Synthesis and Antifungal Activity of Aspirin Derivatives

SUMEI GAO¹, ZHIHONG XU¹, XUESONG WANG¹, HUI FENG², LING WANG¹, YUANJIANG ZHAO¹, YUE WANG¹ and XIAORONG TANG^{1,*}

¹School of Physics and Chemistry, Xihua University, Chengdu 610039, P.R. China

²School of Bioengineering, Xihua University, Chengdu 610039, P.R. China

*Corresponding authors: E-mail: tangxiaorong0901@163.com

Received: 16 October 2013;

Accepted: 14 March 2014;

Published online: 30 September 2014;

AJC-16092

Using aspirin as a lead compound, a series of its derivatives (compounds **1-7**) were designed and synthesized. Their activity of anti-pathogenic fungi of plants has been evaluated in the laboratory. The results showed that these compounds had good antifungal activity against *Sclerotinia sclerotiorum*, *Helminthosporium maydis*, *Botrytis cinerea* and *Rhizoctonia solani*. Among them, the inhibition of growth for compounds **1** and **2** against *Helminthosporium maydis* reached 92.5 and 91.6 % at a concentration of 100 mg L⁻¹, respectively, which was superior to carbendazim.

Keywords: Aspirin derivatives, Antifungal activity.

INTRODUCTION

Sclerotinia sclerotiorum, *Helminthosporium maydis*, *Botrytis cinerea* and *Rhizoctonia solani* are harmful pathogenic fungi of crops or vegetables¹⁻⁴. Over the past decades, synthetic fungicides including carbendazim have been used to prevent them. However, in recent years, they have developed resistance to the fungicides⁵⁻⁹. Moreover, their scope of resistance continues to expand and has already included many new fungicides¹⁰⁻¹². Therefore, new fungicides are continually required.

It is well-known that since aspirin (acetylsalicylic acid) was first marketed in 1899, it has been widely used for the treatment of pains, fever and colds¹³⁻¹⁶. Therefore, in the present study, aspirin derivatives were synthesized based on aspirin. In the meantime, their antifungal activity has been evaluated in the laboratory to find new fungicides with high efficacy and low toxicity.

EXPERIMENTAL

Sclerotinia sclerotiorum, *Helminthosporium maydis*, *Botrytis cinerea* and *Rhizoctonia solani* were obtained from the Chinese

Academy of Agricultural Sciences. They were preserved at 4 °C. All chemicals and solvents were purchased from commercial sources unless specified otherwise. IR spectra were recorded on a Thermofisher Nicolet-6700 spectrophotometer. ¹H NMR spectra were taken on a Varian Unity Inova-400 instrument using deuterium-chloroform as the solvent.

Synthesis of target compounds: The target compounds were synthesized in accordance with the reaction shown in Fig. 1. Aniline or its derivatives (0.01 mol) and pyridine (0.01 mol) were dissolved in CH₂Cl₂ (15 mL). The mixture was stirred and heated to 35-45 °C. 2-(Chlorocarbonyl)phenyl acetate (0.01 mol) with CH₂Cl₂ (15 mL) was slowly added to the above mixture under stirring until the reaction was complete. The precipitate was filtered and washed with still water. The pure compounds were obtained by re-crystallization in anhydrous ethanol.

Compound 1 (C₁₅H₁₃O₃N): Light yellow powder; yield: 75 %; m.p. 152-153 °C; IR (KBr, ν_{max}, cm⁻¹): 3243, 3191, 3131, 3072, 3040, 1760, 1649, 1597, 1542, 1494, 1451, 1370, 1327, 1300, 1195, 1156, 1097, 758, 748, 709, 596; ¹H NMR (400 MHz, CDCl₃) δ (ppm): 8.08 (s, 1H), 7.83 (d, *J* = 7.6 Hz, 1H),

Fig. 1. Synthetic method of target compounds **1-7**

7.60 (d, $J = 8.0$ Hz, 2H), 7.49-7.53 (m, 1H), 7.33-7.38 (m, 3H), 7.13-7.17 (m, 2H), 2.32 (s, 3H).

Compound 2 ($C_{15}H_{12}O_3NCl$): White needle crystal; yield: 40 %; m.p. 133-134 °C; IR (KBr, $\nu_{\max}$, cm^{-1}): 3487, 3285, 3190, 3111, 3048, 1755, 1656, 1608, 1595, 1533, 1493, 1447, 1396, 1317, 1207, 1172, 830, 751, 701, 589; 1H NMR (400 MHz, $CDCl_3$) δ (ppm): 8.10 (s, 1H), 7.80 (d, $J = 7.6$ Hz, 1H), 7.49-7.55 (m, 3H), 7.30-7.36 (m, 3H), 7.14 (d, $J = 8.4$ Hz, 1H), 2.31 (s, 3H).

Compound 3 ($C_{15}H_{12}O_3NBr$): Light yellow needle crystal; yield: 82 %; m.p. 138-139 °C; IR (KBr, $\nu_{\max}$, cm^{-1}): 3485, 3281, 3183, 3105, 3042, 1755, 1656, 1597, 1607, 1587, 1526, 1490, 1446, 1391, 1317, 1208, 1173, 826, 750, 700, 589; 1H NMR (400 MHz, $CDCl_3$) δ (ppm): 8.15 (s, 1H), 7.77 (d, $J = 7.6$ Hz, 1H), 7.42-7.52 (m, 5H), 7.32 (t, $J = 7.6$ Hz, 1H), 7.13 (d, $J = 8.0$ Hz, 1H), 2.30 (s, 3H).

Compound 4 ($C_{15}H_{12}O_5N_2$): Golden yellow crystal; yield: 54 %; m.p. 90-91 °C; IR (KBr, $\nu_{\max}$, cm^{-1}): 3315, 3123, 3068, 3030, 1775, 1675, 1604, 1585, 1501, 1451, 1370, 1298, 1176, 1146, 1121, 812, 744, 693, 592; 1H NMR (400 MHz, $CDCl_3$) δ (ppm): 11.10 (s, 1H), 8.91 (d, $J = 8.4$ Hz, 1H), 8.24 (d, $J = 8.4$ Hz, 1H), 7.95 (d, $J = 7.6$ Hz, 1H), 7.71 (t, $J = 7.6$ Hz, 1H), 7.58 (t, $J = 7.6$ Hz, 1H), 7.40 (t, $J = 7.6$ Hz, 1H), 7.21-7.26 (m, 2H), 2.36 (s, 3H).

Compound 5 ($C_{16}H_{15}O_3N$): Light yellow crystal; yield: 85 %; m.p. 139-140 °C; IR (KBr, $\nu_{\max}$, cm^{-1}): 3494, 3280, 3193, 3122, 3060, 2921, 1756, 1654, 1606, 1533, 1513, 1446, 1382, 1220, 1206, 812, 762, 701; 1H NMR (400 MHz, $CDCl_3$) δ (ppm): 7.98 (s, 1H), 7.84 (d, $J = 7.6$ Hz, 1H), 7.47-7.53 (m, 3H), 7.35 (t, $J = 7.4$ Hz, 1H), 7.16 (t, $J = 7.2$ Hz, 3H), 2.34 (s, 3H), 2.32 (s, 3H).

Compound 6 ($C_{16}H_{15}O_4N$): Light purple solid; yield: 77 %; m.p. 119-120 °C; IR (KBr, $\nu_{\max}$, cm^{-1}): 3341, 3246, 3123, 3044, 3009, 2937, 2839, 1760, 1643, 1608, 1599, 1513, 1445, 1371, 1301, 1219, 1198, 816, 759, 634; 1H NMR (400 MHz, $CDCl_3$) δ (ppm): 7.94 (s, 1H), 7.83 (d, $J = 8.0$ Hz, 1H), 7.49-7.52 (m, 3H), 7.35 (t, $J = 7.2$ Hz, 1H), 7.15 (d, $J = 8.0$ Hz, 1H), 6.90 (d, $J = 9.2$ Hz, 2H), 3.81 (s, 3H), 2.33 (s, 3H).

Compound 7 ($C_{17}H_{15}O_4N$): Orange yellow crystal; yield: 35 %; m.p. 160-161 °C; IR (KBr, $\nu_{\max}$, cm^{-1}): 3502, 3330, 3184, 3104, 3000, 1772, 1682, 1601, 1586, 1522, 1447, 1366, 1314, 1218, 1196, 1180, 820, 759, 698, 595; 1H NMR (400 MHz, $CDCl_3$) δ (ppm): 8.37 (s, 1H), 7.97 (d, $J = 8.8$ Hz, 2H), 7.82-

7.85 (m, 1H), 7.71 (d, $J = 8.4$ Hz, 2H), 7.51-7.56 (m, 1H), 7.34-7.38 (m, 1H), 7.17 (d, $J = 8.0$ Hz, 1H), 2.58 (s, 3H), 2.33 (s, 3H).

Assay of antifungal activity: The antifungal activity of the synthesized compounds against *Sclerotinia sclerotiorum*, *Helminthosporium maydis*, *Botrytis cinerea* and *Rhizoctonia solani* were determined using the plate growth rate method¹⁷.

The synthesized compounds and carbendazim (purity 90 %) were dissolved in dimethyl sulfoxide, respectively. The two solutions were diluted into five different concentrations with distilled water, respectively. They were added to the sterile culture medium (PDA) at 45 °C, mixed to homogeneity and transferred to sterile Petri dishes to solidify. A mycelium agar disc (5 mm in diameter) of the target fungi was placed in the center of PDA plates. They were incubated at 28 °C in the dark until the target fungi used as controls covered the surface of these plates. Control groups were treated with the corresponding solutions without the synthesized compounds or carbendazim. The experiment for each concentration was replicated three times. The diameter of the fungi in the cultures was measured and the inhibition of growth was calculated according to the formula of Abbott.

RESULTS AND DISCUSSION

All the synthesized derivatives of aspirin were screened for their activity against *Sclerotinia sclerotiorum*, *Helminthosporium maydis*, *Botrytis cinerea* and *Rhizoctonia solani*. The results are presented in Table-1. Most of them were active against the four different pathogenic fungi at a concentration of 100 mg L⁻¹. The inhibition rate of compounds **1** and **2** against *Helminthosporium maydis* reached 92.5 and 91.6 %, respectively, which was higher than carbendazim. The inhibition rate of compound **3** against *Botrytis cinerea* reached 90.6 %, which was close to carbendazim.

A series of derivatives of aspirin have been successfully synthesized and tested for their antifungal activity against *Sclerotinia sclerotiorum*, *Helminthosporium maydis*, *Botrytis cinerea* and *Rhizoctonia solani*. The preliminary results suggested that the design and synthesis of these compounds may be conducive to the antifungal activity of derivatives of aspirin. They are also promising and beneficial to further studies in developing new and more effective fungicides in the agricultural chemistry field. However, there is more work

TABLE-1
ANTIFUNGAL ACTIVITY OF COMPOUNDS 1-7 AGAINST *Sclerotinia sclerotiorum*,
Helminthosporium maydis, *Botrytis cinerea* AND *Rhizoctonia solani* AT 100 mg L⁻¹

Compound	R	Inhibition of growth (%) [*]			
		<i>Sclerotinia sclerotiorum</i>	<i>Helminthosporium maydis</i>	<i>Botrytis cinerea</i>	<i>Rhizoctonia solani</i>
1	H	87.5	92.5	72.0	88.7
2	4-Cl	93.7	91.6	88.8	63.8
3	4-Br	87.7	72.7	90.6	61.3
4	4-NO ₂	50.6	67.8	66.4	67.7
5	4-CH ₃	91.1	78.5	68.2	48.9
6	4-OCH ₃	91.1	70.4	60.8	46.2
7	4-COCH ₃	71.7	79.4	77.2	37.1
Carbendazim	—	100	87.5	91.3	100

^{*}Based on the mean of triplicates

to be done. A number of derivatives of aspirin should be further synthesized for screening and surveying quantitative structure-activity relationships so as to find novel fungicides with high effect and low toxicity. Meanwhile, the mode of action of compounds **1** and **2** and their safety to humans and non-target organisms also need to be investigated.

ACKNOWLEDGEMENTS

This project was supported by the Scientific Research Fund of Sichuan Provincial Education Department (No. 13ZA0206), the Key Scientific Research Fund of Xihua University (No. Z1013314), the Chengdu City Science and Technology Bureau (No. 12DXYB001JH), the Innovation Fund of Postgraduate of Xihua University (No. ycjj2014130) and the Research Center for Advanced Computation, Xihua University.

REFERENCES

1. X.J. Chen, M.J. Cao, Y. Wang, Y.H. Tong and J.Y. Xu, *Chin. J. Oil Crop Sci.*, **31**, 503 (2009).
2. Z.Q. Shi, M.G. Zhou and Z.Y. Ye, *J. Jiangsu Agric.*, **16**, 212 (2000).
3. W. Li, Y.J. Zhou and H.G. Chen, *Chin. J. Oil Crop Sci.*, **29**, 63 (2007).
4. K.Y. Liu and F.X. Chen, *Anhui Agric. Sci.*, **35**, 756 (2007).
5. Y.X. Qi, F.X. Chen and K.J. Ding, *Pesticides*, **8**, 567 (2006).
6. A. Have, R. Berloo, P. Lindhout and J.A.L. Kan, *Eur. J. Plant Pathol.*, **117**, 153 (2007).
7. S.I. Ignatova, N.S. Gorshkova and T.A. Tereshonkova, *Acta Physiol. Plant.*, **22**, 326 (2000).
8. H. Egashira, A. Kuwashima, H. Ishiguro, K. Fukushima, T. Kaya and S. Imanishi, *Acta Physiol. Plant.*, **22**, 324 (2000).
9. R.L. Guimaraes, R.T. Chetelat and H.U. Stotz, *Eur. J. Plant Pathol.*, **110**, 13 (2004).
10. R. Finkers, P. Berg, R. Berloo, A. Have, A.W. Heusden, J.A.L. Kan and P. Lindhout, *Theor. Appl. Genet.*, **114**, 585 (2007).
11. R. Finkers, A.W. Heusden, F. Meijer-Dekens, J.A.L. Kan, P. Maris and P. Lindhout, *Theor. Appl. Genet.*, **114**, 1071 (2007).
12. R. Finkers, Y. Bai, P. Berg, R. Berloo, F. Meijer-Dekens, A. Have, J. Kan, P. Lindhout and A.W. Heusden, *Euphytica*, **159**, 83 (2007).
13. D.W. Gilroy, *Prostag. Leukotr. Ess.*, **73**, 203 (2005).
14. D.M. McCarthy, *Best Pract. Res. Clin. Gastroenterol.*, **26**, 101 (2012).
15. R. Eccles, *Int. Pharm. J.*, **6**, 7 (1995).
16. C. Lampl, M. Voelker and H.C. Diener, *J. Neurol.*, **254**, 705 (2007).
17. W. Huang and G.F. Yang, *Bioorg. Med. Chem.*, **14**, 8280 (2006).