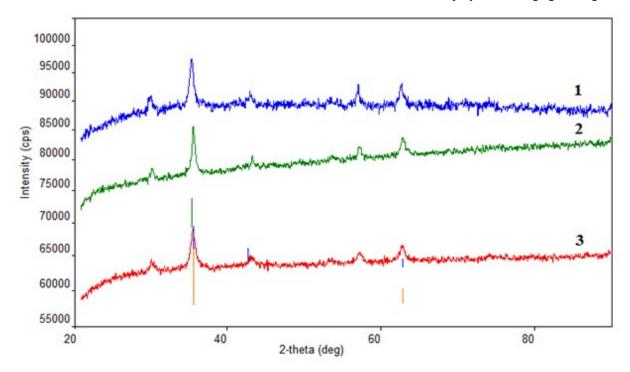
The synthesized and coated magnetic iron-oxide nanoparticles were characterized by SEM, XRD, and FT-IR techniques. The SEM (JEOL JSM-7600 F) was used for the morphology and size characterization. The XRD (Rigaku Mini Flex 600) with Cu Kα radiation was employed for the phase analysis. The FT-IR characterizations of magnetic Fe<sub>3</sub>O<sub>4</sub> nanoparticles with various coating agents were determined with Varian 3600. FT-IR spectra were obtained with KBr pellets and the spectrum was taken from 4000–450 cm<sup>-1</sup>.

### **Results and Discussion**

Figure 1 shows X-ray diffractograms of iron oxide nanoparticles coated with PEG, DEX, and chitosan. Characteristic X-ray lines are observed for magnetic nanoparticles prepared with all three coating agents. So, the d-spacing values of significant peaks match well with (ICDD DB card number 01-073-9877). Furthermore, the average crystallite size of Fe<sub>3</sub>O<sub>4</sub> nanoparticles obtained by different coating agents was calculated using Scherrer's equation as follows:

 $d = 0.9\lambda / \beta \cos\theta$ 

Calculations were carried out for 35,58°; 35,65°; 35,40° values of 2 theta angles for nanoparticles stabillized by PEG, DEX, and chitosan. The average crystallite size for iron oxide nanoparticles obtained with different polymer coating agents is given in Table 1.



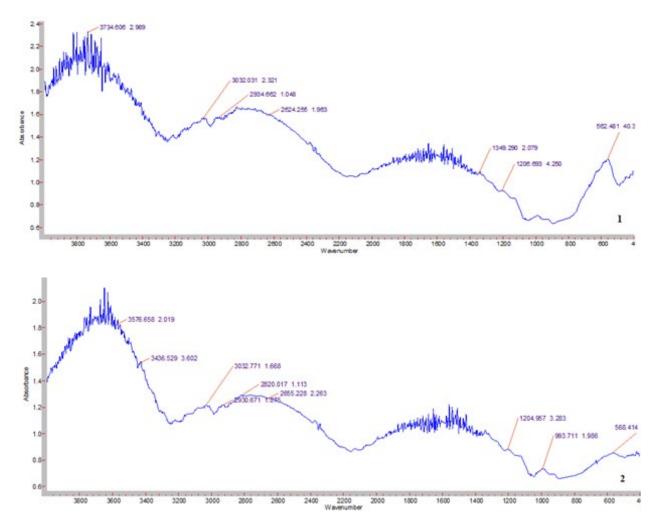
**Figure1:** X-ray diffractogram of iron oxide nanoparticles obtained by different coating agents: Fe<sub>3</sub>O<sub>4</sub>+PEG (1); Fe<sub>3</sub>O<sub>4</sub>+chitosan (2); Fe<sub>3</sub>O<sub>4</sub>+DEX (3).

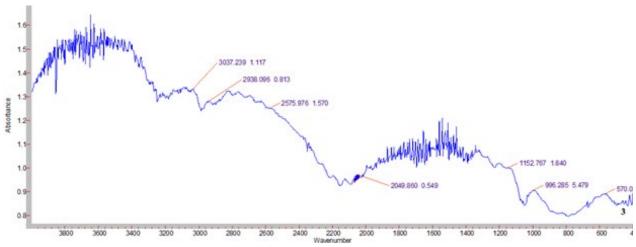
Table 1. The average crystallite size for iron oxide nanoparticles obtained with different coating agents

Samples	Fe <sub>3</sub> O <sub>4</sub> +PEG	Fe <sub>3</sub> O <sub>4</sub> +DEX	Fe <sub>3</sub> O <sub>4</sub> +chitosan
The average crystallite size of nanoparticles (nm)	25,30	25,17	25,10

The relatively narrow and sharp peaks in the XRD patterns of the Fe<sub>3</sub>O<sub>4</sub>+PEG indicate better crystalline characteristics of this sample than Fe<sub>3</sub>O<sub>4</sub>+DEX and Fe<sub>3</sub>O<sub>4</sub>+chitosan samples. Table 1 demosntrates that the magnetic nanoparticles modified by PEG have relatively larger crystallite sizes. It can be explained that a more ordered crystal structure is formed during the nucleation process of the iron oxide nanoparticles coated with PEG molecules [34,35]. Electrical neutrality, significant spatial repulsion, and high hydrophilicity of PEG molecules may have had a positive effect on the magnetic nanoparticles modification by this polymer [36]. It should also be noted that the effect of the molecular weight of PEG macromolecule on the average crystallite size of the magnetite nanoparticles is negligible [34].

Figure 2 shows the IR spectra of iron oxide nanoparticles obtained with different coated agents. The characteristic absorption bands of Fe<sub>3</sub>O<sub>4</sub> nanoparticles in the IR region are in the range of 400-600 cm<sup>-1</sup> [37]. The absorption lines at 570 cm<sup>-1</sup> on the FT-IR spectrum of DEX-coated iron oxide nanoparticles correspond to the vibrational oscillations of the Fe-O bond [38]. The same band was observed at 568cm<sup>-1</sup> on the spectrum of chitosan-coated nanoparticles [39]. On the spectrum of the PEG-coated Fe<sub>3</sub>O<sub>4</sub>, the band corresponding to the Fe-O bond was blue-shifted. The shift can be explained by the strengthening of the interaction between the stabilizer and the nanoparticle at the supramolecular level, the decrease in the degree of freedom of the atoms of the nanoparticle, in other words, the decrease in the frequency of oscillations of the Fe-O bond.





**Figure 2:** FTIR spectra of iron oxide nanoparticles obtained with different coating agents: Fe<sub>3</sub>O<sub>4</sub>+PEG (1); Fe<sub>3</sub>O<sub>4</sub>+chitosan (2); Fe<sub>3</sub>O<sub>4</sub>+DEX (3).

The results obtained from the IR spectra are in agreement with the XRD patterns. Among the considered coated agents, the most optimal coating for magnetic nanoparticles synthesized for biomedical purposes is PEG.

Figure 3 shows the SEM image of the surface morphology PEG-coated iron oxide nanoparticles. The scanning process was performed in SEI mode at an accelerating voltage of 15 kV and a working distance of 4.5 mm.

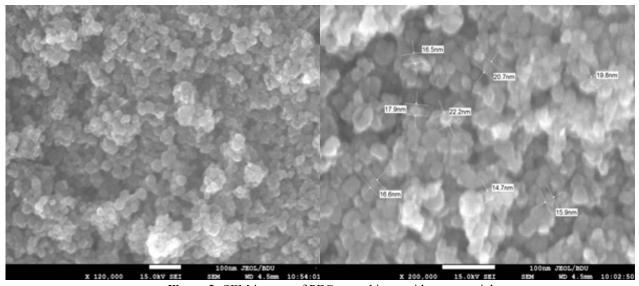


Figure 3: SEM images of PEG coated iron oxide nanoparticles.

As can be seen from the picture, the average size of Fe3O4 nanoparticles is 18-20 nm. This showed that the synthesized nanoparticles meets the size requirements for biomedical application [40, 41].

#### **Conclusion**

In the present study, magnetic iron-oxide nanoparticles was synthesized with the help of different biocompatible coating agents, namely, PEG, DEX, and chitosan. The structure of the produced nanoparticles was investigated by X-ray difractometer, FT-IR spectrometer and Scanning Electron Microsope. It was explored that magnetic nanoparticles coated by PEG macromalecules are more suitable for biomedical application. The nanoparticles stabilized by PEG have relatively larger crystallite sizes, which in-

dicate a more ordered crystal structure of these nanoparticles. Furthermore, FT-IR analysis showed that the Fe<sub>3</sub>O<sub>4</sub>+PEG system possesses a stronger nanoparticle-stabilizer interaction at the supramolecular level. The size of the PEG-coated nanoparticles was about 18-20 nm which is suitable for their biomedical application.

**Acknowledgement:** Many thanks to the late Mahammadali Ramazanov, without whom this project would never have been possible.

#### Funding

This research received no external funding.

### **Competing Interests**

The authors declare no conflict of interest.

### References

- Soares, P. I., Laia, C. A., Carvalho, A., Pereira, L. C., Coutinho, J. T., Ferreira, I. M., ... & Borges, J. P. (2016). Iron oxide nanoparticles stabilized with a bilayer of oleic acid for magnetic hyperthermia and MRI applications. Applied Surface Science, 383, 240-247.
- Teodorescuv, C., M., Săftoiu, A. (2016). Magnetic nanoparticles for hepatocellular carcinoma diagnosis and therapy. J Gastrointestinal Liver Dis, 25: 375-383.
- Lu, J. W., Yang, F., Ke, Q. F., Xie, X. T., & Guo, Y. P. (2018). Magnetic nanoparticles modified-porous scaffolds for bone regeneration and photothermal therapy against tumors. Nanomedicine: Nanotechnology, Biology and Medicine, 14(3), 811-822.
- Johannsen, M., Gneveckow, U., Eckelt, L., Feussner, A., et al. (2005). Clinical hyperthermia of prostate cancer using magnetic nanoparticles: presentation of a new interstitial technique. Int. J. Hyperthermia, 21: 637-647.
- Lee, C. P., Lai, M. F., Huang, H. T., Lin, C. W., & Wei, Z. H. (2014). Wheatstone bridge giant-magnetoresistance based cell counter. Biosensors and Bioelectronics, 57, 48-53.
- Molina, M. M., Seabra, A. B., de Oliveira, M. G., Itri, R., & Haddad, P. S. (2013). Nitric oxide donor superparamagnetic iron oxide nanoparticles. Materials Science and Engineering: C, 33(2), 746-751.
- Aillon, K. L., Xie, Y., El-Gendy, N., Berkland, C. J., & Forrest, M. L. (2009). Effects of nanomaterial physicochemical properties on in vivo toxicity. Advanced drug delivery reviews, 61(6), 457-466.
- 8. Akbarzadeh, A., Samiei, M., & Davaran, S. (2012). Magnetic nanoparticles: preparation, physical properties, and applications in biomedicine. Nanoscale research letters, 7, 1-13.
- Eberbeck, D., Wiekhorst, F., Wagner, S., & Trahms, L. (2011).
  How the size distribution of magnetic nanoparticles determines their magnetic particle imaging performance. Applied physics letters, 98(18), 182502.
- Issa, B., Obaidat, I. M., Albiss, B. A., & Haik, Y. (2013). Magnetic nanoparticles: surface effects and properties related to biomedicine applications. International journal of molecular sciences, 14(11), 21266-21305.
- 11. Ajdary, M., Moosavi, M. A., Rahmati, M., Falahati, M., Mahboubi, M., Mandegary, A., ... & Varma, R. S. (2018). Health concerns of various nanoparticles: A review of their in vitro and in vivo toxicity. Nanomaterials, 8(9), 634.
- 12. Mosayebi, J., Kiyasatfar, M., & Laurent, S. (2017). Synthesis, functionalization, and design of magnetic nanoparticles for theranostic applications. Advanced Healthcare Materials, 6(23), 1700306.
- 13. Veiseh, O., Gunn, J. W., & Zhang, M. (2010). Design and fabrication of magnetic nanoparticles for targeted drug delivery and imaging. Advanced drug delivery reviews, 62(3), 284-

- 304.
- 14. Bohara, R. A., Thorat, N. D., & Pawar, S. H. (2016). Role of functionalization: strategies to explore potential nano-bio applications of magnetic nanoparticles. RSC advances, 6(50), 43989-44012.
- 15. Ankamwar, B. (2012). Size and shape effect on biomedical applications of nanomaterials. Biomedical Engineering-Technical Applications in Medicine, 93-114.
- Mylkie, K., Nowak, P., Rybczynski, P., & Ziegler-Borowska, M. (2021). Polymer-coated magnetite nanoparticles for protein immobilization. Materials, 14(2), 248.
- 17. Rahimi, M., Safa, K. D., & Salehi, R. (2017). Co-delivery of doxorubicin and methotrexate by dendritic chitosan-g-mPEG as a magnetic nanocarrier for multi-drug delivery in combination chemotherapy. Polymer Chemistry, 8(47), 7333-7350.
- 18. Dalal, A. A., Seyed, Y. M., Alexander, S. (2021). Emerging Application of Magnetic Nanoparticles for Diagnosis and Treatment of Cancer. Polymers, 13(23):4146.
- Sundaresan, V., Menon, J. U., Rahimi, M., Nguyen, K. T., & Wadajkar, A. S. (2014). Dual-responsive polymer-coated iron oxide nanoparticles for drug delivery and imaging applications. International journal of pharmaceutics, 466(1-2), 1-7.
- Li, H., Jin, Z., Cho, S., Ko, S. Y., Park, J. O., & Park, S. (2016, August). Polyethylenimine-coated magnetic nanoparticles with improved biocompatibility for hyperthermia. In 2016 13th International Conference on Ubiquitous Robots and Ambient Intelligence (URAI) (pp. 466-470). IEEE.
- Ardiyanti, H., Suharyadi, E., Kato, T., & Iwata, S. (2016, April). Crystal structures and magnetic properties of magnetite (Fe3O4)/polyvinyl alcohol (PVA) ribbon. In AIP Conference Proceedings (Vol. 1725, No. 1, p. 020007). AIP Publishing LLC.
- 22. Jokerst, J. V., Lobovkina, T., Zare, R. N., & Gambhir, S. S. (2011). Nanoparticle PEGylation for imaging and therapy. Nanomedicine, 6(4), 715-728.
- 23. Tansık, G., Yakar, A., & Gündüz, U. (2014). Tailoring magnetic PLGA nanoparticles suitable for doxorubicin delivery. Journal of nanoparticle research, 16, 1-13.
- 24. Yang, G., Zhang, B., Wang, J., Wang, M., Xie, S., & Li, X. (2016). Synthesis and characterization of poly (lactic acid)-modified superparamagnetic iron oxide nanoparticles. Journal of Sol-Gel Science and Technology, 77, 335-341.
- 25. Darwish, M. S., Al-Harbi, L. M., & Bakry, A. (2022). Synthesis of magnetite nanoparticles coated with polyvinyl alcohol for hyperthermia application. Journal of Thermal Analysis and Calorimetry, 147(21), 11921-11930.
- Karimzadeh, I., Aghazadeh, M., Ganjali, M. R., Norouzi, P., Doroudi, T., & Kolivand, P. H. (2017). Saccharide-coated superparamagnetic Fe3O4 nanoparticles (SPIONs) for biomedical applications: An efficient and scalable route for preparation and in situ surface coating through cathodic electrochemical deposition (CED). Materials Letters, 189, 290-294.
- Thapa, B., Diaz-Diestra, D., Beltran-Huarac, J., Weiner, B. R.,
  Morell, G. (2017). Enhanced MRI T 2 relaxivity in con-

- trast-probed anchor-free PEGylated iron oxide nanoparticles. Nanoscale Research Letters, 12(1), 1-13.
- 28. Harris, J. M., & Chess, R. B. (2003). Effect of pegylation on pharmaceuticals. Nature reviews Drug discovery, 2(3), 214-221.
- Yoffe, S., Leshuk, T., Everett, P., & Gu, F. (2013). Superparamagnetic iron oxide nanoparticles (SPIONs): synthesis and surface modification techniques for use with MRI and other biomedical applications. Current pharmaceutical design, 19(3), 493-509.
- 30. Reyes-Ortega, F., Delgado, Á. V., Schneider, E. K., Checa Fernández, B. L., & Iglesias, G. R. (2017). Magnetic nanoparticles coated with a thermosensitive polymer with hyperthermia properties. Polymers, 10(1), 10.
- 31. Wu, W., Wu, Z., Yu, T., Jiang, C., & Kim, W. S. (2015). Recent progress on magnetic iron oxide nanoparticles: synthesis, surface functional strategies and biomedical applications. Science and technology of advanced materials, 16(2), 023501.
- 32. Wroblewski, C., Volford, T., Martos, B., Samoluk, J., Martos, P. (2020). High Yield Synthesis and Application of Magnetite Nanoparticles (Fe3O4). Magnetochemistry, 6:22.
- 33. Wu, S., Sun, A., Zhai, F., Wang, J., Xu, W., Zhang, Q., & Volinsky, A. A. (2011). Fe3O4 magnetic nanoparticles synthesis from tailings by ultrasonic chemical co-precipitation. Materials Letters, 65(12), 1882-1884.
- 34. Namikuchi, E. A., Gaspar, R. D., da Silva, D. S., Raimundo, I. M., & Mazali, I. O. (2021). PEG size effect and its interaction with Fe3O4 nanoparticles synthesized by solvothermal method: morphology and effect of pH on the stability. Nano Express, 2(2), 020022.

- 35. Cîrcu, M., Nan, A., Borodi, G., Liebscher, J., & Turcu, R. (2016). Refinement of magnetite nanoparticles by coating with organic stabilizers. Nanomaterials, 6(12), 228.
- 36. D'souza, A. A., & Shegokar, R. (2016). Polyethylene glycol (PEG): a versatile polymer for pharmaceutical applications. Expert opinion on drug delivery, 13(9), 1257-1275.
- 37. Martínez-Cabanas, M., López-García, M., Barriada, J. L., Herrero, R., & de Vicente, M. E. S. (2016). Green synthesis of iron oxide nanoparticles. Development of magnetic hybrid materials for efficient As (V) removal. Chemical Engineering Journal, 301, 83-91.
- 38. Yusoff, A. H. M., Salimi, M. N., & Jamlos, M. F. (2017, April). Synthesis and characterization of biocompatible Fe3O4 nanoparticles at different pH. In AIP Conference Proceedings (Vol. 1835, No. 1, p. 020010). AIP Publishing LLC.
- 39. Ahangaran, F., Hassanzadeh, A., & Nouri, S. (2013). Surface modification of Fe 3 O 4@ SiO 2 microsphere by silane coupling agent. International Nano Letters, 3, 1-5.
- 40. Dheyab, M. A., Aziz, A. A., Jameel, M. S., Noqta, O. A., Khaniabadi, P. M., & Mehrdel, B. (2020). Simple rapid stabilization method through citric acid modification for magnetite nanoparticles. Scientific reports, 10(1), 10793.
- 41. Dheyab, M. A., Aziz, A. A., Jameel, M. S., Abu, Noqta. O., Khaniabadi, P, M., et al. (2020). Simple rapid stabilization method through citric acid modification for magnetite nanoparticles. Sci. Rep, 10:1-8.

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