Bacterial Load of *Clostridium Perfringens* in the Aquatic Environment of Dal Lake, Kashmir

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Abstract

The study was aimed to investigate the presence of *Clostridium. perfringens* (*C. perfringens*) and estimate its bacterial load in the aquatic environment of the Dal Lake, Kashmir. A total of 45 water samples (9 from each site) were collected randomly from 5 different sites viz., Dhobi Ghat, Telbal Nallah, Char-Chinari, Centaur Lake View and Dalgate of the Dal Lake. Using species-specific 16S rRNA gene amplification, all the 45 (100%) water samples were found to be positive for *C. perfringens* confirming the presence of *C. perfringens* in the lake. The overall *C. perfringens* load in water samples was found to be in the range of 9.2×10 CFU/ml to 1.87×10^2 CFU/ml, which is much less than the critical CFU level of *C. perfringens* (10^5 - 10^8). The highest average CFU/ml value was found at Dalgate (1.87×10^2), followed by Telbal Nallah (1.78×10^2), Dhobi Ghat (1.20×10^2), Centaur Lake View (1.05×10^2) and Char-Chinari (9.2×10), respectively. Statistically, it was observed that there is a significant difference in average CFU/ml values of *C. perfringens* at the five sites of the lake ($P < 0.01$). The current study documented the presence of *C. perfringens* in the aquatic environment of the Dal Lake for the first time in the Kashmir valley.

Keywords: *C. perfringens*; Anaerobe; 16S rRNA; PCR; CFU

Introduction

A large number of infectious diseases are transmitted primarily through water supplies contaminated with human and animal

excreta [25]. There have been many outbreaks of water borne diseases at the global level but the situation is especially serious in developing countries due to their high prevalence and mortality [9].

The water borne pathogens that pose a serious human health risk whenever present in drinking water include *C. perfringens*, *Salmonella* spp., *Shigella* spp., *Campylobacter* spp., among many others [15].

C. perfringens is a Gram-positive, anaerobic, non-motile bacillus. Besides its cosmopolitan distribution, it is a normal inhabitant of human and animal intestines [3,8,12,20]. Under unfavourable environmental conditions, it generates highly resistant endospores [26]. *C. perfringens* is one of the most prolific toxin-producing bacteria, forming at least 20 distinct toxins [10]. Based on the toxins produced, there are 7 strains of *C. perfringens* viz; type A (alpha toxin), type B (alpha, beta and epsilon toxins), type C (alpha and beta toxins), type D (alpha and epsilon toxins), type E (alpha and iota toxins), type F (alpha toxin and enterotoxin) and type G (alpha and NetB toxins) [10]. All the isolates of *C. perfringens* produce alpha toxin [21]. *C. perfringens* is believed to be the most wide-spread bacterial pathogen in nature responsible for causing a number of disease syndromes in humans and animals [16]. *C. perfringens* is also important as an indicator of faecal contamination in aquatic habitats from which the use of water for portable purpose can put human health at risk due to the production of various toxins affecting the nervous and gastrointestinal systems of the consumer [17]. The toxoinfection usually occurs when the load of *C. perfringens* in water or food is above 10^5 to 10^8 CFU/g [13,14,20].

The aquatic resources in the valley of Kashmir have a considerable impact on the socio-economic status of the local people as they provide portable water and sustain commercial activities of fisheries, agriculture and tourism. However, these resources are facing grave pollution problems due to various anthropogenic activities notably the disposal of raw sewage and domestic wastes from catchment areas. These problems are more pronounced in lentic ecosystems like Dal Lake which has witnessed a terrible deterioration in its water quality over the past few decades [19] and the probability of *C. perfringens* in its waters is high. Dal Lake, one of the famous lakes of India with its pristine glory, is a source of potable and recreational water as well as a source of cheap, affordable protein in the form of fish [2]. Water of the lake is continuously being fed with human excreta through sewage dumping, although some partial treatment of the sewage is done via treatment plants. The lake is in a serious state of eutrophication mostly cultural and ways and means for lake restoration have not proved to be fruit full till date. The floating gardens of the lake that have originated with time have now assumed a status of biggest vegetable producing bowl of Kashmir. The polluted water acts as a carrier of viral transmission and since the lake is used as a sewage dump, a tourist hot spot and thus might act as a concentration tool for the bacterial propagation. It is pertinent to mention here that various pathotypes of *Escherichia coli* have been reported from diarrhoeic children of Kashmir [27].

Materials and Methods

Sampling

An urban water body, Dal Lake, located in Srinagar city at 1584 masl (latitude 34° 07' N and longitude 74° 52' E) with total area of 11.4 sq. km and catchment area of 316 sq. km, was selected for the current study [16]. Five 5 sampling sites were chosen in the lake: Dalgate, Telbal Nallah, Dhobi Ghat, Centaur Lake View and Char-Chinari. A total of 45 water samples (9 from each site) were collected in sterile plastic tubes and were transported to the laboratory for immediate processing.

Isolation and identification of *C. perfringens*

For isolation of *C. perfringens*, the water samples were inoculated into Difco™ Cooked meat medium (Becton, Dickinson and Company, USA) and incubated anaerobically in 3.5 litre anaerobic jars (Oxoid Limited, Thermo Fisher Scientific Inc., UK) with Gas-Pak™ Anaerobe Container System (Becton, Dickinson and Company, USA) at 37°C for 48 hours. The enriched samples were then streaked on Sulfite Polymyxin Sulphadiazine agar plates (SPS HiVeg™ Agar, Modified; Hi-media Laboratories, Mumbai, India) and the plates were incubated anaerobically at 37°C for 24 hours. After incubation, typical black centered creamish colonies suggestive of *C. perfringens* [23] were stained with Gram's stain and sub-cultured on the SPS agar plates until they were free from other contaminating bacteria. The pure cultures of suspected *C. perfringens* isolates were subjected to Gram's staining and then PCR for final identification.

An isolated colony was stained on a grease free clean glass slide using Gram staining kit (Becton, Dickinson and Company, Sparks, MD, USA). The violet coloured bacteria indicated the Gram positive bacteria while pink coloured indicated the Gram negative bacteria.

DNA extraction was done by boiling and snap chilling method [4]. Two to three suspected colonies from SPS agar plates were suspended in 1.5 ml microcentrifuge tubes containing 100 µl of distilled water by gentle vortexing. The samples were boiled for 10 minutes, cooled on ice for 5 minutes and centrifuged at 13,000 rpm/ 10,000xg for 10 minutes in a table-top microcentrifuge (Cooling Centrifuge, Eppendorf 5418R, Hamburg, Germany). 3.0 µl of the supernatant was used as the template for PCR.

Isolates of *C. perfringens* from water samples were confirmed by amplifying 16S rRNA gene of the *C. perfringens* as per Tanooka, et al. (2005) [22]. The 25 µl reaction consisted of 3.0 µl template DNA, 2.5 µl of 10X buffer, 2.0 µl of MgCl₂, 0.3 µl of 25mM dNTP mix, 0.5 µl each of forward and reverse primers, 1 U of Taq DNA Polymerase (Fermentas Life Sciences, USA) and rest of sterile distilled water. Well characterized isolates of *C. perfringens* maintained in the Division of Veterinary Microbiology and Immunology, FVSc

and AH, Shuhama, Ganderbal were used as positive control while sterile distilled water served as negative control. The primers used were procured from GCC Biotech, Kolkata, India and have the following sequence: forward- TAACCTGCCTCATAGAGT and reverse- TTTACATCCCACTTAATC. The PCR conditions in the thermal cycler (Mastercycler Gradient, Eppendorf AG, Germany) consisted of initial denaturation at 95°C for 5 minutes, followed by 35 cycles of denaturation at 94°C for 30 seconds, annealing at 49°C for 90 seconds and extension at 72°C for 90 seconds, followed by final extension at 72°C for 10 minutes.

Bacterial load (CFU/ml) of *C. perfringens* in water samples

For assessing the clostridial load of the Dal Lake, Serial dilution spread plate technique was used which is as follows:

1. Test tubes were arranged in the test tube stand and were labelled in a serial manner. All the tubes were filled with 9 ml autoclaved brain heart infusion broth (BHI, HiVeg™; Hi-media Laboratories, Mumbai, India).
2. 1 ml of the water sample was added into the first tube filled with 9 ml of BHI broth. This gives 1:10 or 10^{-1} dilution.
3. Then, 1 ml of the sample suspension was taken from the first tube and added into second tube containing 9 ml of BHI broth. This gives 1:100 or 10^{-2} dilution.
4. In this manner, 10^{-3} , 10^{-4} and 10^{-5} dilutions were prepared.
5. One ml of first dilution was taken and inoculated on a sterile Sulfite Polymyxin Sulphadiazine agar plates (SPS HiVeg™ Agar, Modified; Hi-media Laboratories, Mumbai, India) using L-spreader.
6. This was repeated with all dilutions individually and separately.
7. All the plates were then incubated anaerobically in 3.5 litre anaerobic jars (Oxoid Limited, Thermo Fisher Scientific Inc., UK) with GasPak™ Anaerobe Container System (Becton, Dickinson and Company, USA) at 37°C for 24 hours.
8. Plates containing overlapping or diffused colonies were discarded (usually less than 30 or more than 300 colonies).
9. Plates showing distinct and well separated colonies were selected.

After incubation, the visibly distinguishable *C. perfringens* colonies were identified and the number on each plate was counted using digital colony counter pen (Hi-media, Mumbai, India).

Following formula was used to calculate Colony Forming Unit (CFU):

$$\text{CFU} = \frac{\text{Number of colonies counted} \times \text{Dilution factor}}{\text{Volume of water sample taken (ml)}}$$

Results

The water samples after 48 hours of anaerobic incubation in cooked meat medium at 37°C, showed turbidity with gas bubbling. On Sulphite Polymyxin Sulphadiazine (SPS) agar, *C. perfringens* colonies were typically creamish with black center. Isolated colonies were further subcultured on the SPS agar plates to obtain pure culture of *C. perfringens* (Figure 1).

Figure 1: Cream coloured colonies of *Clostridium perfringens* on Sulphite Polymyxin Sulphadiazine (SPS) agar.

On Gram's staining, the black centered creamish colonies isolated on SPS agar revealed typical Gram-positive stout rods suggestive of *C. perfringens* at 1000X (Figure 2) using compound microscope (Olympus CX21, Olympus Corporation, Tokyo, Japan) with digital camera (10.0 MP Olympus E-520, Olympus Imaging Corporation, China).

All the 45 (100%) isolates from water samples were confirmed positive for *C. perfringens* when subjected to polymerase chain reaction (PCR) using species specific 16S rRNA gene amplification primers. These isolates which were morphologically identified as *C. perfringens*, amplified 481 bp product corresponding to *C. perfringens* (Figure 3).

Figure 2: Gram stained smear of pure culture of *Clostridium perfringens* showing Gram positive violet rods under oil immersion (1000X).

Figure 4: Colonies of *Clostridium perfringens* on SPS agar plate (Spread plate technique).

Figure 3: Detection of 16S rRNA gene of *Clostridium perfringens*
Lane M: 100bp DNA ladder
Lane 1: Positive control
Lane 2-6: Test sample of *C. perfringens* from water
Lane 7: Negative control.

Figure 5: Boxplot showing average CFU/ml values of *Clostridium perfringens* at 5 sites of the Dal Lake.

The clostridial load in the water samples was assessed in terms of colony forming units (CFU/ml) (Figure 4). The overall *C. perfringens* load in Dal Lake at five different sites ranged between 9.2×10 CFU/ml to 1.87×10^2 CFU/ml. The highest average CFU/ml value was found at Dalgate (1.87×10^2), followed by Telbal Nallah (1.78×10^2), Dhobi Ghat (1.20×10^2), Centaur Lake View (1.05×10^2) and Char-Chinari (9.2×10), respectively (Figure 5). Statistically, it was observed that there is a significant difference in average CFU/ml values of *C. perfringens* at the five sites of the Dal Lake ($P < 0.01$) as shown in the table 1.

Site	CFU/ml (Mean value)	Standard Error	F value	P value
Dhobi Ghat	1.20×10^2	3.34	104.6	<0.01
Telbal Nallah	1.78×10^2	4.24		
Char-Chinari	9.2×10	4.85		
Centaur Lake View	1.05×10^2	4.41		
Dalgate	1.87×10^2	3.68		

Table 1: Distribution of average CFU/ml values of *Clostridium perfringens* at five sites of the Dal Lake.

Discussion

Water acts as a mode of transmission for *C. perfringens* to humans and animals which is then responsible for causing varied diseases like gas gangrene and food poisoning in humans, enterotoxaemia, hemorrhagic enteritis, necrotic enteritis and diarrhea in livestock animals and poultry [28]. In the present study, water samples from different sites of the Dal Lake were screened for the presence of *C. perfringens*.

The colonies formed by *C. perfringens* on SPS agar were typical round, umbonate, centrally dense with rhizoid periphery and slightly opaque. These findings are in agreement with those of Das and Jain (2012) and Wani, *et al.* (2018) [5,23]. The Gram's staining of the bacterial smears taken from the pure cultures of *C. perfringens* in the current study revealed typical Gram-positive short rods with blunt ends which correlated well with the reports of Das and Jain (2012), El-Shorbagy (2012) and Wani, *et al.* (2018) [5,6,23]. Based on PCR amplification of 16S rRNA gene, our study revealed that 45 (100%) of 45 water samples were positive for *C. perfringens*.

According to Rios, *et al.* (2017), *C. perfringens* is believed to be an important indicator of faecal contamination in aquatic habitats [18]. For determining the bacterial load of *C. perfringens* in the Dal Lake, a total of 45 water samples from 5 different sites viz., Dhobi Ghat, Telbal Nallah, Char-Chinari, Centaur Lake View and Dalgate (9 samples from each site) were processed. The highest average CFU/ml value was found at Dalgate (1.87×10^2) while as the lowest being observed at Char-Chinari (9.2×10). The possible reason being, that the Dalgate site is more prone to direct impact from human interventions from within (house boats, vegetable gardens) and outside activities (households, hotels, restaurants, shops) and also from point and non point sources of sewage (domestic, agricultural as well as commercial). Moreover, greater tourist influx resulting in the increased movement of house boats in the Dal Lake esp., at Dalgate, where there is no proper disposal system for the night soil results in faecal pollution and deterioration of the water quality coupled with outburst of *C. perfringens*.

C. perfringens load in Dal Lake at different sites was found to be in the range of 9.2×10 CFU/ml to 1.87×10^2 CFU/ml, which is much less than the critical CFU level of *C. perfringens* (10^5 - 10^8) to cause any toxoinfection. However, it must be noted that the water source used for drinking or cleaning purpose should not contain any organism of faecal origin [7,24]. Saleem, *et al.* (2013) concluded that water of the Dal Lake is unfit for drinking purpose as per the standards of WHO after assessing the water quality by studying the coliform bacterial diversity of the lake [19].

The colonization of *C. perfringens* in aquatic habitats is mainly due to indiscriminate dumping of untreated commercial, agricultural or household wastes including sewage [11]. Colonization may also arise from naturally occurring small numbers of *C. perfringens* in aquatic environments [1]. These may have different origins in the study area, such as a result of animal wastes carried by run-off from catchment areas, deposition of fecal material by local as well as migratory birds or atmospheric transport of clostridial spores.

Conclusion

C. perfringens is present in the aquatic environment of the Dal Lake. *C. perfringens* load in Dal Lake was found to be much less than the critical CFU level of *C. perfringens* (10^5 - 10^8) to cause any toxoinfection. However, it must be noted the increasing anthropogenic activities in and around the Lake will eventually result in the elevated levels of this potent human and animal pathogen in its ecosystem and endanger the public health if its water is used for drinking, cooking or sanitary purposes. Thus, in light of the present study, there is a need to conduct an elaborate research on this organism in the various aquatic ecosystems of the valley in the near future in order to minimise its transmission to humans and livestock.

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Conflict of Interest

None.

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