

31. PALYNOLOGY AND DINOCYST BIOSTRATIGRAPHY OF THE LATE MIOCENE TO PLEISTOCENE, NORWEGIAN SEA: ODP LEG 104, SITES 642 TO 644^{1,2}

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ABSTRACT

Cores from ODP Leg 104 Holes 642A, 642B, 642C, 644A, and 644B were sampled at intervals of 40 cm to 100 cm for a detailed study of palynomorphs and palynodebris types in upper Cenozoic sediments of the Vøring Plateau. Core-catcher samples were also studied from Hole 643A on the seaward flank of the plateau. Most of the 600 samples studied contained dinoflagellate cysts, pollen, spores, and various types of palynodebris. Total numbers of indigenous dinocysts, and pollen-spore concentrations show cyclical variations which appear to correspond to climate fluctuations in Pliocene and younger sediments, and to either climatic changes or phytoplankton productivity cycles in the older sediments. Stratigraphic ranges for 68 cyst morphotypes were used to erect a provisional dinocyst zonation that can be correlated with other Northern Hemisphere high-latitude zonations. Four dinocyst zones are defined, with boundaries corresponding to biochronological or magnetostratigraphic ages of ca. 15 Ma, 9 Ma, 4.2 Ma, and 1.4 Ma. Environmental changes are interpreted in terms of (a) a progressive decline in species diversity due to the disappearance of subtropical species; (b) appearance of subarctic North Pacific taxa, presumably from the Arctic Ocean; and (c) an increase in heterotrophic protoperidinioid cyst species during the Pleistocene.

INTRODUCTION

On Leg 104 of the Ocean Drilling Program, Cenozoic sediments were drilled at three sites on a transect across the Vøring Plateau in the eastern Norwegian–Greenland Sea (Fig. 1). The Vøring Plateau is a marginal plateau that extends westward from the Norwegian Shelf at a water depth of ca. 1400 m (Fig. 2), then descends to the Lofoten Basin and the Norway Basin on the north and south sides, respectively. At present, the Norwegian Current transports warm, saline North Atlantic Drift water northward over the Iceland–Faroe Ridge, and the East Greenland Current transports cold water from the Arctic Ocean southward to the North Atlantic through the Denmark Strait (Fig. 1). The Vøring Plateau is thus well located for detailed study of the paleoclimatic and oceanographic evolution of the high-latitude eastern North Atlantic region, including its responses to Cenozoic global climatic changes and the tectonic history of the sills and channels which link the Norwegian–Greenland Sea with the Atlantic and Arctic Oceans. This paper reports the results of high-resolution studies of palynomorphs and palynodebris in the upper Cenozoic sediments, and it presents a detailed dinocyst biostratigraphy which clarifies and refines the initial shipboard results reported by Mudie in Eldholm, Thiede, Taylor, et al. (1987).

On Leg 104, six holes were continuously cored using the advanced hydraulic piston corer to obtain detailed records of the sedimentological and paleoceanographic histories of the sites shown in Figure 2.

The primary purposes of this report are as follows:

1. To compare the palynological assemblages from the near-shore to offshore parts of the Vøring Plateau transect;
2. To establish a regional dinocyst biostratigraphy for the upper Miocene through Pleistocene sediments and its correlation

with other North Atlantic and high-latitude palynostratigraphies;

3. To interpret the palynostratigraphic data in terms of paleoclimatic changes which occurred during the late Cenozoic.

During Leg 104, shipboard palynological studies (Eldholm, Thiede, Taylor, et al., 1987) were made from about 250 samples in an attempt to erect a Cenozoic dinocyst biozonation by correlation with data from DSDP Leg 38 in the Norwegian Sea (Manum, 1976) and with zonations described for deep-sea areas south of the Iceland–Faroe Ridge, e.g., DSDP Leg 94, Site 611 (Mudie, 1986), DSDP Leg 81 (Edwards, 1984), and DSDP Leg 48 (Costa and Downie, 1979; Harland, 1979). Results of the initial Leg 104 shipboard studies were very tentative, however, because of the following limitations:

1. Onboard laboratory processing facilities precluded routine use of HF digestion so that palynomorph concentrations (number per gram) were often low, and detailed cyst morphology was difficult to discern;

2. Large-scale variations in palynomorph concentration and composition characterize most high-latitude upper Cenozoic deep-sea sediments (Mudie, 1986); shipboard examination of core-catcher samples alone therefore provided a very limited perspective of the total palynoflora;

3. Many Norwegian Sea dinocyst species have not yet been described (Manum, 1976; Manum and Boulter, this volume), and there are still many uncertainties regarding equivalence of North Atlantic Neogene taxa reported by different workers (Head et al., in press a, b; de Vernal and Mudie in press a, b).

For the postcruise studies reported in this paper, 500 core section samples and 50 new catcher samples were processed using the same laboratory method as Manum and Boulter (this volume), who have made detailed studies of the early Cenozoic palynology at Sites 643 and 642. Use of the same processing methods allows close comparison of morphotypes from all ODP Leg 104 palynological samples. Many dinocyst species identifications have been cross-checked with Manum and other workers studying Neogene dinocysts, but there is still need for sample exchange, comparative study, and stabilization of nomenclature. The late Cenozoic dinocyst zonation presented in this paper

¹ Eldholm, O., Thiede, J., Taylor, E., et al., 1989. *Proc. ODP, Sci. Results, 104*: College Station, TX (Ocean Drilling Program).

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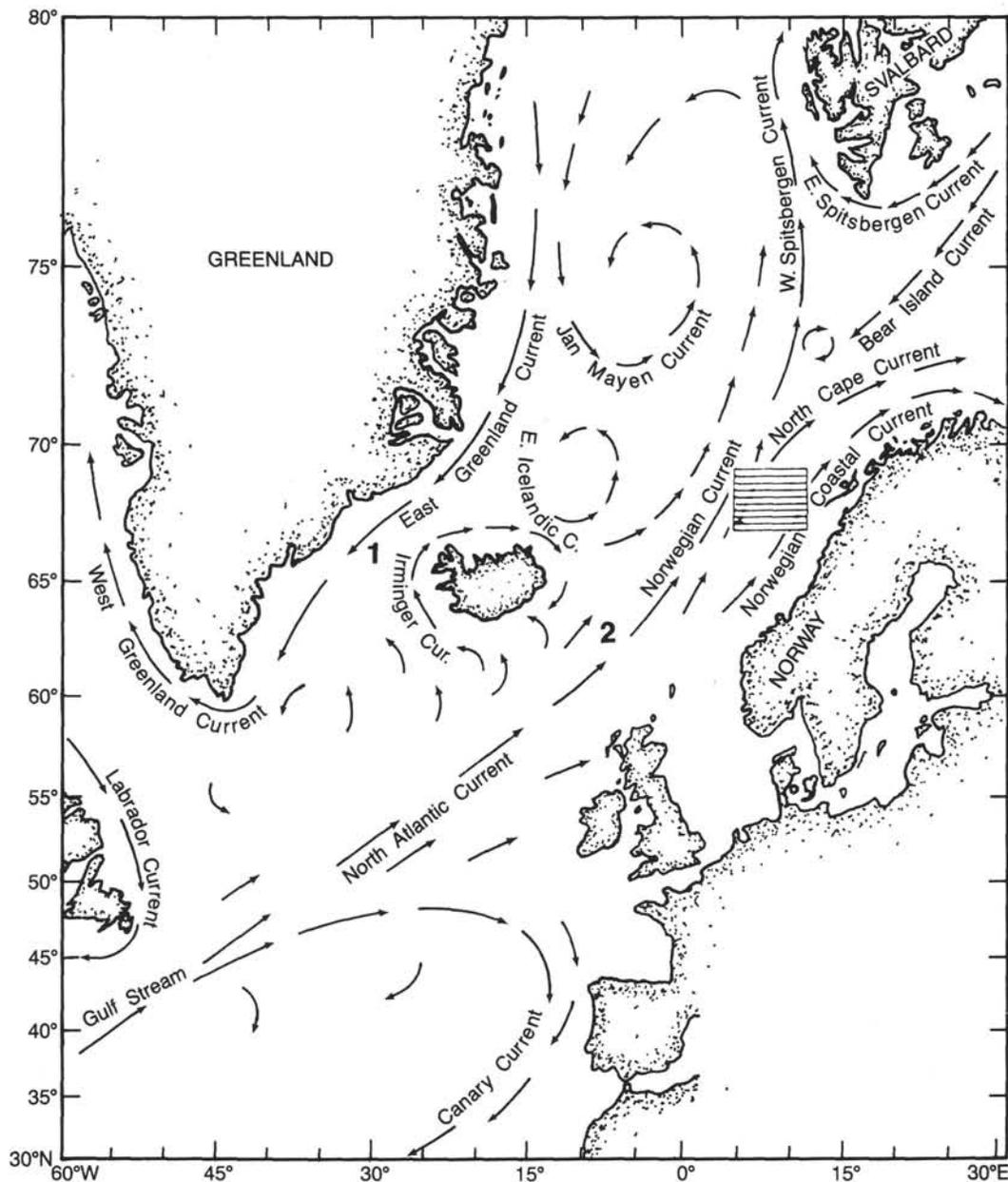


Figure 1. Location of the Vøring Plateau area (shaded square) in the Norwegian-Greenland Sea and the modern surface current circulation. 1 = Denmark Strait; 2 = Iceland-Faeroe Ridge.

may therefore be refined on completion of these taxonomic studies in 1988.

METHODS

Sediment samples (8-cm³ volume) representing 2-cm-long core segments were taken at intervals of 40 to 100 cm for cores from Sites 642 and 644, but only catcher samples were available for Site 643. Samples were disaggregated in 1% Calgon solution and sieved through screens with mesh openings of 150 and 10 μ m. The silt-fine sand fraction was sequentially processed with hot HCl and HF to remove carbonates and silicates, respectively, and one *Lycopodium* tablet was added to the residue to provide a basis for estimating palynomorph concentration (for details, see Mudie, 1986). Residues were stained with Safranin-O and strew mounts were made for palynodebris and biostratigraphic studies. Many preparations, however, contained too much amorphous organic debris to allow detailed study of dinocyst morphology. For these samples, aliquots of the residues were processed further using brief oxida-

tion with NaClO and HCl, following Evitt (1984). This oxidation method is the best one for removing fine detritus that obscures palynomorphs in many samples of high-latitude marine sediments (Manum, 1976; Mudie, 1986), but it severely damages thin-walled protoperidinioid dinocysts and it cannot be used for routine palynological processing.

Slides were first examined with a Zeiss Universal microscope and Orthoplan ($\times 16$) or Normarski interference contrast objectives ($\times 40$ and $\times 100$). Each slide was scanned at $\times 160$ magnification, Polaroid photographs were taken to record palynodebris amount and type, and all known taxa were recorded. Taxonomic verifications were then made at magnifications of $\times 400$ and $\times 1000$, and counts were made for a total of up to 200 dinocysts. Reworked Paleogene and older palynomorphs were recognized on the basis of their morphology. Reworked Neogene palynomorphs were mostly recognized by their yellow color (i.e., absence of staining with Safranin-O), or by their flat shape and presence of mineral imprints. Some questionable specimens were also examined by fluorescence microscopy and were counted as reworked if their fluorescence emission was lower than the average for *in-situ* palynomorphs of similar morphology.

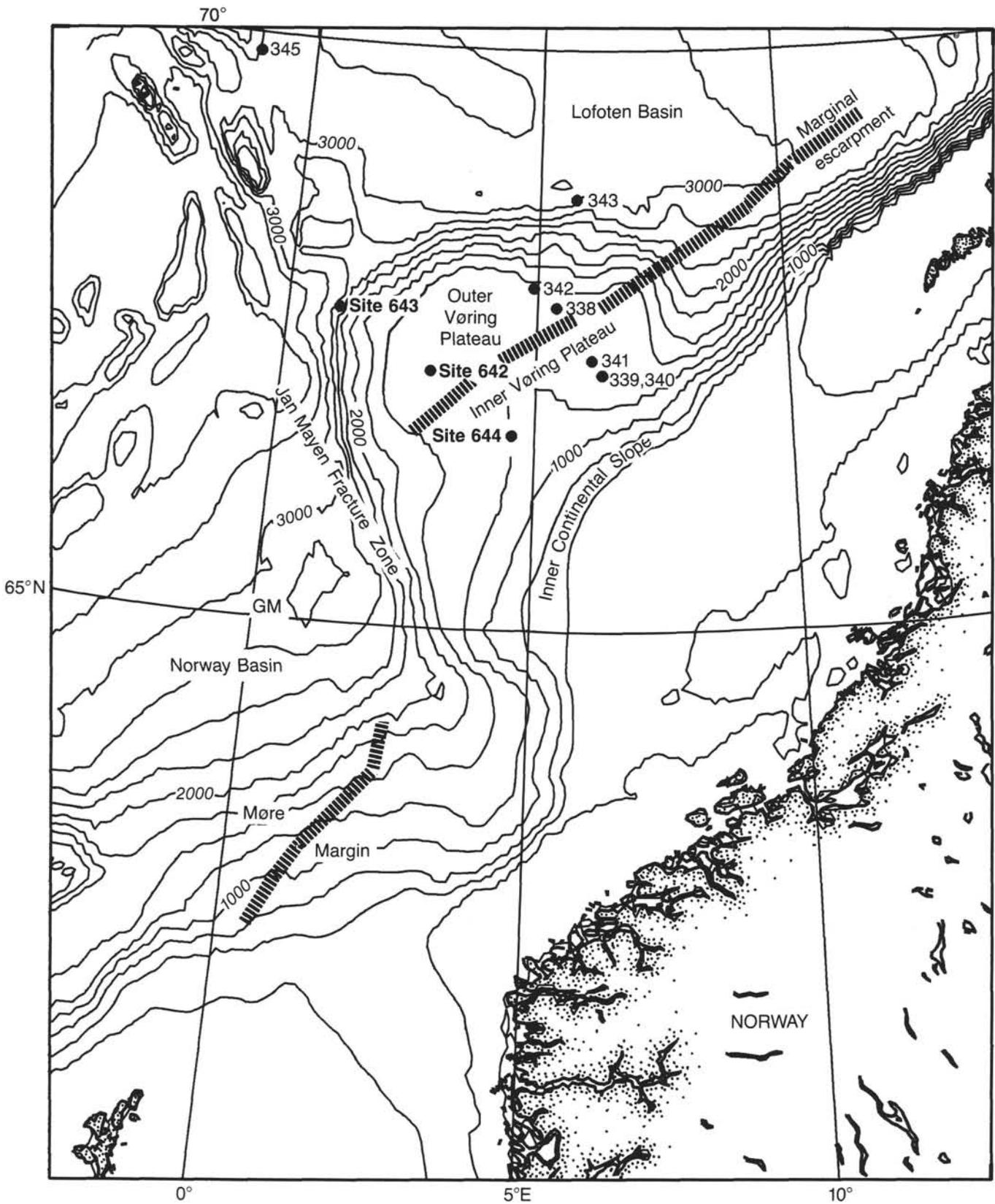


Figure 2. Bathymetric map of the Vøring Plateau area, showing locations of ODP Sites 642, 643, and 644. Contour interval 250 m.

PALYNODEBRIS TYPES AND QUANTITY

About 600 samples from the Vøring Plateau sites were processed for palynological study of the upper Miocene to Pleistocene sediments. All except 20 of these contained dinocysts, pollen, spores, and various types of other acid-resistant plant material. The latter class of organic residues is referred to here as palynodebris. The term palynodebris therefore mainly refers to visual sedimentary organic matter as defined by Boulter and Riddick (1986) but it does not include the microplankton component (dinocysts, acritarchs), pollen, terrestrial plant and algal spores, foraminiferal linings, which are here referred to as palynomorphs. The main palynodebris types in the upper Cenozoic sediments include:

1. Amorphous matter—pale yellow or brown, partially translucent organic matter, often associated with black specks of fine pyrite or other material.
2. Wood fragments—well preserved wood cells with discernible wall structures, e.g., bordered pits, annular or spiral thickenings.
3. Leaf cuticle—leaf epidermis with well-defined cell structures.
4. Degraded debris—blocky, reddish particles without clearly discernible tissue structure.
5. Black debris—opaque, angular fragments with no discernible tissue structure.

The range of palynomorphs and palynodebris types found in the Vøring Plateau drill holes is shown in Plate 1. Graphs of concentrations (no. per cm³ sediment) of indigenous dinocysts and indigenous pollen and spores are shown in Figures 3 and 4; relative abundances of microphytoplankton are shown beside and on the dinocyst range charts (Figs. 5–10). Foraminiferal linings are present in almost all samples, with no clearly discernible stratigraphic distribution pattern; therefore they are not discussed further in this study.

Following the procedure of Manum (1976) for DSDP Leg 38, the palynomorph and palynodebris data can be synthesized to define general characteristics of the visual sedimentary matter in the Leg 104 drill holes, and to discern palynoevents, which are major changes in the ratio of marine dinocysts to terrestrial pollen and spores (Manum, 1976). The high-resolution sampling of Leg 104 cores reveals cyclical fluctuations in indigenous dinocyst and pollen and spore concentrations throughout the stratigraphic column at all three drill sites (Figs. 3 and 4). This cyclicity is superimposed on the palynoevent curves described by Mudie in the Leg 104 shipboard report. The new high-resolution data cast doubt on the validity of palynoevent curves as an index of regional paleoenvironmental changes when low-frequency sampling is used and when redeposited palynomorphs are grouped together with indigenous palynomorphs to calculate cyst ratios (Manum, 1976). The cyclical variations in indigenous palynomorphs, however, correspond to changes in palynodebris type and allow a tentative delimitation of palynofacies sequences (see Plate 1). Final interpretation of these palynofacies requires more closely spaced sampling of, and correlation with, stable isotopic data (Jansen et al., this volume), but an initial interpretation is presented here.

1. Upper Pliocene-Pleistocene sediments show intervals with common to abundant dinocysts (>1000/cm³) and 10% to 80% amorphous palynodebris (Plate 1, Figs. 1–7) alternating with intervals containing fewer dinocysts (<100–500/cm³), <10% or no amorphous debris, and abundant black debris, wood, and/or degraded debris. All of these samples also contain re-

worked Cretaceous-Paleogene palynomorphs, with ratios of reworked/indigenous specimens being >1. The intervals with relatively high numbers of dinocysts correspond to all or part of interglacial stages and can be referred to as interglacial palynofacies. These frequently include coenobia of the freshwater algae *Pediastrum* and/or *Botryococcus* which are found in the vicinity of large rivers in modern arctic environments (Hill et al., 1985). The intervals with few dinocysts and abundant large particles of black debris or wood correspond to glacial stages or to intervals of ice rafted debris (IRD) deposition during interglacial stages.

2. There is a notable increase in amplitude of the interglacial palynomorph influxes between isotopic stages 9 and 11, from ca. 347–440 Ka (Fig. 3, 35 to 43 mbsf in Hole 644B; 18 to 21 mbsf in Hole 642B); this increase is also associated with a change in the amount and nature of amorphous debris which increase in volume, become darker in color, and include abundant black specks. Freshwater algae also become less common. In the lower Pleistocene interval at Site 644, diatom fragments and/or black spores (pyritized diatoms; see Mudie, 1986) become common to abundant, but this change is not seen above the upper Pliocene interval at the more seaward location, Site 642.

3. Large fluctuations in the amplitude of palynomorph frequencies appear throughout the lower Pliocene and Miocene intervals at Sites 642 and 643, with the Miocene interval being distinguished by smaller declines in dinocyst concentrations. These troughs are also associated with reduced amounts of amorphous debris (<50%) and, at Site 642 (Figs. 4 and 7), the troughs are correlated with increased abundances of diatoms, possibly indicating that dinoflagellate productivity was lower during intervals of frequent diatom blooms. Throughout the upper Miocene-lower Pliocene interval, particles of wood and black and unstructured debris particles are relatively small, and reworked Cretaceous-Paleogene palynomorphs are rare (ratio of reworked: indigenous specimens range from 0.01 to <1.0).

4. Middle to lower Miocene sediments at Site 643 (below ca. 119 mbsf, Fig. 3) appear to contain consistently high concentrations of indigenous dinocysts. This signal may indicate more uniformly favorable environmental conditions, but the sampling frequency (ca. one sample per m.y.) is too low to dismiss all possibility of large-amplitude cyclicity. The most notable feature of all samples from this interval is the large quantity of dinocysts and sparseness of palynodebris, pollen, spores, and diatoms.

5. There is an apparently slight decrease in palynomorph concentrations for equivalent time intervals from the inner to outer sites (Fig. 3). Sedimentation rates at the inner Site 644, however, are two to five times higher than at the middle Site 642; thus, the number of palynomorphs deposited per unit time is actually two to five times higher at Site 644 than appears on the graph. In contrast, the sedimentation rate at Site 642 is about 1 cm/ka or less below the Jaramillo event (ca. 47 and 110 mbsf in Holes 642B and 644B, respectively). The number of palynomorphs deposited per unit time at Site 642 is therefore the same or less than the numbers shown on the graphs. This trend of decreased palynomorph abundance across the Vøring Plateau conflicts with the Leg 104 organic carbon data (Eldholm, Thiede, Taylor, et al., 1987) which show amounts of <1% for upper Cenozoic sediments at Sites 644 and 643 and amounts of 0.4 to 1.8% for Site 642. These conflicting data show that, in this area of complex currents, palynomorph content may be a poor index of total organic accumulation because strong bottom currents at the inner site (Site 644) and outer site (Site 643) reduce the deposition rates of fine-grained particulate organic matter. The nearshore-offshore trend from higher to lower palynomorph and palynodebris concentrations, however, is consistent with previous

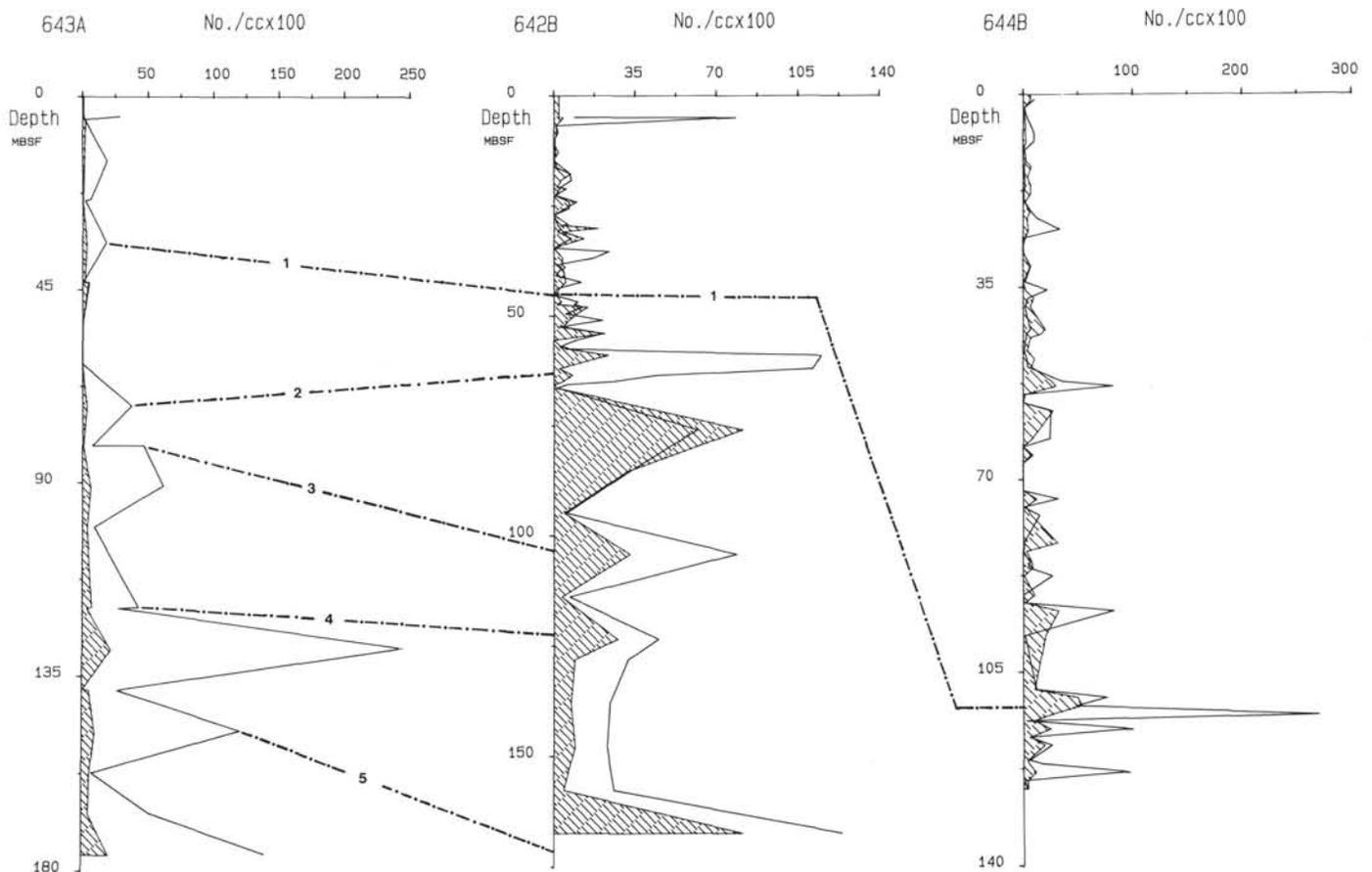


Figure 3. Graphs of fluctuations in concentrations of indigenous dinocysts (unshaded areas) and of pollen and spores (hatched areas) for upper Cenozoic sediments from the outer (Site 643) to inner part (Site 644) of the Vøring Plateau. Dotted lines indicate chronostratigraphic correlations: (1) base of Jaramillo event; (2) Gauss/Gilbert boundary; (3) chron 8 base; (4) middle/late Miocene boundary; (5) early/middle Miocene boundary. Vertical axis shows depth of meters below seafloor (mbsf). Horizontal axis shows 1/100th of the number of palynomorphs per cm^3 sediment.

observations that maximum dinocyst productivity occurs in neritic environments (Wall et al., 1977) and that pollen-spore deposition declines exponentially with distance offshore (Mudie, 1982).

DINOCYST STRATIGRAPHY

Taxonomy and Zonation

More than 100 late Cenozoic dinoflagellate and acritarch morphotypes were observed in the Leg 104 samples, of which about 50 may be new species. Many of these morphotypes require detailed study and comparison with similar forms illustrated by Manum (1976) and other workers (e.g., Powell, 1986 a, b, c; Wrenn and Kokinos, 1986; Matsuoka, 1983; Bujak and Matsuoka, 1986a). In this report, therefore, informal taxonomy is frequently used, and descriptions are confined to notes which explain how certain taxonomic problems have been treated in this report. Some of the new taxa which also occur in the Labrador Sea and/or Baffin Bay are described elsewhere (Head et al., in press a,b; de Vernal and Mudie, in press a, b). Not all bibliographic references are included here for taxa described before 1985, but throughout the report, the nomenclature of Lentin and Williams (1985) is followed. That publication provides the necessary bibliography. The Appendix lists all the names and authors of the dinocysts and acritarchs referred to in this paper.

Wherever possible, the proposed zonation emphasizes previously well-documented species, with the zones being based on concurrent ranges of these index species. The proposed zonation

may require some revision on completion of detailed taxonomic studies and chronostratigraphic correlation of the Leg 104 Sites (Goll, this volume). Similarity between the dinocyst zones described here and in previous studies of other high-latitude regions (Fig. 11) suggests, however, that the provisional biozones are approximately valid.

Dinocyst Distribution: Site 642

The most complete sequence of upper Miocene to Pleistocene sediments (150 m) was recovered from Holes 642B and 642C. Magnetostratigraphic data (Bleil, this volume) and planktonic foraminiferal distributions also provide relatively good time control for these holes (note that the revised time scale of Berggren et al., 1985 is used throughout the Leg 104 reports). Dinocysts were initially studied in 91 samples taken at 1-m intervals in cores from Holes 642A and 642B (Core 104-642A-1H and Cores 104-642B-1H through -8H), and at 20 to 70-cm intervals in Core 104-642C-11 of Hole 642C. Unfortunately, it only later become evident from postcruise biostratigraphic studies that the upper Miocene section lies below Cores 104-642B-9H and -642C-11H in Holes 642B and 642C, respectively. In this study, therefore, only core-catcher samples could be examined from the upper Miocene sediments.

The distributions of the main late Cenozoic taxa are shown in Figure 5, and high-resolution data for the uppermost Miocene and lower Pliocene interval of Hole 642C are shown in Figure 6. These new data, combined with detailed onshore studies

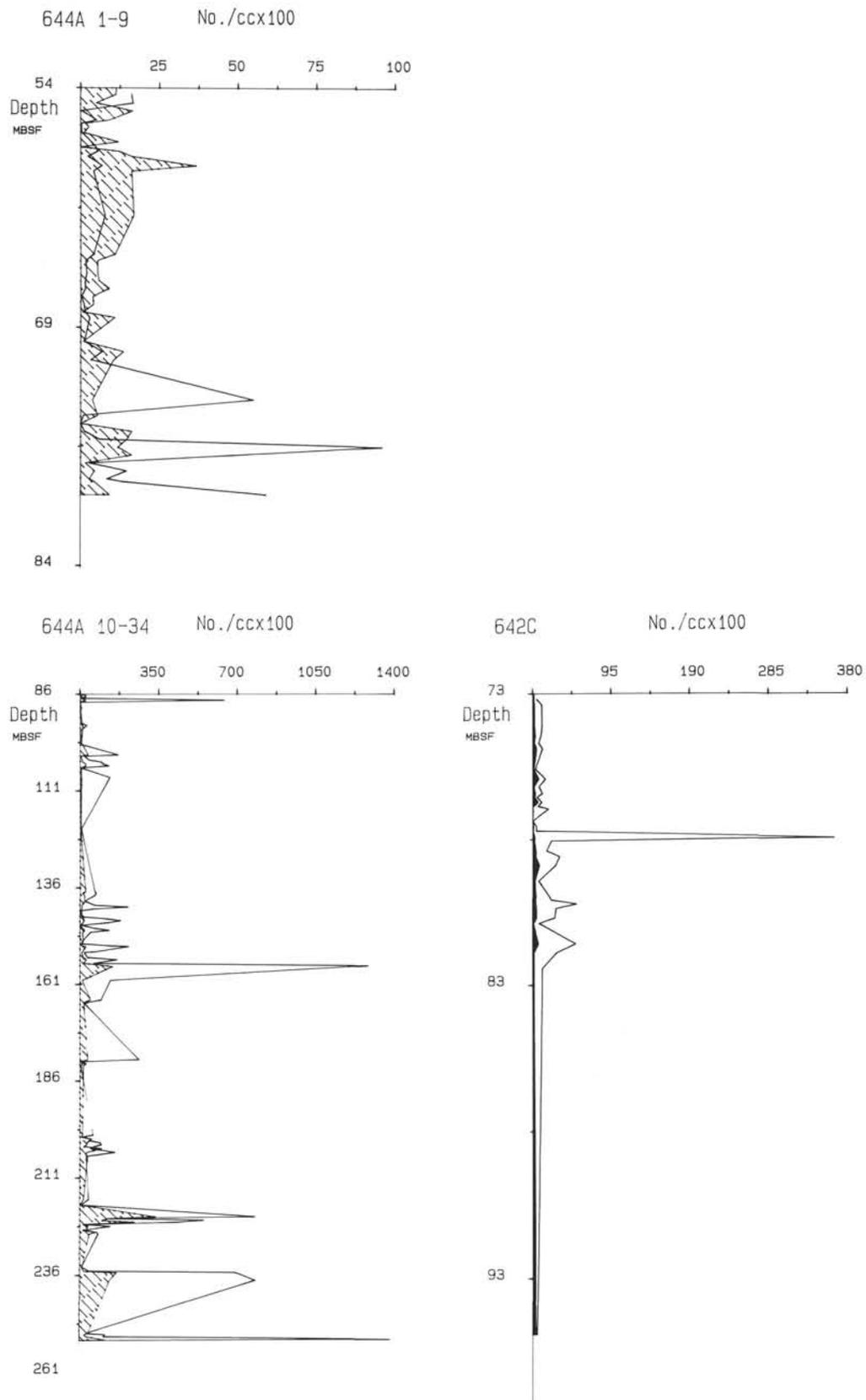


Figure 4. Graphs of fluctuations in concentrations of indigenous dinocyst (unshaded) and pollen and spores (hatched) for very high-resolution sample intervals at Site 644 (upper Pliocene-upper Pleistocene) and for Site 642 (upper Miocene-middle Pliocene). Vertical scale: depth in meters below sea floor (mbsf). Horizontal scale: 1/100th number of palynomorphs per cm³ sediment.

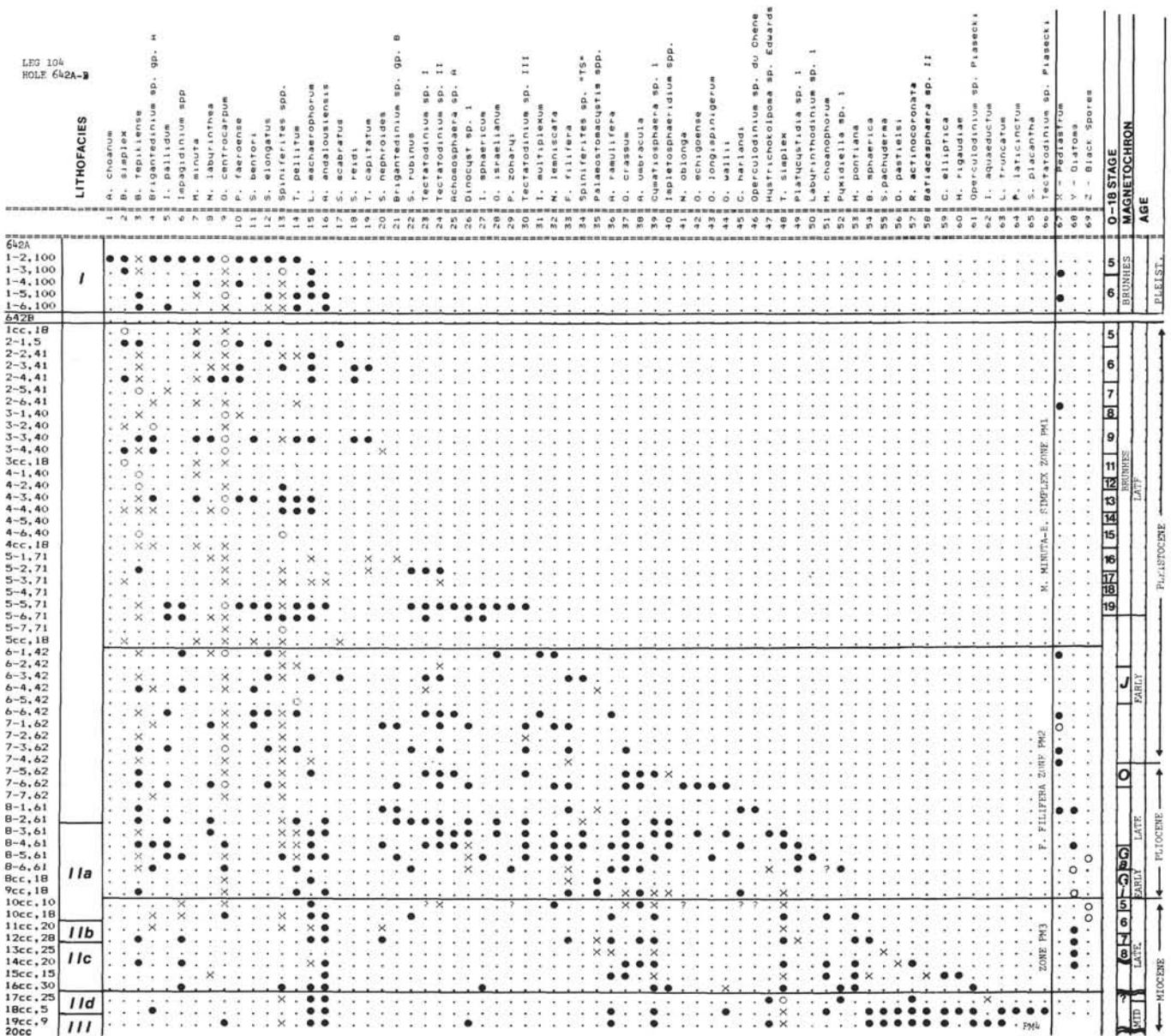


Figure 5. Range chart for selected dinocysts and acritarchs, Holes 642A and 642B. Species are ordered by highest appearance and are listed by number of Appendix; relative abundances are indicated by: O = abundant (>49%); X = common (10-49%); • = rare to frequent (1-9%); . = not present. Lithofacies are I—glacial mud; IA—dark glacial mud interbedded with foraminifer-nannofossil ooze; IC—siliceous and nannofossil mud interbedded with dark glacial sand; IIA—siliceous nannofossil ooze and mud; IIB—siliceous mud or sandy mud and ooze; IIC—interbedded nannofossil/siliceous ooze and mud; IID—siliceous-calcareous ooze, siliceous mud and nannofossil ooze; III—diatom or siliceous mud and ooze. Wavy lines below magnetochron 8 indicate possible stratigraphic hiatuses.

of cyst morphologies, require that several changes to be made for the informal Zones PM1 to PM4 of the shipboard palynostratigraphy for Leg 104 (Eldholm, Thiede, Taylor, et al., 1987).

1. The top of Zone PM4 is now placed between Samples 104-642B-16H,CC, 30-32 cm, and -17H,CC, 20-22 cm, at the last appearance datum (LAD) of *Impagidinium aquaeductum* (= *Nematosphaeropsis aquaeducta*). Younger occurrences of poorly preserved *I. aquaeductum* cysts are now considered to be reworked. The base of Zone PM4 is provisionally placed at Sample 104-642B-19H,CC, 8-10 cm, but it cannot be defined with certainty because of a possible hiatus in Core 104-642B-19H and poor recovery of cysts from Cores 104-642B-20H to -22H.

2. The base of Zone PM3 is now extended to Sample 104-642B-16H,CC, 30-32 cm, which corresponds to the first ap-

pearance datum (FAD) of *Hystrichosphaeropsis ?pontiana* (this taxon was included with *H. obscura* in the shipboard study). The top of Zone PM3 is marked by the last common occurrence of *Amiculosphaera umbracula*. This boundary is placed in Sample 104-642B-9H,CC of Hole 642B and in Sample 104-642C-11H-3, 56-58 cm of Hole 642C. The provisional name of Zone PM3 is now changed to the *Achomosphaera andalouisiensis* acme zone.

3. The base of Zone PM2 is still located at Sample 104-642B-9H,CC, but it is redefined as being marked by the FAD of *Filisphaera filifera* and *Tectatodinium pellitum*. The top of Zone PM2 is now relocated at Sample 104-642B-6H-IG, 42-44 cm, which marks the LAD of *Nematosphaeropsis lemniscata*. Zone PM2 now contains the range of *F. filifera* and is therefore renamed the *F. filifera* zone.

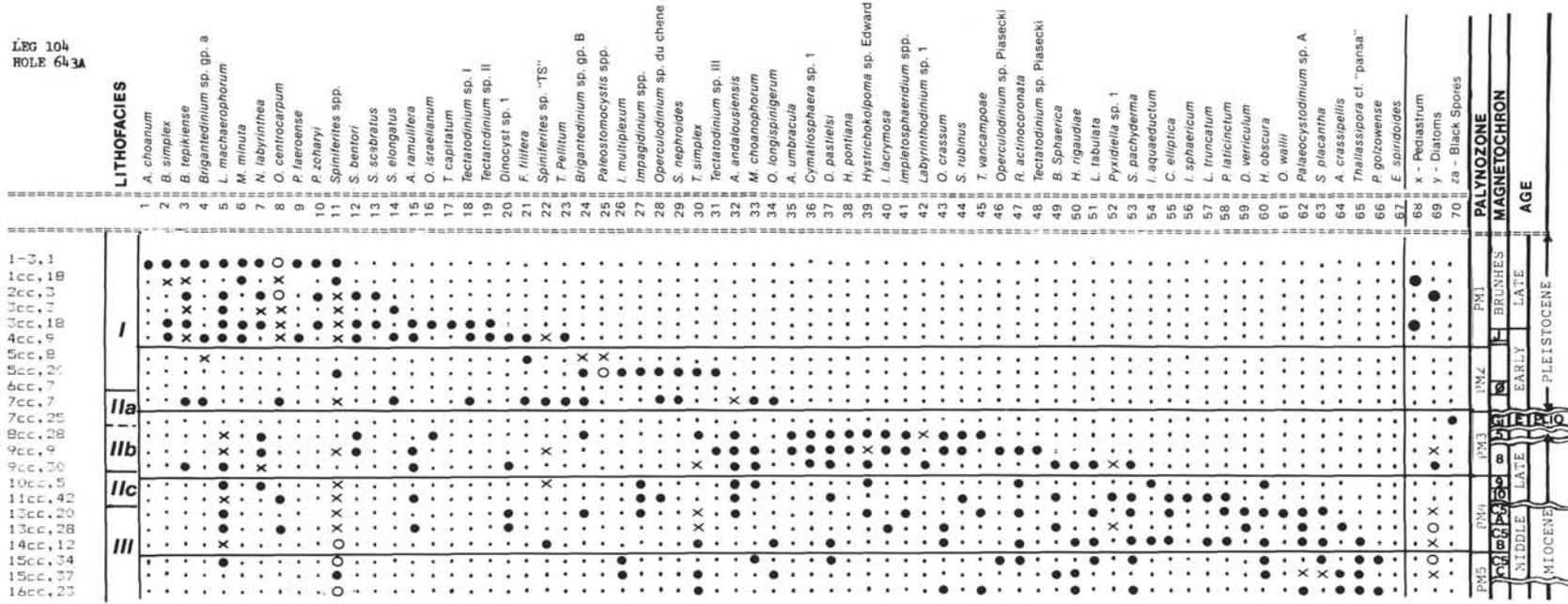


Figure 7. Range chart for selected dinocysts and acritarchs, Hole 643A. Species are ordered by highest appearance and are listed by number in Appendix; relative abundances and lithofacies are as in Figure 5. Wavy lines on right indicate probable stratigraphic hiatuses.

Dinocyst Distribution: Site 644

For this study, 200 samples from core intervals of 40-cm spacing were examined from the base (upper Pliocene) to top (upper Pleistocene) of Hole 644A. Figures 8 and 9 show dinocyst distributions for most of these samples, excluding the top seven cores which are well represented by data for Hole 644B. A total of 65 samples from 100-cm intervals were examined from the Pleistocene sediments cored in Hole 644B (Fig. 10). Both holes recovered continuous sequences of sediments for which biostratigraphic and magnetostratigraphic data apparently provide good age control (Goll, and Bleil, this volume). According to the revised palynozonation described above for Hole 642B, several changes have been made to update the shipboard palynostratigraphy for Site 644.

1. Zone PM3 is no longer recognized in Hole 644A. The revised stratigraphy for Site 642 now clearly shows that the ranges of several common late Miocene species, e.g., *A. umbracula* and *Operculodinium crassum*, continue upwards in Zone PM2 to at least the Pliocene/Pleistocene boundary. Small numbers of other species with more restricted occurrences in Zone PM3 at Site 642, e.g., *Corrudinium harlandi*, *Tectatodinium simplex*, and *Reticulatosphaera actinocoronata*, occur rarely and sporadically in the upper Pliocene interval of Hole 644A, and they are probably reworked. The occurrence of *A. andalousiensis* is also sporadic in Pleistocene sediments of high-latitude North Atlantic regions (Askú et al., in press), whereas coherent assemblages of late Miocene and Pliocene index species were not found at Site 644.

2. The base of Hole 644A contains Zone PM2; the zone top occurs at Sample 104-644A-10H-3, 35–37 cm, which marks the LAD of *N. lemniscata* and the FAD of *M. minuta* and *B. simplex*. Zone PM1 extends from this sample to the top of the hole.

3. In Hole 644B, Zone PM2 extends from the base to Sample 104-644B-10H-5, 86–88 cm, which lies just above the LAD of *N. lemniscata* and contains the FAD of *M. minuta* and *B. simplex*. Zone PM1 extends from this sample to the top of the hole.

Provisional Norwegian Sea Zonation and Correlation

Impagidinium aquaeductum—Zone PM4

This is the oldest Miocene palynozone that can be delimited with reasonable certainty from the samples in this study. As defined here, the base of Zone PM4 is marked by the FAD of *Labyrinthodinium truncatum* and *Cyclopsella elliptica*, and by the LAD of *Palaeocystodinium golzowense* and a new cyst form, *Thalassiphora* sp. cf. *Th. pansa*, which is common together with *Emslandia spiridoides* in the underlying early Miocene Zone PM5 (= *Emslandia spiridoides* Zone). The top of Zone PM4 is defined as the LAD of *I. aquaeductum*. In both Holes 642B and 643A, three well-known middle Miocene and older species have their LAD just below the zone top: *L. truncatum*, *Pentadinitium laticinctum*, and *Systematophora placacantha*. Other characteristic species of Zone PM4 include *Operculodinium* sp. of Piasecki (1980), *O. wallii*, *Hystrichokolpoma rigaudiae*, *Batiacasphaera sphaerica*, and cf. *Batiacasphaera* sp. II of Edwards (1984), *Spiniferites* sp. cf. *S. pachyderma*, *Dapsilidinium pastielsii*, *Reticulatosphaera actinocoronata*, *Tectatodinium simplex*, and *Lingulodinium machaerophorum*. *Spiniferites* species (i.e., various undifferentiated taxa, excluding those listed separately on the range charts) are particularly abundant and diverse in this zone and they will require detailed study to determine their taxonomy and stratigraphic value. In the lower section of Zone PM4 (represented only at Site 643 in this study), cf. *Achomos-*

phaera crassipellis, *Dinopterygium verriculum* (= *Heteraulacysta verricula*) and *Palaeocystodinium* sp. A of Costa and Downie (1979) are common.

The age of the Zone PM4 boundaries are presently not very well constrained due to the paucity of calcareous microfossils and apparent presence of hiatuses between Cores 104-643A-13H and -11H, and two hiatuses between Cores 104-642B-20H and -13H (Bleil, this volume and Goll, this volume). The base of Zone PM4 in Hole 643A, however, lies above the planktonic foraminifer Zone N4-6 (early Miocene, ca. 17.5–23.5 Ma) and occurs just above the base of magnetochron C5B (16.22 Ma) in Hole 642B. The top of Zone PM4, both Holes 643A and 642B, occurs in the middle of foraminifer Zone 16 which gives it an age of about 9 Ma. These constraints indicate a middle to early late Miocene age for the zone. Most of the species listed above for Zone PM4 are common throughout all or part of the middle to lowermost upper Miocene sediments of DSDP Leg 81, Hole 555, on Rockall Bank (Edwards, 1984), the Langhian to Serravallian dinocyst zones LAN-5 to LAN-8 in Italy (Powell, 1986a), and the *L. truncatum* to *D. verriculum* zones of the Gram Section in Denmark (Piasecki, 1980). An age of late Langhian (15.6 Ma) to early Tortonian (9 Ma) for this Norwegian Sea dinocyst zone is therefore broadly consistent with the North Atlantic middle Miocene zonation (see Powell, 1986a). A significant difference, however, is found in the younger age of the LAD for *I. aquaeductum*, which has an early Tortonian LAD at both Sites 642 and 643 on the Vøring Plateau. This discrepancy may be due to the sedimentary hiatuses at these sites, which might facilitate upward mixing of sediment across erosional surfaces. It is notable, however, that the top of Zone PM4 is approximately the same age as that of Dinocyst Zone 1a at DSDP Site 611 (Mudie, 1986) in the subpolar region of the eastern North Atlantic, although the marker species of Zone 1a differ from those in Zone PM4 and do not include *I. aquaeductum* or *L. truncatum* (see Head et al., in press a, b). The similar age of the zone tops suggests that a regional overturn of dinocyst species took place in the high-latitude North Atlantic seas during the early late Miocene.

Achomosphaera andalousiensis—Zone PM3.

This zone is presently best defined by the acme of *A. andalousiensis* and the FAD of *Amiculospaera umbracula* which occurs just above the base of the zone in Holes 643A and 642B. The basal zone boundary is presently not clearly definable because of the lack of high-resolution sampling for these holes; high-resolution data for core 104-642C-11H, however, provides detailed information for the zone top. The base of Zone PM3 contains the FAD of *Hystrichosphaeropsis ?pontiana* which probably has its range within this zone (isolated occurrences above the top of Chron 5 in Hole 642C are probably reworked). *Impagidinium sphaericum* also has its FAD at the zone base. At Site 642, *Operculodinium crassum* has its FAD just above the zone base, *Bitectatodinium tepikiense* has its FAD in the same sample as *A. umbracula*, and *Selenopemphix* sp. B of Powell (1986b) has its FAD in the middle of the zone. Several common middle Miocene taxa have their LAD between the base and middle of the zone, including *Operculodinium* sp. of Piasecki (1980), *H. rigaudiae*, *Cyclopsella elliptica*, *Batiacasphaera* sp. II of Edwards (1984), *B. sphaerica*, and *Dapsilidinium pastielsii*. The top part of Zone PM3 is marked by the FAD of *Nematosphaeropsis lemniscata*, *Spiniferites rubinus*, *Corrudinium harlandi*, *Operculodinium* sp. of Jan du Chêne (1977), *Tectatodinium* sp. I, and *Tectatodinium* sp. II. Other taxa which are common in all or part of Zone PM3 include *Melitasphaeridium choanophorum*, *Tectatodinium simplex*, Dinocyst sp. 1. The cyst forms referred to as *Hystrichokolpoma* sp. 1, *Platycystidia* sp. 1, and *Imple-*

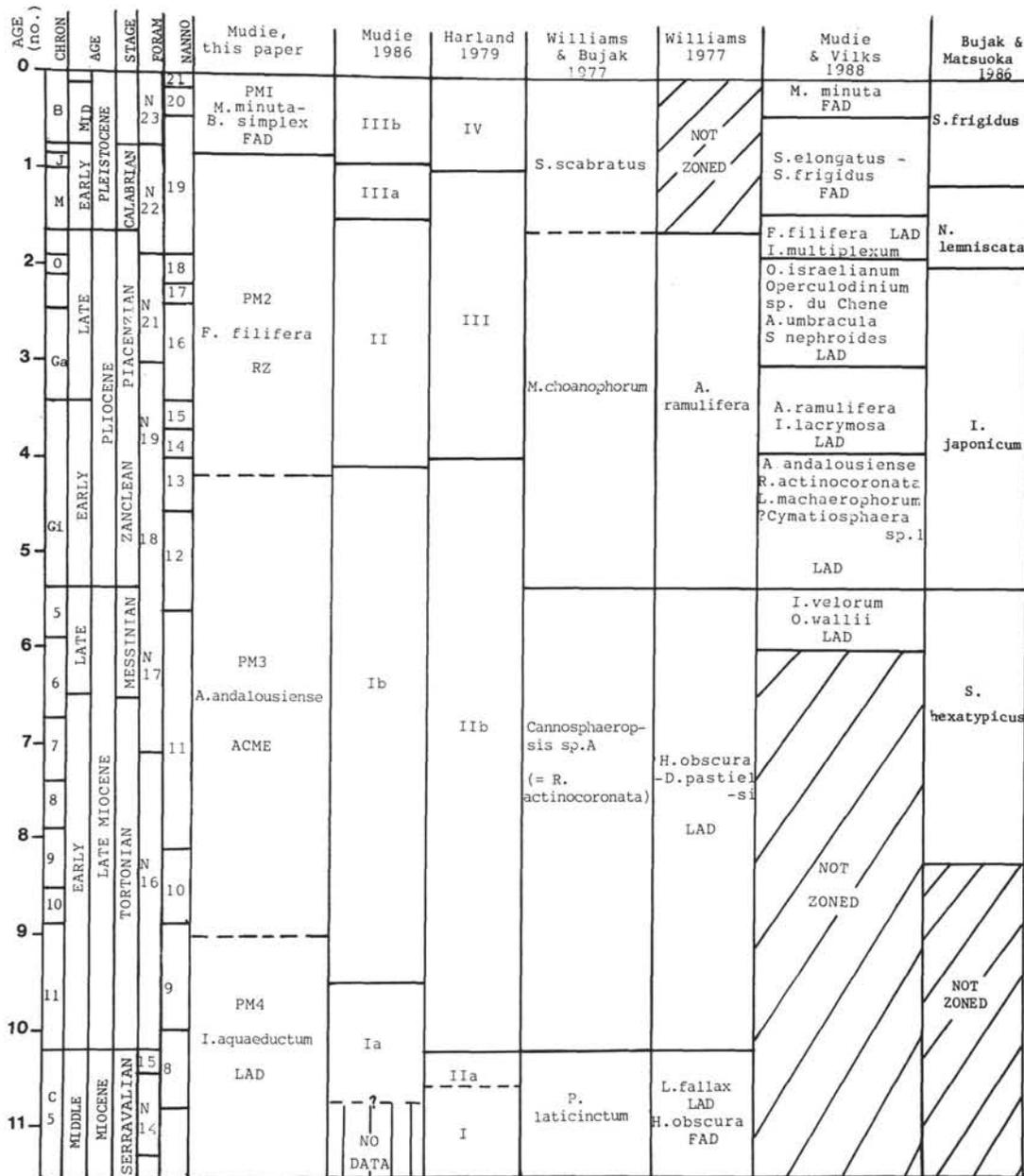


Figure 11. Correlation chart showing the chronostratigraphic relation between the provisional Leg 104 dinocyst zonation and zonations at DSDP Site 611 (Mudie, 1986), Site 400 (Harland, 1979), eastern Canadian offshore wells (Williams and Bujak, 1977; Williams, 1977), northwestern North Atlantic Ocean (Mudie and Vilks, 1988), and the North Pacific Ocean (Bujak and Matsuoka, 1986 a,b). FORAM = foraminifer Zone; NANNO = nannofossil Zone; CHRON indicates magnetostratigraphy, where B = Brunhes, J = Jaramillo, M = Matuyama, O = Olduvai, Ga = Gauss, Gi = Gilbert event.

required to refine the zonation for the upper Miocene interval of the Norwegian Sea.

Filisphaera filifera—Zone PM2.

This zone is presently defined primarily by the range of *F. filifera* and by the partial range of *N. lemniscata*, which has its LAD at the top of the zone. In Holes 642B and 643A the base of the zone is clearly marked by the FAD of *F. filifera* and *Tectatodinium pellitum*; in Hole 642C the FAD of *T. pellitum* occurs about one meter above the FAD of *F. filifera*. Overall, the assemblages in Zone PM2 are dominated by *Operculodinium centrocarpum* (which first becomes abundant just above the base of this zone), *B. tepikiense*, *Tectatodinium* sp. I and sp. II, *Spiniferites ramosus*, *S. membranaceus*, and *S. mirabilis*. *Brigante-*

dinium species group A (undifferentiated, thick-walled, dark brown morphotypes, excluding *B. simplex*) and *Brigantedinium* species group B (thin-walled morphotypes) also become common in this zone. Species diversity, measured as total number of species (S) is relatively high (S = 16–26) in the lower half of the zone below the Olduvai magnetochron (1.88 Ma) but it decreases sharply (S = 6–12) in the upper part. The age of Zone PM2 is well constrained by magnetostratigraphic data which show that the zone extends from the middle of the early Pliocene (ca. 4.2 Ma) to the top of the early Pleistocene, between the Jaramillo and Brunhes magnetochrons (ca. 0.8 Ma).

The base of Zone PM2 includes most of the dinocyst and acritarch species found at the top of Zone PM3, but between the Gauss magnetochron (3.4 Ma) and the top of the Olduvai

event all the small acritarchs disappear, and seven dinocyst species have their LAD in Hole 642B: *M. choanophorum*, *C. harlandi*, *T. simplex*, *Operculodinium* sp. of Jan du Chêne (1977), *O. wallii*, *O. echigoense*, and *Nematosphaeropsis oblonga* (Mudie, 1986). It is also notable that *A. andalusiensis* disappears at, or just above, the Gauss magnetochron at all the Vøring Plateau sites, but it reappears for brief intervals during the Pleistocene at Sites 642 and 644. The upper part of Zone PM2 is marked by the LAD of *O. crassum* and *A. umbracula* just above the Pliocene/Pleistocene boundary. *Spiniferites elongatus* and the cyst-form of *Peridinium faeroense* (Dale, 1977) have their FAD at this boundary in Holes 642B and 644A; *S. scabratus* and *Impagidinium multiplexum* have their FAD at or just below the top of the zone.

In general, the composition and distribution of dinocyst assemblages in Zone PM2 are very similar to those observed in dinocyst Zones II and IIIa at DSDP Site 611, where most of the species listed above show a similar succession and age for the LADs and FADs (Mudie, 1986). The assemblages in Zone PM2 are also comparable to those in Zone III at DSDP Site 400 (Harland, 1979) and there is a notable similarity in the ages of the LADs for *M. choanophorum*, *C. harlandi*, *T. simplex*, *O. crassum*, and *A. umbracula* at all the North Atlantic sites from the Bay of Biscay to Vøring Plateau. The FAD of *S. elongatus* and *I. multiplexum* also have similar ages at Site 611 and the Vøring Plateau, but Harland (1979) reports a younger LAD (middle Pleistocene) for *S. elongatus* at Site 400, while Bujak and Matsuoka (1986a) report a late Miocene FAD for *S. elongatus* in the North Pacific. At present, no subdivision is made for Zone PM2, but future studies may warrant a division at the Pliocene/Pleistocene boundary to delimit an early Pleistocene subzone characterized by the FAD of *S. scabratus* and *I. multiplexum* and by the LAD of *N. lemniscata*. This subzone would then be comparable to the *N. lemniscata* zone in the North Pacific (Bujak and Matsuoka, 1986a).

Multispinula minuta-*Brigantedinium simplex*—Zone PM1.

This zone is delimited by the ranges of *M. minuta* and *B. simplex* which have their FAD at the zone base, at or just above the LAD of *N. lemniscata*. *Peridinium faeroense* cyst-form of Dale (1977) is commonly present throughout this zone. Other characteristic species are *S. elongatus* (here including *S. frigidus* and intergrades), *S. scabratus*, *Impagidinium pallidum*, *T. peltatum*, *Trinovantedinium capitatum*, *Stelladinium reidi*, and *Selenopemphix nephroides*, but their occurrences are often sporadic and their distribution within Zone PM1 varies largely with sampling density. In general, dinocyst assemblages in Zone PM1 are dominated by *O. centrocarpum*, *B. tepikiense*, and *Brigantedinium* spp. At all of the Vøring Plateau sites, however, there is a notable decrease in species diversity from the base to the middle of the zone, and at Sites 642 and 644 the LAD of the *Tectatodinium* sp. I, II, and III, *Operculodinium israelianum*, and *Polysphaeridium zoharyi* occurs in this interval, between the Brunhes/Matuyama boundary (0.73 Ma) and the middle part of the late Pleistocene (isotopic stage 8, ca. 0.44 Ma). In contrast to the disappearance of these gonyaulacoid taxa, the diversity of protoperidinioid species increases during the late Pleistocene. This part of Zone PM1 includes the FAD of *T. capitatum*, *Selenopemphix quanta*, and at Site 644, *Brigantedinium* species group A (including *B. cariacensis*, *B. auranteum*, and other indeterminate thick-walled round, brown protoperidinioid cysts).

PALEOENVIRONMENTAL INTERPRETATION

At present, few definite statements can be made about the paleoenvironmental significance of dinocyst assemblages from deep-sea locations because of conflicting views concerning their

probable sources as indigenous phytoplankton vs. current-transported particles (Dale, 1986). A few outstanding features that have been observed elsewhere in the North Atlantic nonetheless appear worthy of comment.

1. The high total diversity of dinocyst species ($S > 22$) in Zones PM5 and PM4, combined with the abundance of *Tuberculodinium vancompoae*, large number of *Spiniferites* species, common *Impagidinium* species, and the presence of many large cyst-forms with inflated periphragms or processes appear to be typical features of lower to middle Miocene sediments in the North Atlantic region and to be indicative of warm (subtropical), saline ($> 3.4\%$) surface waters (Harland, 1983; Wrenn and Kokinos, 1986). If the Iceland-Faroe Ridge was indeed very shallow or emergent during the earlier Miocene (Eldholm, Thiede, and Taylor, this volume), the inflow of subtropical water would have been very restricted. The warm water dinocysts thus suggests that the inflow of a large volume of cold arctic water to the Norwegian-Greenland Sea had not yet commenced, so that the relatively small volume of warm North Atlantic surface water could have a strong effect on the climate of the region.

2. Seven middle Miocene dinocyst species with widespread distributions in the North Atlantic (*I. aquaeductum*, *L. truncatum*, *C. ellipticum*, *P. laticinctum*, *H. obscura*, *S. placacantha*, and *D. pastielsii*) disappear in Zone PM3 prior to the Tortonian/Messinian boundary at 6.5 Ma. This attrition is contrary to the supposed subsidence of the Iceland-Faroe Ridge by 10 Ma which should have increased the inflow of North Atlantic water, thereby maintaining the warm surface water and introducing more North Atlantic species rather than eliminating species. Furthermore, a similar attrition of species is seen at DSDP Site 611 south of Iceland (Mudie, 1986), and it is believed that this widespread extinction may reflect the early stages of high-latitude climatic cooling, as found in eastern Iceland pollen studies (Mudie and Helgason, 1983).

3. Although total species diversity remains relatively high ($S = 16 - 26$) in the uppermost Miocene and lower Pliocene intervals at the Vøring Plateau, the most abundant palynomorphs are often small acritarchs of uncertain botanical origin (*Impletosphaeridium* spp., *Platycystidia*, and ?*Cymatiosphaera*). At the Vøring Plateau and central North Atlantic sites from Iceland to the Azores (Mudie, 1986), these small cyst-forms are associated with abundant diatoms and/or black spores. It is thus possible that these acritarchs are organic walled resting spores of marine algae, e.g., Chrysomonads or Archeomonads. It is also notable that dinocyst abundance in these samples is often lower in the presence of abundant diatoms. This may reflect the fact that diatoms are more efficient users of inorganic nutrients; hence, they suppress growth of dinoflagellates when light and nutrients are not limiting growth factors.

4. It is notable that three common oceanic subarctic North Pacific dinocysts, *F. filifera*, *N. lemniscata*, and *I. pallidum*, have their first appearances in the Norwegian Sea at or just above the Miocene/Pliocene boundary. This suggests that the East Greenland Current may have formed at that time, thereby transporting arctic water into the Norwegian-Greenland Sea. *F. filifera* and *I. pallidum* occur in Pliocene sediments of the central Arctic Ocean (Aksu and Mudie, 1985a), indicating that there was more inflow of North Pacific water prior to the Pleistocene lowering of sea level over the Bering Strait; hence, more outflow of arctic water would also be expected. This appearance of arctic/subarctic indicators also coincides with a major reduction in the Norwegian Sea occurrence of *I. patulum* and *I. aculeatum* which are typical warm temperate North Atlantic oceanic species (Harland, 1983). In contrast, the ubiquitous opportunistic species, *O. centrocarpum* started to become abundant during the Miocene/Pliocene transition often in association with *B. tepikiense* which is another species that appears to be toler-

ant of major shifts in seasonal temperature and salinity (Harland, 1983; Mudie, 1980).

5. Within the Pliocene interval, twelve cyst taxa disappear from the Norwegian Sea sites, with six of these losses occurring at the Pliocene/Pleistocene boundary. Most of these species are gonyaulacoid taxa which are common in middle Miocene sediments of the North Atlantic. It is likely that their disappearance during the late Pliocene reflects the onset of major continental glaciations in the Northern Hemisphere.

6. Another major reduction in dinocyst species occurs at or just above the lower/middle Pleistocene boundary. This involves the disappearance of *Polysphaeridium zoharyi* and *Operculodinium israelianum* which are presently dominant species in subtropical-warm temperature regions (Harland, 1983). There is a major reduction in the amplitude of palynomorph abundance cycles around this time interval, and productivity appears to diminish greatly above isotopic stage II (ca. 440 Ka). In general, the latest Pleistocene is marked by a relatively low total species diversity, with about ten species during interglacial stages and five species (mostly protoperidinioids) in the glacial stages. This pattern is consistent with detailed records from sediments of late Weichselian and Flandrian age in the southern Norwegian Sea region (Dale, 1985; Harland, 1984) and upper Pleistocene sediments in the Labrador Sea (Aksu and Mudie, 1985b). In general, there appears to be a correlation between dominance of protoperidinioids and adverse conditions of low-light levels resulting from either a thick ice cover or the discharge of sediment-laden meltwater. It is also notable that only one new gonyaulacoid cyst-form, *Ataxiodinium choanum*, appeared in the Norwegian Sea during the late Pleistocene, in contrast to the appearance of seven protoperidinioid taxa (*B. simplex*, *M. minuta*, *S. quanta*, *P. faeroense* cyst-form of Dale (1977), *T. capitatum*, *Xandarodinium xanthum*, and *Z. lenticulatum*). This feature seems to support previous observations (Dale, 1985; Mudie, 1980) that heterotrophic peridinioid dinoflagellates are better adapted to survive in the low-light conditions dictated by the presence of extensive sea-ice cover and low-angle irradiation north of the Arctic Circle.

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APPENDIX: List of dinocysts and acritarchs in range charts

(Note: L & W = Lentin and Williams)

Figure No.						Taxon
5	6	7	8	9	10	
1	—	1	—	—	20	<i>Ataxiodinium choanum</i> Reid 1974
2	—	2	14	—	1	<i>Brigantedinium simplex</i> (Wall) Reid, 1977
3	3	3	8	2	2	<i>Bitectatodinium tepikiense</i> Wilson, 1973
4	4	4	16	3	3	<i>Brigantedinium</i> species group A (this paper)
5	9	—	9	18	—	<i>Impagidinium pallidum</i> Bujak, 1984
6	38	27	10	28	19	<i>Impagidinium</i> species (mostly <i>I. patulum</i> (Wall) Stover and Evitt, 1978, and <i>I. aculeatum</i> (Wall) L & W, 1981)
7	—	6	2	—	5	<i>Multispinula minuta</i> Harland and Reid in Harland et al., 1980
8	13	7	11	20	15	<i>Nematosphaeropsis labyrinthea</i> (Ostenfeld) Reid, 1974
9	14	8	3	6	6	<i>Operculodinium centrocarpum</i> (Deflandre & Cookson) Wall, 1967
10	8	9	4	30	7	<i>Peridinium faeroense</i> cyst from of Dale (1977)
11	12	—	24	—	18	<i>Spiniferites bentori</i> (Rossignol) Wall & Dale, 1970
12	41	14	5	7	21	<i>Spiniferites elongatus</i> Reid, 1974 and intergrades with <i>Spiniferites frigidus</i> Harland and Reid in Harland et al., 1980
13	15	11	6	10	9	<i>Spiniferites</i> species (species other than those listed here)
14	15	23	28	17	14	<i>Tectatodinium pellitum</i> Wall, 1967
15	11	5	1	21	4	<i>Lingulodinium machaerophorum</i> (Deflandre & Cookson) Wall, 1967
16	1	32	7	22	24	<i>Achomosphaera andalusiensis</i> Jan du Chêne, 1977
17	—	12	25	9	27	<i>Spiniferites scabratus</i> (Wall) Sarjeant, 1970
18	—	—	18	—	26	<i>Stelladinium reidi</i> Bradford, 1975
19	—	17	—	—	10	<i>Trinovantedinium capitatum</i> Reid, 1977
20	25	29	13	16	15	<i>Selenopemphix nephroides</i> Benedek, 1972
21	—	24	15	4	13	<i>Brigantedinium</i> species group B (this paper)
22	37	44	32	25	—	<i>Spiniferites rubinus</i> (Rossignol) Sarjeant, 1970
24	34	18	26	14	34	<i>Tectatodinium</i> species I of de Vernal and Mudie, 1988a
24	18	19	17	15	25	<i>Tectatodinium</i> species II of de Vernal Mudie, 1988a
25	45	—	31	24	—	<i>Achomosphaera</i> species A of Matsuoka, 1983
26	5	20	—	33	—	Dinocyst species 1 (this paper)
27	36	56	—	27	28	<i>Impagidinium sphaericum</i> (Wall) L & W, 1981
28	30	16	23	29	30	<i>Operculodinium israelianum</i> (Rossignol) Wall, 1967
29	31	10	12	—	23	<i>Polysphaeridium zoharyi</i> (Rossignol) Bujak et al., 1980
30	19	31	21	11	—	<i>Tectatodinium</i> species III (this paper)
31	—	25	29	41	22	<i>Impagidinium multiplexum</i> (Wall and Dale) L & W, 1981
32	28	—	30	39	31	<i>Nematosphaeropsis lemniscata</i> Bujak, 1984
33	6	21	22	5	35	<i>Filispheera filifera</i> Bujak, 1984
34	42	22	33	13	—	<i>Spiniferites</i> species "TS" (this paper)
35	—	25	—	31	—	<i>Palaeostomocystis</i> species (a heterogeneous group of cysts resembling species of <i>Palaeostomocystis</i> Deflandre, 1973 described by Mudie (1986))

Appendix (continued).

Figure No.						Taxon
5	6	7	8	9	10	
36	2	15	19	1	23	<i>Achomosphaera ramulifera</i> (Deflandre) Evitt, 1963
37	33	43	—	32	—	<i>Operculodinium crassum</i> Harland, 1979
38	20	35	—	23	—	<i>Amiculosphaera umbracula</i> Harland, 1979
39	27	36	—	—	—	<i>Cymatiosphaera</i> species 1 of de Vernal and Mudie, 1988a
40	10	41	—	34	—	<i>Impletosphaeridium</i> species (undifferentiated small forms of <i>Impletosphaeridium</i> Morgenroth, 1966 as described by Mudie (1986))
41	29	—	—	40	—	<i>Nematosphaeropsis oblonga</i> Mudie, 1986
42	22	—	—	45	—	<i>Operculodinium echigoense</i> Matsuoka, 1983
43	—	34	—	43	—	<i>Operculodinium longispinigerum</i> Matsuoka, 1983
44	23	61	—	36	—	<i>Operculodinium walli</i> Matsuoka, 1983
45	40	—	—	—	—	<i>Corrudinium harlandi</i> Matsuoka, 1983
46	39	28	—	42	—	<i>Operculodinium</i> species of Jan du Chêne, 1977
47	8	38	—	35	—	<i>Hystrichokolpoma</i> species of Edwards (1984)
48	17	30	—	37	38	<i>Tectatodinium simplex</i> (Harland) Edwards, 1984
49	24	—	—	48	—	<i>Platycystidia</i> species 1 of Mudie (1986)
50	—	42	—	44	—	<i>Labyrinthodinium</i> species 1 (this paper)
51	12	33	—	47	—	<i>Melitasphaeridium choanophorum</i> (Deflandre & Cookson) Harland and Hill, 1979
52	44	52	—	—	—	<i>Pyxidiella</i> species 1 of Mudie (1986)
53	35	38	—	—	—	<i>Hystrichosphaeropsis pontiana</i> (Baltes) Stover and Evitt, 1978
54	—	—	—	46	—	<i>Batiacasphaera sphaerica</i> Stover, 1977
55	—	53	—	—	—	<i>Spiniferites pachyderma</i> (Rossignol) Reid, 1974
56	—	37	—	—	—	<i>Dapsilidinium pastielsii</i> (Davey & Williams) Bujak et al., 1980
57	32	47	—	—	—	<i>Reticulosphaera actinocoronata</i> (Benedek) Bujak and Matsuoka, 1986b
58	—	49	—	—	—	<i>Batiacasphaera</i> species II of Edwards (1984)
59	—	55	—	—	—	<i>Cyclopsella elliptica</i> Drugg & Loeblich, 1967
60	—	50	—	—	—	<i>Hystrichosphaeropsis obscura</i> Habib, 1972
61	—	46	—	—	—	<i>Operculodinium</i> species of Piasecki (1980)
62	—	54	—	—	—	<i>Impagidinium aquaeductum</i> (Piasecki) L & W, 1985
63	—	57	—	—	—	<i>Labyrinthodinium truncatum</i> Piasecki, 1980
64	—	58	—	—	—	<i>Pentadinium laticinctum</i> Gerlach, 1961
65	—	63	—	—	—	<i>Systematophora placacantha</i> (Deflandre & Cookson) Davey et al., 1969
66	—	48	—	—	—	<i>Tectatodinium</i> species of Piasecki (1980)
—	7	—	—	—	—	<i>Hystrichokolpoma</i> species 1 of Mudie (1986)
—	21	40	—	—	—	<i>Invertocysta</i> species (mostly <i>I. lacymosa</i> Edwards, 1984)
—	—	51	—	—	—	<i>Invertocysta tabulata</i> Edwards, 1984

Appendix (continued).

Figure No.						Taxon
5	6	7	8	9	10	
—	26	—	—	—	8	<i>Selenopemphix quanta</i> (Bradford) Matsuoka, 1985
—	43	—	—	—	—	<i>Quinquecuspis</i> species (undifferentiated)
—	—	45	—	—	12	<i>Tuberculodinium vancampoe</i> (Rossignol) Wall, 1967
—	—	59	—	—	—	<i>Dinopterygium verriculum</i> Piasecki, 1980 = <i>Heteraulacacysta</i> (Piasecki) L & W, 1981
—	—	60	—	—	—	<i>Hystichosphaeropsis obscura</i> Habib, 1972
—	—	62	—	—	—	<i>Palaeocystodinium</i> species A of Brown and Downie (1984)
—	—	64	—	—	—	<i>Achomosphaera crassipellis</i> (Deflandre & Cookson) Stover and Evitt, 1978
—	—	65	—	—	—	? <i>Thalassiphora</i> cf. <i>Th. ?pensa</i> Stover, 1977
—	—	66	—	—	—	<i>Palaeocystodinium golzowense</i> Albert, 1961
—	—	67	—	—	—	<i>Emslandia spiridoides</i> (Benedek) Benedek and Sarjeant, 1981
—	—	—	20	12	32	<i>Quinquecuspis</i> species (undifferentiated)
—	—	—	27	—	—	<i>Selenopemphix</i> cf. species B of Powell (1986b)
—	—	—	34	26	29	? <i>Cordosphaeridium</i> species 1 of de Vernal and Mudie, 1988b
—	—	—	—	—	8	<i>Spiniferites pseudofurcatus</i> (Klumpp) Sarjeant, 1970
—	—	—	—	—	11	<i>Xandarodinium xanthum</i> Reid, 1977
—	—	—	—	—	17	<i>Zygabikodinium lenticulatum</i> (Wall & Dale) Bujak and Davis, 1983

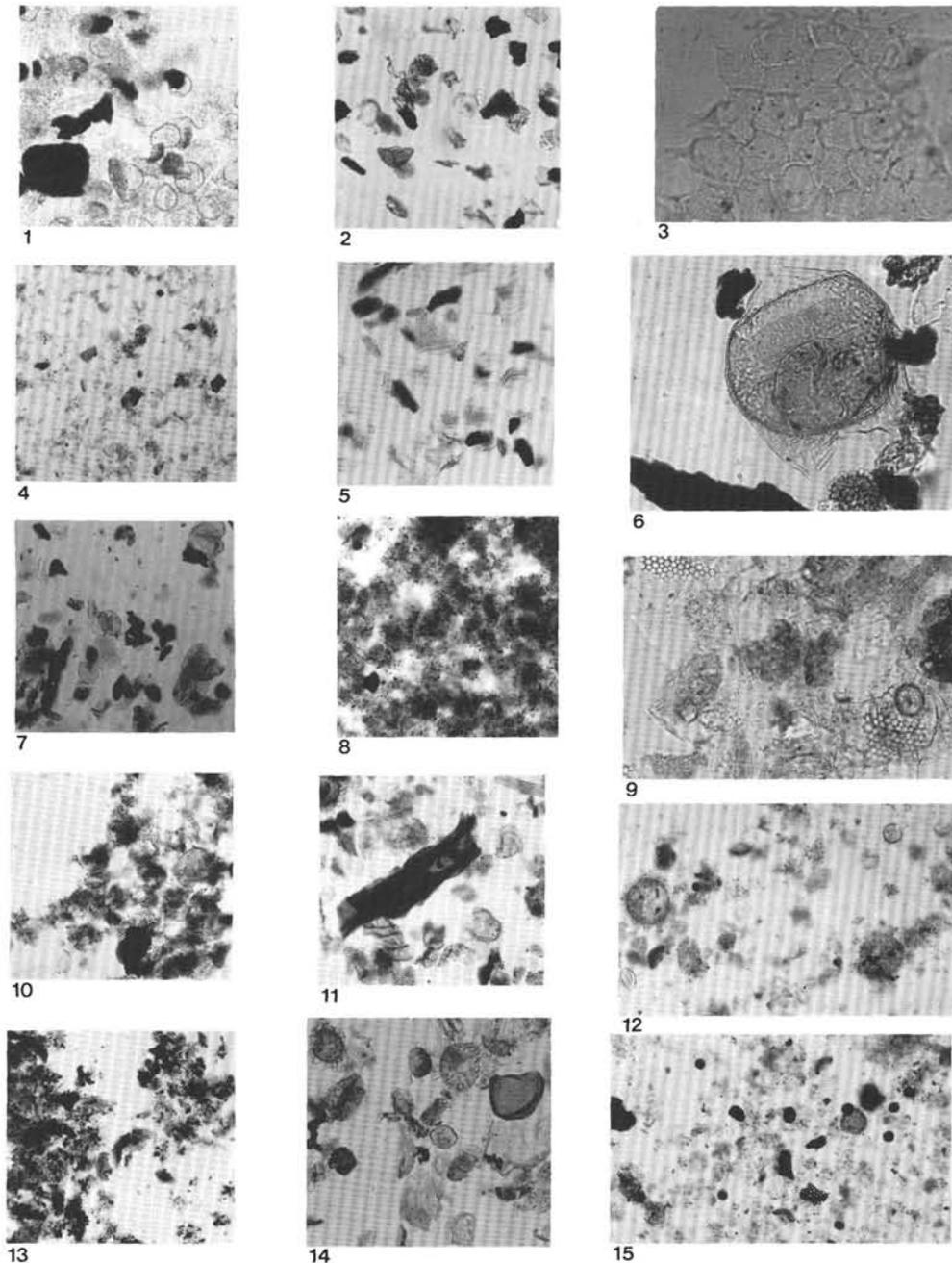


Plate 1. Magnification $\times 160$ for all figures except 3 and 6 ($\times 1000$) and 9 ($\times 400$). 1, 2, 3, 6. Upper Pleistocene palynofacies. 1, 3: Interglacial stage marine palynofacies; 1. 104-642B-1H-1, 0-5 cm: indigenous dinocysts embedded in yellow amorphous debris, with scattered wood fragments and black debris. 3. 104-644B-6H-4: *Pediastrum* coenobium. 2, 6: Ice-rafted palynofacies; 2. 104-642B-2H-6, 41-43 cm: large wood fragments, black debris, redeposited (Cretaceous-Paleogene) palynomorphs; 6. 104-644B-2H-6, 86-88 cm: well-preserved Paleogene dinocyst and poorly sorted wood fragments. 4, 5, 7. Upper Pliocene-lower Pleistocene palynofacies, Site 642. 4. 104-642-7H-5, 62-64 cm: marine palynofacies with well-dispersed small flocs of yellowish amorphous debris, small particles of wood and black debris; 5. 104-642B-7H-7, 62-64 cm: ice-rafted or fluvialite palynofacies with large wood and leaf fragments, degraded debris, poorly preserved reworked Tertiary palynomorphs; 7. 104-642B-5H-1, 71-78 cm: terrigenous or fluvialite palynofacies with abundant pollen, coarse wood fragments, black debris and reddish brown degraded debris. 8, 11. Upper Pliocene palynofacies, Site 644. 8. Marine palynofacies, 104-644A-29H-2, 35-37 cm: palynomorphs embedded in dense masses of dark brown amorphous debris with black specks; 11. 104-644A-34H-5, 85-87 cm: terrigenous palynofacies, with dinocysts and pollen interspersed with large wood particles, rare small flocs of amorphous debris. 9, 12, 15. Biosiliceous palynofacies. 9. Lower Pleistocene 104-644A-16H-4, 35-47 cm: dinocysts in matrix of amorphous yellow debris, faecal pellets, diatom fragments and resting spores, with thin sheets of translucent siliceous shards; 12. Upper Miocene 104-642C-11H-6, 136-137 cm: dinocysts coated with amorphous debris and black specks, interspersed with irregular and pellet-shaped flocs of amorphous debris. 15. Lower Pliocene 104-642C-11H-1, 19-20 cm: dinocysts interspersed with amorphous debris and black specks, common pyritised black spores (?diatoms), rare pyritised diatom fragments and reddish brown degraded debris. 10, 13. Upper Miocene-middle Pliocene palynofacies, Site 642. 10. 104-642B-8H-2, 61-63 cm: Pliocene dinocysts interspersed with or embedded in large flocs of brown amorphous debris with abundant black flecks and rare degraded brown debris; 13. 104-642B-16, CC: Upper Miocene dinocysts embedded in subangular flocs of brown amorphogen with abundant poorly sorted unstructured debris. 14. Lower-middle Miocene palynofacies, 104-643A-18H, CC (0-8 cm). Abundant dinocysts with rare flocs of yellowish amorphous debris and wood fibers.

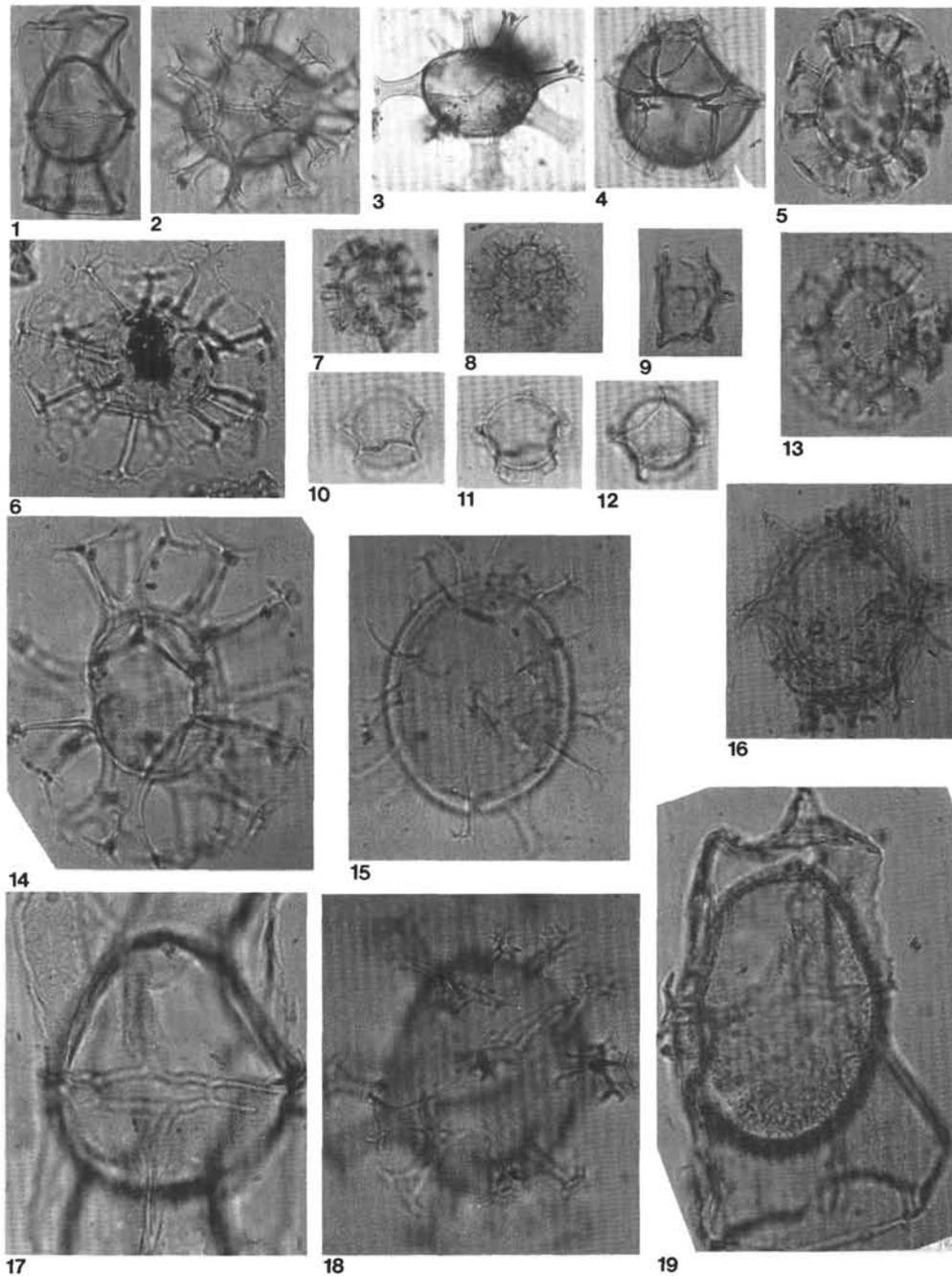


Plate 3. Magnification $\times 400$ for Figures 1-4, and $\times 1000$ for Figures 5-19. **1, 17** *Hystrichosphaeropsis ?pontiana*, 104-643A-9H, CC (9-11 cm), AGC 870226-30 (E22/0). **1**. Dorsal mid focus; **17**. dorsal surface focus, showing the smooth ovate body, wide cingulum, and finely scabrate periphragm surface. **2, 3** *Hystrichokolpoma* sp. of Edwards, 1984. **2**. 104-643A-16H, CC (23-25 cm); AGC 870226-22 (N25/2): ?Left lateral surface. **3**. 104-642B-19H, CC (9-11 cm), AGC 870122-15 (Z14/1): Apical mid focus. **4**. *Pentadinium laticinctum*, 104-643A-13H, CCG (20-22 cm), AGC 870318-1 (W25/0): Ventral surface. **5, 13** *Labyrinthodinium* sp. 1. 104-643A-8H, CC (28-30 cm), AGC 870226-7 (R27/4). **5**. Optical section, orientation unknown; **13**. Surface of trabeculate periphragm. **6**. *Reticulosphaera actinocoronata* 104-643A-14H, CC (10-12 cm), AGC 870226-16 (M31/0): Optical section. **7, 8** *Cymatiosphaera* sp. 1. 104-644A-24H-1, 35-37 cm; **7**. AGC 870404-1 (N19/3). Optical section of crests separated at the distal ends in places. **8** AGC 870404-2 (N17/0). Surface of undulating margins of crest fields. **9, 10, 11, 12**. *Platycystidia* sp. 1 104-642C-11H-1, 38-39 cm. **9**. AGC 870404-3 (P48/1). Surface of specimen with rectangular body, showing ?apical separation of outer wall layer **10, 11, 12**. AGC 870122-1 (W55/0). Apical view of specimen with circular body. **10**. Surface, showing small pylome with subrectangular outline; **11**. Optical section; **12**. Antapical focus. **14**. *Spiniferites* sp. "TS". 104-643A-8H, CC (28-30 cm), AGC 870226-4 (F37/3): Optical section of long narrow gonal spines with trifurcate distal ends. **15**. *Achomosphaera ?crassipellis*, 104-643A-11H, CC (42-44 cm), AGC 870318-35 (D34/2): Dorsal tectate body wall. **16**. *Achomosphaera* ?sp. A of Matsuoka (1983). 104-644A-16H-3, 135-137 cm, AGC 870205-11. (Z45/1: Dorsal surface focus. **18**. *Achomosphaera andalouisiensis*. 104-642A-1H-4, 100-102 cm, AGC 870220-20 (E19/4). Orientation uncertain, focus on process tips, with complex branching and fusion of tips. **19**. *Hystrichosphaeropsis obscura* 104-643A-8H, CC (28-30 cm); AGC 870226-5 (E28/3). Surface view of ellipsoidal central body with coarsely tectate ornament and granular appearance.

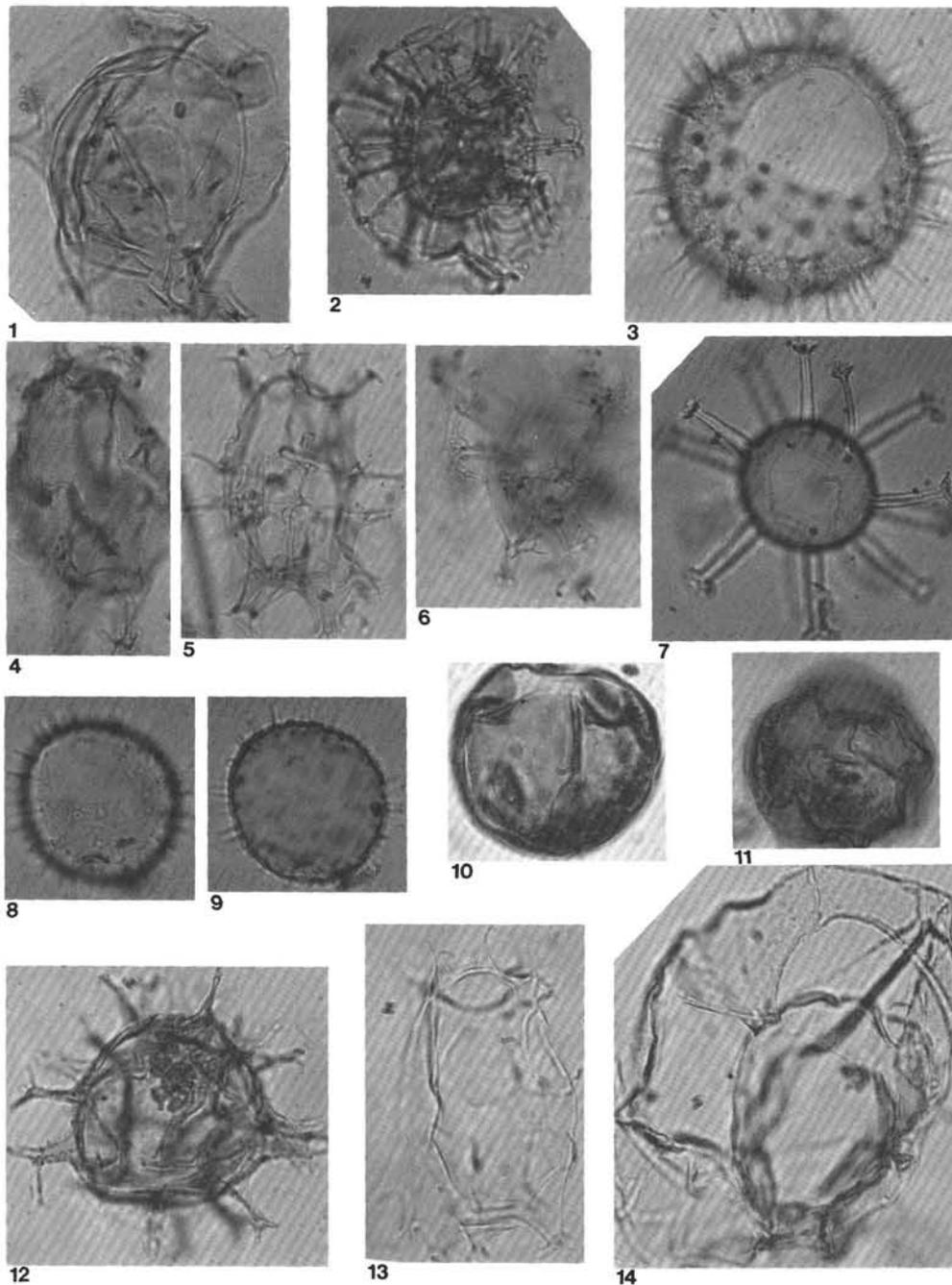


Plate 4. Magnification $\times 1000$. 1, 4 *Spiniferites rubinus* (Rossignol) 1. 104-642B-7H-3, 62-64 cm, AGC870220-32 (W18/1): Dorsal surface of cyst with wide crests linking gonal processes. 4. 104-644A-16H-3 (135-137 cm, AGC870205-35 (U45/0): Dorsal view of weak, box-like gonal processes. 2, 6 *Nematosphaeropsis lemniscata* 104-644A-10H-3, 135-137 cm, AGC870318-23 (T39/4): 2. ?Right lateral, optical section. 6. Surface of undulating ribbon-like trabeculae. 3. *Operculodinium crassum*, 104-642C-11H-4, 42-43 cm, AGC870122-20 (W33/2): Dorsal mid focus. 5, 13 *Spiniferites elongatus-frigidus* intergrades. 5. *S. elongatus* cyst type; 104-644A-16H-2, 35-37 cm, AGC 870205-7 (U29/1) 5. Dorsal surface figure. 13. *S. frigidus* cyst type. 104-642A-1H-6 100-102 cm, AGC870220-22 (Z38/3): Dorsal mid focus. 7. *Melitasphaeridium choanophorum*, 104-643A-8H,CC (28-30 cm), AGC870206-6 (D30/0): Dorsal epicyst, apical process in focus. 8, 9 *Peridinium faeroense* cyst form of Dale (1977). 104-642A-1H-2, 100-102 cm, AGC 870220-13 (U18/1): 8. ?Apical view of outer wall surface; 9. Optical section of the variable spine length. 10. *Brigantedinium* species group B. 104-644B-6H-2, 86-88 cm, AGC 870318-18 (G24/0): Dorsal antapical surface. 11. *Selenopemphix* ?sp. B of Powell (1986b). 104-644A-10H-5, 85-87 cm, AGC870404-7 (Z47/4). Right lateral epicyst surface. 12. *Spiniferites scabratus* 104-644B-15H-2, 86-88 cm, AGC870318-32 (Z50/4): Dorsal epicyst surface showing microgranular ornament on processes. 14. *Amiculosphaera umbracula* 104-644A-29H-2, 85-87 cm, AGC870205-38 (O44/4): Dorsal apical periphragm arch in focus.

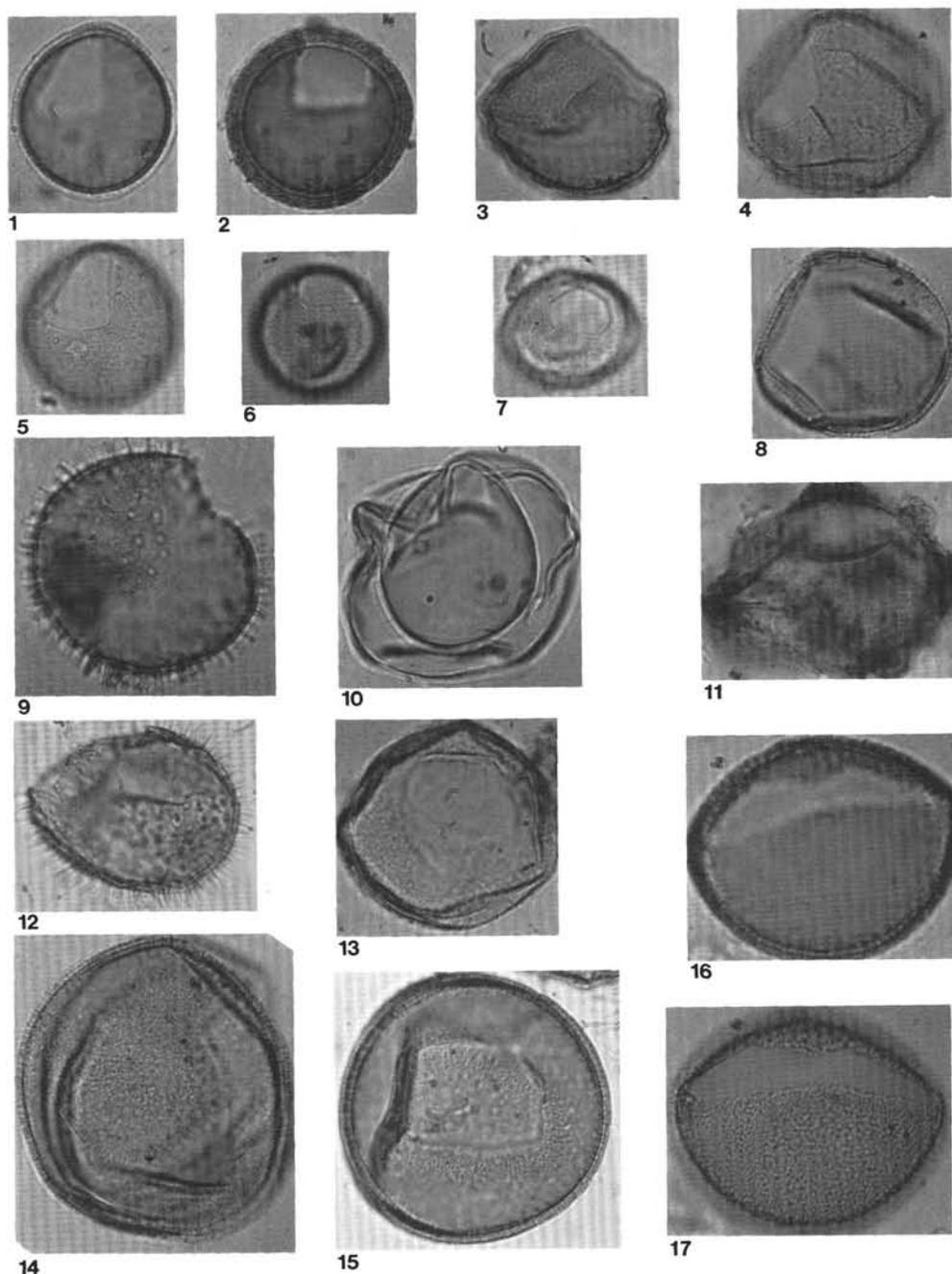


Plate 5. Magnification $\times 1000$ 1, 5 *Tectatodinium pellitum* 104-642A-1H-2, 100-102 cm, AGC870220-26 (Z16/1). 1. Dorsal optical section of outer wall layer and tectate ornament. 5. Surface of finely corrugated ornament. 2. ?*Tectatodinium* sp. of Piasecki, 1980. 104-643A-8H, CC (28-30 cm), AGC870220-35 (O46/4): Dorsal optical section. 3. *Tectatodinium* sp. III. 104-644A-24H-3, 35-37 cm, AGC870213-17 (W30/3): ?Ventral view, mid focus on paracingulum and tectate wall. 4, 8 Dinocyst sp. 1. 104-644A-29H-2, 35-37 cm, AGC870213-11 (O43/0) 4. ?Right lateral surface. 8. Optical section of coarsely tectate ornament. 6, 7 ?*Pyxidiella* sp. 1. 104-643A-13H, CC (20-22 cm), AGC870318-12 (Y37/0): 6. Surface of specimen with notched apical pylome. 7. AGC870318-13 (W45/0): Apical view of specimen with pentagonal pylome. 9, 12 *Multispinula minuta*: 9. 104-642A-1H-4, 100-102 cm, AGC870220-21 (X29/4): Left lateral optical section of short, acuminate processes and attached archeopyle; 12. Sample 104-644B-1H-1, 100-102 cm; AGC860413-2 (Y31/4): Dorsal optical section of cyst form with acicular processes (wall on right is torn). 10. *Ataxiodinium choanum* 104-642A-1H-2 100-102 cm, AGC870220-12 (O37/0): Dorsal surface. 11. ?*Cordosphaeridium* sp. 1. 104-644A-17H-2, 31-33 cm, AGC 870205-30 (W38/2): ?Dorsal optical section. 13. *Tectatodinium* sp. II. 104-644A-24-3, 84-87 cm, AGC870404-5 (K12/0): Dorsal optical section of surface. 14. *Filisphaera filifera*, 104-642B-7H-4, 62-64 cm, AGC870220-33 (T23/1): Left lateral mid focus on thin finely granular outer wall and pilose ornament. 15. *Tectatodinium* sp. I. 104-644A-16H-4 (35-37 cm, AGC870205-17 (P15/1): Dorsal optical section of thick outer wall, coarsely tectate ornament and finely corrugate wall surface. 16, 17 *Bitectatodinium tepikiense*, 104-642A-1H-2, 100-102 cm, AGC870220-25 (Y40/4): 16. Dorsal optical section of thick outer wall and coarsely tectate ornament; 17. Surface of heavily corrugate outer wall.