Design, Synthesis and Characterization of Biogenic Chloroquine Silver Nanoparticles as Potential Anticancer Agent against Neuroblastoma Cells

S.C. VIVEKANANDHAN¹, M. CHANDRAMOHAN² and P. SELVAM^{3,*}

¹Department of Medicine, Thoothukudi Medical College, Thoothukudi-628 008, India

²Bharat Ratna Kamarajar Jaundice Liver Hospital and Research Centre, Madurai-625 001, India

³Sir CV Raman-K.S. Krishnan International Research Centre, Kalasalingam University, Krishnankoil-626 126, India

*Corresponding author: Fax: +91 4563 289322; E-mail: selvaamin@gmail.com

Received: 4 August 2017;

Accepted: 20 October 2017;

Published online: 31 January 2018;

AJC-18739

Chloroquine (4-aminoquinoline derivative) an old antimalarial drug is a versatile bioactive agent and reported to have board spectrum of activity including anticancer and antiviral activity and firmly repositioned in these latter indications. In literature report of chloroquine retinopathy upto 0.5 % was noted to circumvent this toxicity and increase the efficacy of the drug. We ventured to develop a novel approach of the green synthesis of chloroquine silver nanoparticles and anticancer activity of the same was tested by MTT against Neuroblastoma (SH-SY5Yc) and cytotoxicity was also investigated against normal Vero cells by MTT assay. We prepared chloroquine silver nanoparticles by an ecofriendly method and was characterized using FT-IR, UV-visible, SEM and PXRD techniques. Anticancer activity of chloroquine silver nanoparticles was tested against Neuroblastoma (SH-SY5Yc) by MTT assay and cytotoxicity in vero cells. The UV-visible spectrum for prepared silver nanoparticles shows sharp peak at 434 nm confirms the presence of silver nanoparticles. SEM analysis showed that the nanoparticles are spherical in shape and average size of the particles are 254 nm. Powder XRD studies, 2θ value (220), (111), (211) confirms that the biosynthesized nanoparticles are in crystalline nature. The prepared chloroquine silver nanoparticles showed significant anticancer activity against Neuroblastoma cells by MTT assay with inhibitory concentration (IC₅₀) of 33.79 µg/mL, where as cytotoxicity (CC₅₀) of chloroquine silver nanoparticles in Vero cells was found to be 173.79 µg/mL.

Keywords: Chloroquine, Silver nanoparticles, Anticancer Activity, Neuroblastoma cells.

INTRODUCTION

Cancer is a major threat to human health and enormous drugs in the market for the treatment of cancer. However, the cancer cells could develop rapid resistance to approved drugs due to high mutation rate. In order to overcome the resistance, new drugs with novel molecular mechanisms of action should be developed and recently much attention has been devoted for searching of potential anticancer agents. Chloroquine phosphate (CQ) is a versatile therapeutic agents and approved as antimalarial agent also, used for treatment of amoebiasis, rheumatoid arthritis, lupus erythematosus, photo allergic disorder, as a non specific anti-inflammatory agent. Chloroquine (CQ) also reported for board-spectrum antiviral activity [1] against HIV [2], HBV [3], HCV [4,5], influenza [6], dengue [7], SARS-CoV [8] and chikungunya virus [9].

Chloroquine reported for anticancer activity [10-12] and also enhancing anticancer activity of radiotherapy [13]. Mechanism of anticancer activity of chloroquine in by inhibiting autophagy and activating apoptosis [14,15]. Chloroquine

causes central nervous system toxicity of retinopathy and epilepsy up to 25 to 40 % in rheumatoid arthritis patients at 20 mg/kg dosage for prolonged period [16-18] and nanoparticles may reduce the same. Current work is to biosynthesis and characterization of the chloroquine silver nanoparticles (CQ AgNPs) and tested anticancer activity against Neuroblastoma (SH-SY5Yc) and cytotoxicity was also investigated against normal Vero cells by MTT assay.

EXPERIMENTAL

Preparation of silver nanoparticles: Silver nitrate (AgNO₃) was obtained from Hi Media (Mumbai, India). Aqueous solution of 1 mM AgNO₃ was used for the synthesis of silver nanoparticles. The 50 mL of silver nitrate solution was stirred for 10 min. 10 mg of Chloroquine was dissolved in ethanol and 0.5 mL of chloroquine solution was added to silver nitrate solution drop by drop continuously which was under stirring at 40° C. After 30 min the solution changes to reddish brown in colour. Colour change from colourless to reddish brown indicates the formation of chloroquine silver nanoparticles.

Characterization of silver nanoparticles: UV-visible spectra of the silver nanoparticles obtained from chloroquine for plasmon resonance band analyzed in Shimadzu Double beam UV-visible spectrophotometer. The scanning electron microscopy (SEM) analysis was carried by a Carl Zeiss EVO-18 electron micro-scope and visualized the morphological size and shape. A sample of a nanoparticles solution was placed on a carbon strip attached to a SEM brass, extra solution was detached using blotting paper and then allowed to dry by putting it under a mercury lamp for 5 min. The elemental compositions of chloroquine silver nanoparticles were obtained using BRUKER EDS-(QUANTAX 200-XFLASH) SDD (silicon drift detector) by variable pressure mode, at acceleration voltage of 20 KeV. The Fourier transform infrared spectra were recorded using IR Tracer-100 Shimadzu, FTIR and using the spectral range 4000-400 cm^{-1} . 50 mg chloroquine silver nanoparticles placed in ATR diamond disc (zinc selenide) for FTIR measurement. The crystalline and the lattice characteristics of the synthesized silver nanoparticles were measured by powder X-ray diffraction analyzed. The XRD measurement was carried out on thoroughly dried thin films of the purified chloroquine silver nanoparticles was dried on a glass slab on a D8 Advance ECO XRD Systems with SSD160 1D Detector-Bruker XRD 6000 instrument operated at a voltage of 20 keV and a current of 30 mA with $\text{CuK}_{\alpha 1}$ radiation ($\lambda = 0.1542$) in a 2θ ($^\circ$) configuration. The particle size of chloroquine silver nanoparticles were determined by using Beckman coulter particle size analyzer (Delsa™ Nano common).

Anticancer activity and cytotoxicity: Chloroquine silver nanoparticles were investigated for the anticancer activity against Neuroblastoma cell (SH-SY5Y cells), cytotoxicity also tested against Vero cells by MTT assay [19]. 50 % inhibitory concentration (IC_{50}) and 50 % cytotoxic concentrations (CC_{50}) were measured in $\mu\text{g/mL}$ to understand the anticancer activity and cytotoxicity effect, respectively.

RESULTS AND DISCUSSION

In the present work, authors synthesized silver nanoparticles by bioreduction method using silver nitrate solution as a source and chloroquine as a reducing agent. The process of silver nanoparticles generation is presented in Fig. 1a. In brief, the chloroquine added to the source solution and the mixture was stirred on a magnetic stirrer till colour changed to reddish brown. The colour change indicate the reduction of silver nitrate to silver nanoparticles. The UV-visible spectrum for prepared silver nanoparticles shows noticeable absorptions maximum at 434 nm, confirms the presence of silver nanoparticles and also absorptions maximum same upto 5 h (Fig. 1b). The SEM studies used to investigate the morphology of the silver nanoparticles and from SEM images confirms that the prepared nanoparticles in spherical shape and their morphology are shown in the Fig. 2. Fig. 3 revealed three peaks at degree 2θ 28.2°, 32.7° and 38.2° corresponding to three diffraction from (220), (111), (211) planes of silver (as correlated to JCPDS: File No. 4-783). The XRD studies show that biosynthesized nanoparticles are crystalline in nature. Chloroquine silver nanoparticles were of the sizes range from 17-400 nm and average size of the particles is 254 nm (Fig. 4).

Fig. 1a. UV-visible spectra of chloroquine silver nanoparticles at 30 min

Fig. 1b. UV-visible spectra of chloroquine silver nanoparticles at 1-5 h

Fig. 2. SEM analysis of chloroquine silver nanoparticles

Fig. 3. Particle size analysis of chloroquine silver nanoparticles

Fig. 4. XRD analysis of chloroquine silver nanoparticles

Polydispersity index was found to be 0.208, this indicates no nanoparticles aggregation and mono disperse of synthesized nanoparticles. IR spectrum indicates (Fig. 5) the broad band at near 3500 cm^{-1} shows the presence of N-H group and 2968 and 2929 cm^{-1} is for C-H stretching. The frequencies 900 - 690 cm^{-1} is for $=\text{C-H}$ bending occurs out of plane. 1610 and 1452 cm^{-1} is for the C=C ring stretch also 1579 cm^{-1} is for the N-H and C-N stretching in the spectrum.

The prepared chloroquine silver nanoparticles showed significant anticancer activity against Neuroblastoma cells with inhibitory concentration (IC_{50}) of $33.79\text{ }\mu\text{g/mL}$, whereas cytotoxicity (CC_{50}) of AgNPs in Vero cells was found to be $173.79\text{ }\mu\text{g/mL}$ both by MTT assay (Figs. 6a,b and 7).

Conclusion

Chloroquine nanoparticles were reported for anticancer activity with inhibition of autophagy [20,21], antimalarial and

Fig. 5. FT IR spectrum of chloroquine silver nanoparticles

Fig. 6a. Cytotoxicity of chloroquine silver nanoparticles in Vero cells

Fig 6b. Anticancer activity of chloroquine silver nanoparticles in Neuroblastoma cells

antidengue virus activities [22,23]. We have successfully synthesized silver nanoparticles using chloroquine as a reducing agent and the prepared nanoparticles were confirmed by the characterization technique UV-visible, FTIR, SEM and XRD. We have shown the anticancer potential of chloroquine silver nanoparticles and the inhibitory concentration (IC_{50}) of $33.79\text{ }\mu\text{g/mL}$.

ACKNOWLEDGEMENTS

The authors sincerely thank Dr. Ashish Wadhwani, Head, Department of Pharmaceutical Biotechnology, JSS College of Pharmacy, Uthagamandalam, India for cytotoxicity studies.

REFERENCES

1. J.M. Rolain, P. Colson and D. Raoult, *Int. J. Antimicrob. Agents*, **30**, 297 (2007); <https://doi.org/10.1016/j.ijantimicag.2007.05.015>.
2. M.R. Fesen, K.W. Kohn, F. Leteurtre and Y. Pommier, *Proc. Natl. Acad. Sci. USA*, **90**, 2399 (1993); <https://doi.org/10.1073/pnas.90.6.2399>.
3. G. Civitico, Y.Y. Wang, C. Luscombe, N. Bishop, G. Tachedjian, I. Gust and S. Locarnini, *J. Med. Virol.*, **31**, 90 (1990); <https://doi.org/10.1002/jmv.1890310205>.
4. M. Chandramohan, S.C. Vivekananthan, D. Sivakumar, P. Selvam, J. Neyts, G. Katrien and E. De Clercq, *Indian J. Pharm. Sci.*, **68**, 538 (2006); <https://doi.org/10.4103/0250-474X.27842>.

Fig. 7. Anticancer activity and cytotoxicity of chloroquine silver nanoparticles

5. U.A. Ashfaq, T. Javed, S. Rehman, Z. Nawaz and S. Riazuddin, *Virol. J.*, **8**, 163 (2011); <https://doi.org/10.1186/1743-422X-8-163>.
6. N.I. Paton, L. Lee, Y. Xu, E.E. Ooi, Y.B. Cheung, S. Archuleta, G. Wong and A.W. Smith, *Lancet Infect. Dis.*, **11**, 677 (2011); [https://doi.org/10.1016/S1473-3099\(11\)70065-2](https://doi.org/10.1016/S1473-3099(11)70065-2).
7. K.J. Farias, P.R. Machado, J.A. Muniz, A.A. Imbeloni and B.A. Da Fonseca, *Viral Immunol.*, **28**, 161 (2015); <https://doi.org/10.1089/vim.2014.0090>.
8. E. Keyaerts, L. Vijgen, P. Maes, J. Neyts and M.V. Ranst, *Biochem. Biophys. Res. Commun.*, **323**, 264 (2004); <https://doi.org/10.1016/j.bbrc.2004.08.085>.
9. M. Khan, S.R. Santhosh, M. Tiwari, P.V. Lakshmana Rao and M. Parida, *J. Med. Virol.*, **82**, 817 (2010); <https://doi.org/10.1002/jmv.21663>.
10. S. Hatzieremia, P. McCall, J. Willder, C. Orange, M. Seywright, M.A. Underwood and J. Edwards, *Cancer Res.*, **73**, 1 (2013); <https://doi.org/10.1158/1538-7445.AM2013-1>.
11. M. Gwenola, O. Florine, K. Guido, V. Illo and G. Lorenzo, *Mol. Cel. Oncol.*, e29911 (2014); <https://doi.org/10.4161/mco.29911>.
12. V. Solomon and H. Lee, *Eur. J. Pharmacol.*, **625**, 220 (2009); <https://doi.org/10.1016/j.ejphar.2009.06.063>.
13. Y.C. Lin, J.F. Lin, S.I. Wen, S.C. Yang, T.F. Tsai, H.E. Chen, K.Y. Chou and T.I. Hwang, *Kaohsiung J. Med. Sci.*, **33**, 215 (2017); <https://doi.org/10.1016/j.kjms.2017.01.004>.
14. M.E. Egger, J.S. Huang, W. Yin, K.M. McMasters and L.R. McNally, *J. Surg. Res.*, **184**, 274 (2013); <https://doi.org/10.1016/j.jss.2013.04.055>.
15. H. Zhao, Y. Liao, D. Xu, Q. Wang, Q. Gan, C. You and C. Yang, *BMC Neurol.*, **16**, 1 (2016); <https://doi.org/10.1186/s12883-015-0524-9>.
16. M.F. Marmor, U. Kellner, T.Y. Lai, R.B. Melles and W.F. Mieler, *Am. Acad. Ophthalmol.*, **123**, 1386 (2016); <https://doi.org/10.1016/j.ophtha.2016.01.058>.
17. L.S. Leung, J.W. Neal, H.A. Wakelee, L.V. Sequist and M.F. Marmor, *Am. J. Ophthalmol.*, **160**, 799 (2015); <https://doi.org/10.1016/j.ajo.2015.07.012>.
18. E.V. Navajas, H. Crema, D.S. Hammoudi, J.H. Lipton, E.R. Simpson, S. Boyd and M. Easterbrook, *Can. J. Ophthalmol.*, **50**, 442 (2015); <https://doi.org/10.1016/j.cjco.2015.08.003>.
19. P. Skehan, R. Storeng, D. Scudiero, A. Monks, J. McMahon, D. Vistica, J.T. Warren, H. Bokesch, S. Kenney and M.R. Boyd, *J. Natl. Cancer Inst.*, **82**, 1107 (1990); <https://doi.org/10.1093/jnci/82.13.1107>.
20. P. Joshi, S. Chakraborty, S. Dey, V. Shanker, Z.A. Ansari, S.P. Singh and P. Chakrabarti, *J. Colloid Interf. Sci.*, **355**, 402 (2011); <https://doi.org/10.1016/j.jcis.2010.12.032>.
21. P. Joshi, S. Chakraborty, J.E. Ramirez-Vick, Z.A. Ansari, P. Chakraborty and S.P. Singh, *Colloids Surf. B Biointerf.*, **15**, 195 (2012); <https://doi.org/10.1016/j.colsurfb.2012.02.039>.
22. S. Tripathy, S. Das, S.P. Chakraborty, S.K. Sahu, P. Pramanik and S. Roy, *Int. J. Pharm.*, **434**, 292 (2012); <https://doi.org/10.1016/j.ijpharm.2012.05.064>.
23. K. Murugan, J. Wei, M.S. Alsallhi, M. Nicoletti, M. Paulpandi, C.M. Samidoss, D. Dinesh, B. Chandramohan, C. Paneerselvam, J. Subramaniam, H. Wei, C. Vadivalagan, P. Amuthavalli, A. Jaganathan, S. Devanesan, A. Higuchi, S. Kumar, A.T. Aziz, D. Nataraj, B. Vaseeharan, A. Canale and G. Benelli, *Parasitol. Res.*, **116**, 495 (2017); <https://doi.org/10.1007/s00436-016-5310-0>.