ANALYSIS

DETERMINATION OF FLUCONAZOLE AND ITS OXIDATION PRODUCTS WITH KINETIC EVALUATION UNDER POTASSIUM PERMANGANATE TREATMENT IN ACIDIC SOLUTIONS BY ULTRA PERFORMANCE LIQUID CHROMATOGRAPHY-TANDEM MASS SPECTROMETRY

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Abstract: For the determination of fluconazole (FLU) oxidation stability under permanganate treatment at the acidic pH, a sensitive, reproducible, and stability-indicating ultra-performance liquid chromatography–mass spectrometry (UPLC/MS) method was developed. Three additional products ($t_R = 5.79, 6.98, 7.54$) were observed besides the FLU ($t_R = 6.22$). The proposed method was used to study the kinetics of FLU oxidative degradation. An oxidation process followed the kinetics of the second-order reaction. The degradation rate constant and the corresponding half-life obtained for the FLU oxidative degradation were 0.5626 h^{-1} and 16.69 h , respectively. The putative oxidation products were characterized and their fragmentation pathways, on a basis of MS/MS data, were proposed.

Keywords: fluconazole, stability, oxidation studies, kinetic evaluation, UPLC/MS

Fluconazole (FLU), [2-(2,4-difluorophenyl)-1,3-bis(1*H*-1,2,4-triazol-1-yl)propan-2-ol], a synthetic antifungal agent of the imidazole class, is systematically used in oral, esophageal, and vaginal candidiasis, as well as in the treatment of cryptococcal infections in patients with the acquired immunodeficiency syndrome (AIDS). FLU interacts with 14- α demethylase, a cytochrome P-450 enzyme necessary to convert lanosterol to ergosterol—an essential component of the fungal cell membrane. Fluconazole is eliminated primarily by renal excretion, with approximately 80% of the administered dose appearing in the urine as unchanged drug (1-3). Different analytical methods, for example, spectrophotometry (4, 5); capillary electrophoresis (6); micellar electrokinetic capillary chromatography (7); RP-HPLC (1, 3, 8), UPLC (9), or UPLC-MS/MS (10); and gas chromatography (11, 12), were reported for the determination of FLU in pharmaceutical preparations and biological matrices.

The stability of active pharmaceutical ingredients (APIs) affects quality, efficacy and usage safe-

ty of APIs. The chemical and physical stability evaluation process consists of many tests. These tests should be conducted in a planned way following the guidelines issued by ICH, WHO, or FDA (13-15). The most important stability tests are stress tests, which enable to identify degradation products. A list of stress factors suggested for forced degradation studies must include acid and base hydrolysis, thermal degradation, photolysis, and oxidation (13). The regulations do not specify the oxidation agents, which should be used, in the case of oxidation. A 0.1–3% solutions of hydrogen peroxide are widely used in forced degradation studies. Other chemicals useful in oxidation reactions include, for example, potassium permanganate (KMnO_4), sodium hypochlorite, singlet oxygen, or Fenton's reagent (16).

KMnO_4 is widely used as an oxidizing agent in synthetic and in analytical chemistry and is also applied as a disinfectant. It is effective over a wide pH range. Potassium permanganate is the most potent oxidation state both in alkaline and in acid

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mediums. KMnO_4 can be used to differentiate degradation products that can be related to drug and its degradation products. Oxidation by permanganate ion is applied in organic syntheses since the advent of phase-transfer catalysis (17, 18).

There are reports in which FLU was subjected to stress conditions of acid, alkali, and neutral hydrolysis; oxidation; photolysis (in solutions and solid phase); and thermal decomposition by RP-HPLC. Extensive degradation was found to occur in oxidative medium under thermal stress (19). The forced oxidation degradation of FLU using hydrogen peroxide by spectrophotometric method was performed (2). The literature survey also evidenced the results of the stability of reconstituted FLU oral suspension (20).

Shortage of information concerning reaction of FLU with oxidants prompted us to investigate the influence of KMnO_4 on this drug stability. Herein, we report on the development of a new UPLC-MS/MS method for the determination of FLU and its oxidation products during reaction with KMnO_4 at the acidic pH. The method was used for kinetic studies and identification of obtained degradation products of FLU.

Chemicals and methods

Chemicals and reagents

Fluconazole; 99.35%, SlovakoFarma, Batch 00/1040/01. HPLC grade glacial acetic acid was purchased from AppliChem (Germany). HPLC grade acetonitrile and formic acid (98%) were purchased from J.T. Baker. HPLC grade water was obtained from HLP 5 (HYDROLAB Poland) apparatus and was filtered through 0.2 μm filter before use.

Standard solution

The amount of 50.8 mg of FLU, weighed with a precision of 0.1 mg, was dissolved in the volume of 2.5 mL of glacial acetic acid and filled (up to the 25 mL) with dilution water. For method validation, five solutions with concentration of 0.0609, 0.0812, 0.1015, 0.1218, and 0.1421 mg/mL of FLU were prepared.

Oxidation study of the drug substance

The amount of 0.5 mL of solution of FLU (2.03 mg/mL), 3.5 mL demineralized water, 1.0 mL of concentrated sulfuric acid, 5.0 mL 0.02 M KMnO_4 , and 5 μL of manganese dioxide solution was added

Table 1. Validation of the method.

Parameter	Fluconazole
t_R (min)	6.22 $\pm$ 0.05
Limit of detection, [$\mu\text{g/mL}$]	4.10
Limit of quantitation, [$\mu\text{g/mL}$]	12.60
Linearity range, [mg/mL]	0.0609 – 0.1421
Regression coefficients $P = a \cdot c + b \pm S_e^b$	$a = 4696.7$ $b = 6.7240$
Standard deviation of the regression coefficients	$S_a = 53.54194$ $S_b = 5.64771$
Correlation coefficient, R	0.9992
Determination coefficient, R^2	0.9982
Normality of residuals ^c (Shapiro-Wilk test)	0.9493 ($p = 0.5131$)
Precision (% RSD)	level 50%: 0.56% level 100%: 0.63% level 150%: 0.89%
Indirect precision (% RSD)	level 50%: 0.93% level 100%: 0.79% level 150%: 1.02%
Recovery [%] $n = 3$	level 80%: 100.17% level 100%: 99.67% level 120%: 98.11%

^aMean $\pm$ SD ($n = 5$). ^bP = peak area; c = concentration; a and b = regression coefficients, S_e = standard error of the estimate, S_a = standard deviation of the slope, S_b = standard deviation of the intercept, ^cNormal distribution of residuals if $p > 0.05$, RSD = relative standard deviation.

Figure 1. UPLC chromatogram overlay of FLU oxidation induced by KMnO_4 in acidic conditions

to 10.0 mL flasks. The test solutions were incubated at room temperature and 1 μL of each reaction mixture was injected onto ACQUITY UPLC system after 36, 47, 54, 71, and 110 h, respectively.

Before the measurements of test samples, the analysis of solutions containing identical components as test samples but without KMnO_4 was done. The analyses were performed in triplicate.

UPLC/MS/MS analysis

The UPLC-MS/MS system consisted of a Waters ACQUITY® UPLC® (Waters Corporation, Milford, MA, USA) coupled to a Waters TQD mass spectrometer (electrospray ionization mode ESI-tandem quadrupole). Chromatographic separations were carried out using the Acquity UPLC BEH (bridged ethyl hybrid) C18 column; 2.1×100 mm, and 1.7 μm particle size, equipped with Acquity UPLC BEH C18 VanGuard pre-column; 2.1×5 mm, and 1.7 μm particle size. The column was maintained at 40°C , and eluted under gradient conditions from 100% to 50% of eluent A over 13 min, at a flow rate of 0.3 mL/min. Eluent A: 0.1% (v/v) formic acid in water; eluent B: 0.1% (v/v) formic acid in acetonitrile. 1 μL of each sample was injected. Chromatograms were made using Waters eë PDA detector. Spectra were analyzed in 200–700 nm range with 1.2 nm resolution and sampling rate of 20 points/s.

MS detection settings of Waters TQD mass spectrometer were as follows: source temperature 150°C , desolvation temperature 350°C , desolvation gas flow rate 600 L/h, cone gas flow 100 L/h, capillary potential 3.00 kV, cone potential 20 V. Nitrogen was used for both nebulizing and drying gas. The data were obtained in a scan mode ranging from 50 to 1000 m/z in time 0.5 s intervals.

Collision-activated dissociations (CAD) analyses were carried out with the energy of 30 eV, and all the fragmentations were observed in the source. Consequently, the ion spectra were obtained by scanning from 50 to 1000 m/z range. Data acquisition software was MassLynx V 4.1 (Waters).

Method validation

The described method was validated for the determination of FLU in the presence of oxidation products by UPLC method according to ICH guidelines (21).

Specificity

To demonstrate the specificity of the developed UPLC method the solution of FLU after oxidation stress was analyzed. Oxidation study was performed in KMnO_4 solution at the acidic pH; the solution was left for 15 min at room temperature. Peak purity test was carried out for FLU peak by using MS detector in a stressed sample.

Linearity

The calibration plot for the method was constructed by analysis of five solutions containing different concentrations of FLU in the range 0.0609–0.1421 mg/mL. Solutions were injected into a column in the amounts of 1 μL . A further analytical procedure was as described in the UPLC Conditions. Linearity was assessed in triplicate on the basis of the relationship between peak areas and concentration, in milligrams per milliliter. The linear coefficients, standard deviation of slope and intercept, correlation coefficient R, determination coefficient (R^2), standard error of residuals of the calibration curve and to determine whether the residuals have a normal distribution, the Shapiro-

Wilk statistical test were calculated using the program Statistica v. 10.

Limit of detection (LOD) and limit of quantification (LOQ)

Based on the standard error of residuals (Se) and the slope (a) of the calibration plot and following the formula $LOD = 3.3Se/a$ and $LOQ = 10Se/a$, the LOD and LOQ for FLU was estimated.

Precision

The repeatability of the method was checked by a threefold analysis of the three concentrations levels 0.0508 mg/mL, 0.1015 mg/mL and 0.1523

mg/mL (50%, 100% and 150%) of FLU solutions. The same protocol was followed for three different days to study the intermediate precision of the proposed method. Different analysts prepared different solutions on different days. The RSD (%) of the peak area of FLU was calculated.

Accuracy

The accuracy of the method was evaluated in triplicate (n = 3) at the three concentrations of 0.0812, 0.1015 and 0.1218 mg/mL (80%, 100%, and 120%) of FLU solutions, and the recovery was calculated for each added (externally spiked) concentration.

Table 2. Kinetic results of FLU oxidation by KMnO₄ in acidic media at room temperature.

Rate constant, k [h ⁻¹]	t _{0.1} [h] ^a	t _{0.5} [h] ^a	R ^a
0.5626	1.85	16.69	0.9716

^at_{0.5} = half-life, t_{0.1} = time after which 10% of the substrate decomposes, and R = correlation coefficient.

Figure 2. MS/MS spectra of FLU and its degradation products: FP-2 and FP-3

Figure 3. The $1/c = f(t)$ graph of oxidation of FLU by KMnO_4 in acidic medium

Figure 4. Fragmentation pattern of product FLU

Robustness

To demonstrate the robustness of the method deliberate small changes of flow rate, the content of acetonitrile and column temperature were made around the optimal values. The mobile phase flow rate was 0.30 mL/min; to study the effect of the flow rate on the resolution, the flow rate was changed to 0.27 and 0.33 mL/min. The effect of the column temperature was studied at 36°C and 44°C (instead of 40°C), and the mobile phase composition was changed +10% from the initial composition.

RESULT AND DISCUSSION

Result and discussion

ICH guideline requires stress testing to be performed to elucidate the inherent stability characteristics of the active substance (13). Ideal stability-indicating method is one that quantifies the standard drug alone without any interference from its degradation products. For the separation and identification of examined constituents and for the kinetic studies of FLU in the presence of KMnO_4 in acidic

solution, the developed and validated UPLC-MS/MS method is suitable.

Optimization of chromatographic conditions

The UPLC/MS method was chosen to evaluate the degradation process of FLU under potassium permanganate treatment in an acidic solution. With the development of the chromatographic method enabling the determination of FLU and its oxidation products, a test procedure was started. After oxidation stress, the FLU solution was analyzed. Satisfactory results were achieved using gradient conditions from 100% to 0% of water/formic acid (0.1%, v/v) over 13 min as eluent A and acetonitrile/formic acid (0.1%, v/v) as eluent B, at a flow rate of 0.3 mL/min. The retention time for FLU in the developed condition was $\approx$ 6.22 min.

Method validation

The developed UPLC method was specific to FLU and guaranteed to obtain well-shaped peaks both for active substance and coexisting oxidation products. Peaks of FLU were well resolved from

products FP-1, FP-2, FP-3 in chromatograms, and no interference that could have an influence on the obtained results was observed.

Good linearity was observed in the concentration range 0.0609 – 0.1421 mg/mL of FLU. The correlation coefficient and determination coefficient (R^2) obtained for the linear model were greater than 0.99. The y-intercept of the linear equation for FLU was statistically insignificant. The distribution of the residuals can be approximated with a normal distribution as it is shown by the p-value ($p > 0.05$) of the Shapiro–Wilk normality test. Sensitivity of the method was good. The limit of detection (LOD) and limit of quantification (LOQ) values were found to be 4.10 and 12.60 $\mu\text{g/mL}$, respectively. Accuracy of the method expressed as % recovery at three concentration levels was from 98.11% to 100.17%. Good precision and intermediate precision with % RSD less than 1.02% was observed. Table 1 presents the detailed results. In all the deliberately varied chromatographic conditions (flow rate, column temperature, mobile phase composition), FLU and

Table 3. Proposed products of degradation of FLU.

Product Id	tR	[M+H] ⁺	Fragmentation ions	Proposed structure
FP-1	5.79	323.1	70.0, 98.0, 139.0, 151.0, 169.0, 208.1, 220.1, 236.1, 305.1
FLU	6.22	307.1	70.0, 127.0, 139.0, 151.0, 169.0, 220.1, 238.1
FP-2	6.98	224.1	70.0, 127.0, 155.0
FP-3	7.54	159.0	-

Figure 5. Fragmentation pattern of product FP-1

Figure 6. Fragmentation pattern of product FP-2

degradation products were adequately resolved, and the order elution remained unchanged.

Oxidation of FLU by KMnO_4 in acidic medium

The oxidative ability of KMnO_4 depends on the acidity of a reaction solution. The oxidation potential of KMnO_4 decreases with increasing pH value and counts +1.23 V at pH 3, and at pH 5, it reaches +1.07 V; in neutral solution or in basic solution, it was lower and equaled +0.58 V and +0.56 V, respectively (22). Chromatograms obtained for acid solution containing only FLU showed one peak with retention time $t_R \approx 6.22$ min, whereas in chromatograms obtained for test solutions containing

KMnO_4 , with the addition of concentrated sulfuric acid, three additional peaks of oxidation products of FLU with retention times such as **FP-1** $t_R \approx 5.79$, **FP-2** $t_R \approx 6.98$, and **FP-3** $t_R \approx 7.54$ were observed besides peak of FLU $t_R \approx 6.22$ min (Fig. 1). Figure 2 presents the MS/MS spectra of FLU and degradation products such as FP-2 and FP-3. The degradation process of FLU increased with the prolongation of incubation time, and after 110 hours, decomposition of FLU was 82.12%.

Kinetic evaluation

The analysis of the equation $1/c = f(t)$ for the oxidation of FLU induced by KMnO_4 under acidic condi-

tions revealed that the process followed the kinetics of second-order reaction (Fig. 3). Table 2 present the calculated values of k , $t_{0.1}$, and $t_{0.5}$. The linear regression equation was $y = -11.41x + 0.5626$. The correlation coefficient and determination coefficient (R^2) obtained for linear model were found to be 0.9857 and 0.9716, respectively. The standard deviation of slope and intercept were 0.06 and 3.82, respectively.

Identification of oxidation products of FLU

The identification of photodegradation products of fluconazole was performed on a basis of UPLC/MS analysis and supported with fragmentation patterns obtained from MS/MS experiments. The structures of the proposed degradation products are shown in Table 3. The structures of the presented stable products of the degradation were proposed on basis of the results of Collisionally Activated Decomposition (CAD) experiments.

FLU was found at the retention time of 6.22 min with a molecular ion peak of m/z 307 in the positive mode. Methylene group was most susceptible to oxidation in close vicinity to triazole moiety. Hydroxylation of this group yielded product FP-1. Further oxidation of this compound led to the loss of triazolylmethylene moiety and dehydration, as it is observed in the case of product FP-2. Finally, degradation of FP-2 involved loss of triazole-1-carbonyl moiety and oxidation of methylene group to carboxylic group, yielding 2,4-difluorobenzoic acid (FP-3). Figures 4-6 show the degradation patterns. This quantity of degradation products found is the largest compared with previously published papers (2, 19). The proposed chemical structures of the oxidation products shown in Table 3 are new, compared to those reported in earlier publications. Azole compounds, which are imidazole and triazole derivatives, showed strong differences in their stability (23-26). One of the most possible routs of degradation of azole antifungal drugs is hydroxylation that can result in the ring opening or loss of imidazole moiety as observed for, for example, clotrimazole or bifonazole (23, 24). Similarly, in this case, hydroxylation was the first step in the oxidative degradation of FLU leading to FP-1. The product FP-2 at a retention time of 6.98 min and the molecular ion peak (m/z 224) were the consequences of further hydroxylation, which resulted in loss of one of triazole ring.

CONCLUSIONS

We have developed new stability indicating UPLC-MS/MS method for the determination of FLU and its oxidation products. The method meets

the acceptance criteria for validation and may be used for the kinetic analysis of FLU oxidation. The process followed kinetics of the second-order reaction for the substrate. Finally, MS/MS experiments allowed to determine that FLU oxidation results in three derivatives.

REFERENCES

1. Abdel-Moetty E.M., Khattab F.I., Kelani K.M., AbouAl-Alamein A.M.: *Farmaco* 57, 931 (2002).
2. Lotfy H.M., Monir H.H., Abd El-Aleem A.E.E.: *J. Chil. Chem. Soc.* 57, 1447 (2012)
3. Meshram D.B., Bagade S.B., Tajne M.R.: *J. Chromatogr. Sci.* 47, 885 (2009).
4. Belal F., Sharaf El-Din M.K., Eid M.I., El-Gamal R.M.: *J. Chromatogr. Sci.* 4, 298 (2014).
5. El-Bayoumi A., El-Shanawany A.A., El-Sadek M.E., Abd El-Sattar A.: *Spectrosc. Lett.* 30, 25 (1997).
6. Crego A.L., Marina M.L., Lavandera J.L.: *J. Chromatogr. A* 1-2, 337 (2001).
7. Heeren F., Tanner R., Theurillat R., Thormann W.: *J. Chromatogr. A* 745, 165 (1996).
8. Ayub A.C., Vianna-Soares C.D., Ferreira L.A.M.: *J. Chromatogr. Sci.* 45, 286 (2007).
9. Mistretta V., Dubois N., Denooz R., Charlier C.: *Acta Clin. Belg.* 1, 53 (2014)
10. Decosterd L.A.: *Antimicrob. Agents Chemother.* 12, 5303 (2010).
11. Ekiert R.J., Krzek J., Czekaj J.S., Hubicka U.: *Acta Chromatogr.* 21, 273 (2009).
12. Lima D.M., Nunes Junior, G.P., Ferri P.H., Santos S.C.: *Braz. J. Pharm. Sci.* 2, 223 (2005).
13. http://www.ich.org/fileadmin/public/Web_Site/ICH_Products/Guidelines/Quality/Q1A_R2/Step4/Q1A_R2_Guideline.pdf (2003).
14. http://www.who.int/medicines/areas/quality_safety/quality_assurance/stblty-testing-APIsand_FPPS-QAS17-694_12012017.pdf (2017).
15. Jamrógiewicz M.: *Front Pharmacol.* 7, 17 (2016).
16. Baertschi S.W., Jansen P.J., Alsante K.M.: in *Pharmaceutical Stress Testing: Predicting Drug Degradation*, 2nd Ed., Baertschi S.W., Alsante K.M., Reed R.A., p. 27, Taylor & Francis, 2005.
17. Fingerman M.: *Bioremediation of Aquatic and Terrestrial Ecosystems Chapter 8 Remediating RDX and HMX Contaminated Soil and Water*. CRC Press, 2016.
18. Dash S., Patel S., Mishra B.K.: *Tetrahedron* 65, 707 (2009).

19. Lotfy H.M., Al-Byoumy Abdel-Aleem A.A., Monir H.H.: J. Liq. Chromatogr. Rel. Technol. 36, 1013 (2013).
20. Dentiger P.J., Swenson Ch.: Ann. Pharmacother. 43, 485 (2009).
21. [http://www.ich.org/fileadmin/public Web Site/ICH Products/Guidelines/Quality/Q2 R1/Step4/Q2 R2 Guideline.pdf](http://www.ich.org/fileadmin/public/WebSite/ICH_Products/Guidelines/Quality/Q2_R1/Step4/Q2_R2_Guideline.pdf) (2005)
22. Kocjan R. (Ed), Analytical Chemistry, pp. 518-519, PZWL, Warszawa 2000.
23. Kryczyk A., Żmudzki P., Hubicka U.: Biomed. Chromatogr. 31, 9 (2017).
24. Kryczyk A., Żmudzki P., Koczurkiewicz P., Piotrowska J., Pękała E., Hubicka U.: J. Pharm. Biomed. Anal. 145 (2017).
25. Kryczyk A., Żmudzki P., Hubicka U.: Biomed. Chromatogr. 30, 11 (2016).
26. Ekiert R.J, Krzek J.: Curr. Issues Pharm. Med. Sci. 26, 3 (2013).

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