Extraction Optimization and Functional Properties of Protein from *Amygdalus pedunculatus* Seeds

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Amygdalus pedunculatus seeds contain good-quality proteins. In this paper, maximum yield (55.85 %) was obtained when temperature, extraction time and liquid/solid ratio were 39.49 °C, 80 min and 15.88:1 (v/w) by using response surface methodology. The amino acid composition of *Amygdalus pedunculatus* seed protein was analyzed and the proportion of the essential amino acids was closer to the FAO recommended values, among which, glutamic acid was in a particular high level (25.24 % of total amino acid content). The maximum and minimum protein solubility of *Amygdalus pedunculatus* seed protein were found at pH 10 and pH 4. Water and oil absorption capacity were 2.20 mL H₂O/g protein and 2.70 mL oil/g protein. Foaming capacity and emulsifying activity of *Amygdalus pedunculatus* seed protein were affected by pH (2-10). The least gelation concentration and surface hydrophobicity (*S*₀) were 12 % and 290, respectively. These results indicated that *Amygdalus pedunculatus* seed protein as a potential ingredient could be used in the food industry.

Key Words: *Amygdalus pedunculatus* seed, Response surface methodology, Protein, Functional property.

INTRODUCTION

The proteins from two major sources (animal proteins and plant proteins) are most widely used in the food and non-food markets, either as a general nutrients supply or as functional ingredients¹. However, proteins from animal sources (milk proteins) are expensive and low production. Demand of relatively inexpensive sources of proteins that can be incorporated to value-added food products is increasing². Worldwide, much of the research is going on various sources of plant proteins that may help in increasing the nutritional value of food products at low cost³⁻⁶.

Amygdalus pedunculatus (AP) is a rosaceous deciduous shrubs, low-input trees mainly grown in northwestern China and Mongolia. The leaves of this plant are spindle shape and have waxiness on its surface (Fig. 1). As a deciduous shrubs plant, it has a lot of rotted vegetation which can effectively improve the soil quality. The water and soil resources' protection and ecological balance around the desert areas were signally improved by the extensive root system of *Amygdalus pedunculatus*. Since the huge ability of preventing and controlling desert areas, *Amygdalus pedunculatus* has received considerable attention in recent years and has planted a lot in China. Meanwhile, a mass of *Amygdalus pedunculatus* seed (APS) were reaped every year and most of them were scrapped.

Fig. 1. A *Amygdalus pedunculatus* trees; B *Amygdalus pedunculatus* blooming; C *Amygdalus pedunculatus* fruiting; D *Amygdalus pedunculatus* seeds

This would cause the waste of resource. However, our research has found that *Amygdalus pedunculatus* seed was rich in protein and at present no scientific information is available on the

protein from *Amygdalus pedunculatus* seed. Thus, it was deserved us to do further research. As known to all, to develop dietary protein for utilization as ingredients in the food industry, it is necessary to determine the physicochemical and functional properties of the extracted proteins⁷.

The objective of the present study was to evaluate the extraction condition, physicochemical and functional properties of *Amygdalus pedunculatus* seed protein. The amino acid composition, water/oil absorption, protein solubility, foaming and emulsifying properties, gelling properties and surface hydrophobicity, were reported to evaluate the quality of extracted protein isolates. The basic information about the physicochemical properties of isolates that would help to determine their application in foods was provided⁸.

EXPERIMENTAL

Sample preparation: *Amygdalus pedunculatus* seed were dehulled to get kernels and then smashed by a high speed grinder (WND-200, Wei neng da, China). The ground kernels lipids were removed by cold squeeze method and extracted two times with ethanol ratio (flour ethanol of 1:5 w/v) to remove residual lipids. The organic solvent was recycled by a rotary evaporator (RE-5205, Ya rong, China). The deoiled meal samples were grounded again and sieved to an uniform particle size. The samples were stored at - 20 °C till use. All other materials and reagents were of analytical grade and purchased from regular suppliers. The experiment was carried out in triplicate.

Proximate analysis: Association of Official Analytical Chemists AOAC (1997) methods were used to determine moisture, fat, protein, ash and fiber content.

Extraction of protein using response surface methodology (RSM): The defatted *Amygdalus pedunculatus* seeds (DAPS) were extracted with distilled water at varying temperature (x_1), reaction time (x_2) and liquid: solid ratio (x_3) by using response surface methodology (Table-1). According to the experimental data and Design-Expert 7.1.6 software analysis, the optimum protein extraction condition could be selected⁹⁻¹¹. The suspension was centrifuged at 10,000g for 20 min and the supernatants were collected. The supernatants were adjusted to pH 4 (*Amygdalus pedunculatus* seed protein' isoelectric pH) and centrifuged at 10,000 g for 10 min. Then collected precipitation and freeze drying. And protein samples were stored at 4 °C until used for the determination of physicochemical and functional properties.

TABLE-1
INDEPENDENT VARIABLE VALUES OF THE
PROCESS AND CORRESPONDING LEVELS

Independent variables	Symbol	Levels		
		-1	0	1
Temperature (°C)	x_1	30	40	50
Extraction time (min)	x_2	70	90	110
Liquid: solid ratio (v/w)	x_3	10:1	15:1	20:1

Amino acid analysis: Amino acid profiles were determined at the Pony Testing International Group chemical labs using AOAC Official Method 994.12.

Water/oil absorption: The water or oil absorption was determined by using a modified method described by Cao

*et al.*¹². Samples (1 g) were mixed with distilled water or peanut oil (10 mL) and the mixtures were centrifuged at 3000 g for 0.5 h. Then the remaining water/oil's volume was measured and the water/oil capability could be calculated. The gain in volume per unit weight was recorded as water or oil absorption capacity.

Protein solubility (PS): Protein solubility was studied using a modified method of Shand *et al.*¹³. Every 20 mL distilled water were mixed with each samples (0.20 g) and adjusted to pH (2-10) by using 1 mol/L HCl or 1 mol/L NaOH. Samples were stirred in an orbital shaker at ambient temperature for 0.5 h and then centrifuged at 3000 g 10 min. The protein content of supernatants was tested by the method of Kjeldahl and the protein solubility could be calculated.

Foaming properties (foaming capacity and stability): The foaming capacity (FC) and foaming stability (FS) of the protein were evaluated according to the modified method of Lin *et al.*¹⁴. A sample (1 g) was mixed with 100 mL distilled water and the mixture was homogenized for 1 min by a Waring Blender (ds-1, Biao ben, China) at a certain speed (approximately 10,000 rpm). The mixture was transferred into a 250 mL graduated cylinder, so as to measure the foam volume. The foaming capacity was described as the increased percentage of volume at pH 7. Foaming stability was determined by measuring the decrease in volume of foam after 0.5 h at pH 7.

Emulsifying properties (emulsifying activity and stability): The emulsifying activity (EA) and emulsifying stability (ES) were determined by using a modified method of Naczek *et al.*¹⁵. Distilled water (50 mL) was mixed with samples (2.00 g) and the mixture was homogenized for 1 min by using a Waring Blender at a certain speed (10,000 rpm). An equal quantity of peanut oil was added and mixtures were homogenized at another certain speed (12,000 rpm) for 1 min again. The emulsion was divided evenly into 50 mL centrifuge tubes and centrifuged at 3000 g for 5 min. Emulsifying activity could be deduced by the ratio of the height of the emulsified layer to the liquid layer at pH 7. Then the emulsion was heated 15 min at 85 °C and cooled 15 min. The emulsifying stability was expressed as the percentage of emulsifying activity remaining after heating.

Gelling properties: The gelling properties of *Amygdalus pedunculatus* seed protein was described according to the method of Sathe and Salunkhe with some modification¹⁶. Different amounts of protein were weighed into 50 mL test tubes which contained 20 mL distilled water and made protein suspensions from 2-16 %. The pH was adjusted to 7. Test tubes were heated in water bath at 85 °C for 20 min and cooled immediately. Then cooled at 4 °C overnight. The least gelation concentration (LGC) was denoted as the concentration when the sample did not skid along the test tube walls in inverted position¹⁷. The results were expressed as liquefied (-), gluey ($\pm$) and gel (+)⁸.

Surface hydrophobicity (S_0): Surface hydrophobicity of protein was determined by using a hydrophobic fluorescence probe, 1-anilino-8-naphthalene sulfonate (ANS), with the help of a spectrofluorometer (RF-5301PC, Shimadzu, Japan). *Amygdalus pedunculatus* seed protein solutions of 0.15-0.30 mg/mL were prepared using 0.01 M phosphate buffer (pH 7)

by serial dilution. Protein solutions of 2.0 mL were mixture with ten microliters of 8.0 mM 1-anilino-8-naphthalene sulfonate solution (prepared in 0.01 M phosphate buffer, pH 7). The fluorescence intensity was measured at wavelengths of 390 nm (excitation) and 470 nm (emission) by a spectrofluorometer. According to the plotted slope of fluorescence intensity against protein concentration, surface hydrophobicity was calculated by linear regression¹⁸.

Statistical analysis: The Design-Expert 7.1.6 software set up the design of response surface methodology and analysis of the maximum protein extraction under the optimum conditions. All trials were carried out in triplicate and all data were reported as means $\pm$ SD (standard deviation). Treatment differences were evaluated at the 95 % confidence level with three treatment replicates.

RESULTS AND DISCUSSION

Proximate composition of *Amygdalus pedunculatus* seed: The composition of *Amygdalus pedunculatus* seed was as follows (% w/w): protein, 28.6; fat, 43.2; moisture, 6.0; ash, 2.5; fiber, 15.2. *Amygdalus pedunculatus* seed was rich in protein when compared favorably with other seeds as a potential non-conventional source of protein such as green lentils (23.0 g/100 g)¹⁹, paprika seed (24.43 g/100 g)²⁰, red lentils (25.9 g/100 g)¹⁹, desi chickpea (20.5 g/100 g)¹⁹, yellow pea (21.09 g/100 g)¹⁹, peanut (22.04 g/100 g)²¹ and kabuli chickpea (16.7 g/100 g)¹⁹. In brief, *Amygdalus pedunculatus* seed was a kind of potential plant protein resource and very worth studying.

Optimization for protein extraction using response surface methodology: Response surface methodology was selected to investigate optimum conditions for maximizing the yield of protein extraction from *Amygdalus pedunculatus* seed. The influences of temperature, extraction time and liquid: solid ratio on the experimental and predicted yield values of the protein extraction were depicted in Table-2. The coefficient of determination (R^2) was 86 % and the standard error was 0.927 which indicates the adequacy of applied model. The response surfaces based on these coefficients with one variable was at the optimum level and varying the other two within the

experimental range were shown in Fig. 2. In general, the exploration of response surfaces demonstrated a complex interaction between the variables. Optimum protein extraction was obtained at 39.49 °C, 80 min extraction time and 15.88:1 (v/w) liquid: solid ratio. Under this condition, the protein yield of theoretical value was 57.30 %, the actual protein yield could reach 55.85 % and protein content is 94.8 %. It was observed that experimental optimal value was slightly lower than predicted value by the regression model. It confirmed that these conditions were optimal for *Amygdalus pedunculatus* seed protein extraction. The protein was extracted using the optimal condition and then analysed its amino acid composition.

Amino acid composition: The amino acid composition of *Amygdalus pedunculatus* seed protein has been quantified and the results were shown in Table-3. Amino acid composition of *Amygdalus pedunculatus* seed protein was partially conformed to the FAO requirements for the amino acids (FAO/WHO/ONU, 1985) except for lysine, histidine and methionine + cysteine, while SPI was also partially satisfied with the standard. However, *Amygdalus pedunculatus* seed protein could be considered as a high quality protein which contains 18 kinds of amino acids. The total amino acid contents of *Amygdalus pedunculatus* seed protein was 94.75 %, slightly higher in comparison with SPI (91.69 %) and tea protein (77.31 %)²². Glutamic acid was the most abundant amino acid in *Amygdalus pedunculatus* seed protein, followed by aspartic acid, arginine, leucine, phenylalanine and glycine. The least abundant amino acid in *Amygdalus pedunculatus* seed protein was methionine, which only accounts for 0.8 %. The glutamic acid was widely used as pharmaceuticals, liver function promoting agents and fatigue recovery agents. Aspartic acid could promote the red blood cells grow and improve the nutrition of brain cells. It can be seen that *Amygdalus pedunculatus* seed protein was full of nutrients and deserved to do functional property research.

Absorption properties: Protein structure has both hydrophilic and hydrophobic properties and thereby interacts with water/oil in the food system²³. Under limited water/oil conditions, the water and oil absorption capacity symbolizes the ability of binding water/oil molecules. The water and oil's

TABLE-2
BOX-BEHNKEN DESIGN ARRANGEMENT, RESPONSES AND PREDICTED VALUES FOR PROTEIN YIELD

Run	Coded variables			Uncoded variables			Protein yield (%)	
	x_1	x_2	x_3	x_1	x_2	x_3	Experimental	Predicted
1	-1	0	1	30	60	20	51.84	51.19
2	1	-1	0	50	40	15	54.82	55.25
3	-1	0	-1	30	60	10	45.59	55.48
4	-1	1	0	30	80	15	55.91	55.28
5	0	0	0	40	60	15	54.44	47.49
6	0	0	0	40	60	15	56.64	50.32
7	0	1	-1	40	80	10	53.48	51.78
8	0	0	0	40	60	15	57.11	52.81
9	0	0	0	40	60	15	56.99	48.11
10	1	0	-1	50	60	10	50.26	52.01
11	1	1	0	50	80	15	53.86	53.24
12	0	-1	-1	40	40	10	49.59	54.66
13	0	1	1	40	80	20	54.17	56.26
14	1	0	1	50	60	20	54.71	56.26
15	0	0	0	40	60	15	56.11	56.26
16	-1	-1	-1	30	40	15	52.61	56.26
17	0	-1	1	40	40	20	51.76	56.26

Fig. 2. 3D graphic surface optimization of protein yield versus: A solvent:solid ratio and extraction time; B temperature and solvent:solid ratio; C temperature and extraction time

TABLE-3
AMINO ACID COMPOSITION^a OF *Amygdalus pedunculatus* SEED PROTEIN (APSP) AND SPI²² (g/100 g PROTEIN)

Amino acid	FAO ^b	APSP	SPI
Aspartic acid	—	11.61 ± 0.55	9.09 ± 0.16
Glutamic acid	1.9	25.24 ± 1.02	3.08 ± 0.03
Serine	—	3.61 ± 0.06	4.17 ± 0.08
Histidine	3.4	2.21 ± 0.21	5.30 ± 0.17
Glycine	—	4.60 ± 0.08	15.7 ± 0.18
Threonine	—	2.00 ± 0.10	1.44 ± 0.05
Arginine	—	10.69 ± 0.16	5.84 ± 0.28
Alanine	—	4.37 ± 0.07	2.89 ± 0.07
Proline	—	4.21 ± 0.10	5.86 ± 0.15
Tyrosine	6.3 ^c	2.91 ± 0.09	5.53 ± 0.16
Valine	3.5	4.54 ± 0.13	3.52 ± 0.08
Methionine	2.5 ^d	0.27 ± 0.02	2.42 ± 0.13
Cysteine	—	0.82 ± 0.05	1.15 ± 0.01
Isoleucine	2.8	3.55 ± 0.06	3.54 ± 0.15
Leucine	6.6	6.71 ± 0.14	5.77 ± 0.13
Phenylalanine	—	5.06 ± 0.11	4.69 ± 0.07
Lysine	5.8	1.26 ± 0.03	11.20 ± 0.19
Tryptophan	—	1.09 ± 0.02	0.50 ± 0.02
Total	—	94.75 ± 0.99	91.69 ± 1.00

^aData are the mean ± SD of three analyses. ^bFAO/WHO/ONU. Energy and protein requirement, 1985. ^cTyrosine + phenylalanine. ^dMethionine + cysteine.

mean values of absorption capacities of *Amygdalus pedunculatus* seed protein were shown in Table-4. In this experiment, the water absorption capacity was 2.20 mL/g, which was lower than that reported for casein (2.48 mL/g)² and unstabilized rice bran protein isolates (Un-PI) (3.80 mL/g)⁸. And it was higher than white rice protein (1.78 mL/g)¹². Aletor *et al.*²⁴ considered that water absorption capacity values ranging from 1.49 to 4.72 g/g could be used in viscous foods. The data indicated that *Amygdalus pedunculatus* seed protein had good capacity for water absorption and could be used in products which was required high water retention. The oil absorption of *Amygdalus pedunculatus* seed protein was 2.70 mL/g, which is higher than white rice protein (2.56 mL/g)¹², casein (2.15 mL/g)² and Un-PI (2.40 mL/g)⁸. High oil absorption is essential in the formulation of many processed foods and then improves mouthfeel and flavor retention of the final product.

Protein solubility (PS): In general, superior functional attributes for most applications in food processing were associated with the solubility of proteins. It was depicted in Fig. 3 that *Amygdalus pedunculatus* seed protein exhibited pH dependent protein solubility. The solubility of *Amygdalus pedunculatus* seed protein was the minimum at pH 4, which might be due to the isoelectric region. Under isoelectric pH, the electrostatic repulsion and ionic hydration were minimum and hydrophobic interactions between surface non-polar patches were maximum²⁵. The protein solubility increased on either side of pH 4 including acidic and alkaline. A moderate increase was observed above pH 4 until pH 8, followed by a marked increase up to pH 10. At lower pH values, the increased net positive charge contribute to the solubility⁶. At higher pH values, the solubility increased may be due to the increased net negative charge on the protein dissociates the protein aggregates²⁶. The total contents of negatively charged amino acids (aspartic acid and glutamic acid) of *Amygdalus pedunculatus* seed protein were 36.85 %. The residues of these

TABLE-4
PHYSICO-CHEMICAL^a PROPERTIES OF *Amygdalus pedunculatus* SEED PROTEIN

Index	<i>Amygdalus pedunculatus</i> seed protein									
Water absorption (mL/g)	2.20 ± 0.16									
Oil absorption (mL/g)	2.70 ± 0.21									
Surface hydrophobicity (S ₀)	290 ± 8									
Gelation										
Concentration	2 %	4 %	6 %	8 %	10 %	12 %	14 %	16 %	18 %	
	(-)	(-)	(-)	(-)	(±)	(+)	(+)	(+)	(+)	
Least gelation concentration	12									

^aGelation levels: (-) liquefied, (±) gluey and (+) gel. The values are the mean of three replicates.

Fig. 3. Protein solubility of *Amygdalus pedunculatus* seed protein. Data are shown as means. Error bars represent the SD

ionic amino acids might be at least partially responsible for the protein solubility at high pH levels.

Foaming properties: The foaming capacity of *Amygdalus pedunculatus* seed protein (Fig. 4A) was pH-dependent. At pH 4 the lowest foaming capacity (9 %) which was due to the protein behaviour at its isoelectric point. On either side of pH 4, foaming capacity significantly increased, especially at 10. It was probably due to the net charges' increase on the protein surface, which weakened the hydrophobic interactions but increased the flexibility of the protein. This allowed the protein to diffuse more rapidly to the air-water interface to encapsulate air particles and then enhance the foam formation²⁶. The effect of pH on foaming stability of *Amygdalus pedunculatus* seed protein was shown in Fig. 4B. It was found that *Amygdalus pedunculatus* seed protein had a minimum foaming stability (50 %) at pH 4 with an increase on both sides of pH 4. Foaming stability was dependent on the formation of a thick cohesive layer around the air bubble²⁵.

Emulsifying properties: Emulsifying activity and emulsifying stability were used to depicted the emulsifying properties of food proteins. The effect of pH on emulsifying activity and emulsifying stability of *Amygdalus pedunculatus* seed protein were shown in Fig. 5. Results showed that *Amygdalus pedunculatus* seed protein had a minimum emulsifying activity (40.2 %) at pH 4 (Fig. 5A) with an increase on both sides of pH 4. Emulsifying activity was pH-dependent and alkaline pH improved the emulsifying activity more than did the acidic pH²⁷. Similar observations on the pH dependence of emulsi-

Fig. 4. Effects of pH on foaming capacity and foaming stability of *Amygdalus pedunculatus* seed protein. Data are shown as means. Error bars represent the SD

fying activity have been reported²⁸. Nakai indicated that protein solubility and its surface hydrophobicity were very important properties for determining protein emulsifying activity²⁸. Moreover, the relationship between emulsifying activity against pH for *Amygdalus pedunculatus* seed protein was more or less similar to that between protein solubility against pH. This was in agreement with the general correlation between emulsifying activity and protein solubility found in previous studies^{29,30}. emulsifying stability was also pH-dependent (Fig. 5B). Hung and Zayas³⁰ suggested that various factors,

Fig. 5. Effects of pH on emulsifying activity and emulsifying stability of *Amygdalus pedunculatus* seed protein. Data are shown as means. Error bars represent the SD

including pH, net charge, heat treatment and protein conformation, could affect the values of emulsifying stability. The high emulsifying stability of protein isolates might be due to the partial unfolding of protein structure, by exposing hydrophobic units and facilitating protein interaction with non-polar solvents and resisting oil drop flocculation, thereby increasing the overall stability of emulsion⁸. Present study of emulsifying activity and emulsifying stability were quite similar to those reported earlier by Jiamyangyuen *et al.*³¹.

Least gelation concentration: It is an important functional property that the proteins' ability to form gels in food processing and formulation. The gel is resistant to flow under pressure when proteins form a three-dimensional network, in this case gelation occurs. The least gelation concentration is often used as an indication of the gelation capacity of food proteins¹⁹. The gelling properties of *Amygdalus pedunculatus* seed protein was summarized in Table-4. There was no gel formed at a concentration of 2, 4, 6 and 8 % (w/v). A weak gel was formed at 10 % concentration of *Amygdalus pedunculatus* seed protein. At 12 % (least gelation concentration), a strong gel was formed. *Amygdalus pedunculatus* seed protein had the

best gelling properties compared with the rice bran protein⁸ yellow pea protein and kabuli chickpea protein (least gelation concentration of 14 %, w/v)¹⁹. The results might be due to the denaturation of protein and the reinforcement of gel strength that made lower least gelation concentration values of *Amygdalus pedunculatus* seed protein which had good gelation characteristics. *Amygdalus pedunculatus* seed protein would be a useful additive and it could deliver the desired gelling property in thick textured foods.

Surface hydrophobicity (So): Surface hydrophobicity is affected by the presence of hydrophobic patches on the surface of proteins that are available to interact with the food system, especially molecules in polar aqueous environments⁸. The surface hydrophobicity value of *Amygdalus pedunculatus* seed protein was presented in Table-4. Surface hydrophobicity of *Amygdalus pedunculatus* seed protein (290) was significantly higher than that of parboiled rice bran protein isolates (29)⁸, but lower than SPI (399)²². This demonstrated that *Amygdalus pedunculatus* seed protein had more hydrophobic grouping on the surface of protein. The functional property of a protein might be impacted by higher surface hydrophobicity, especially in foam and emulsion where this property is needed for a specific food product application³².

Conclusion

The results of the present study provided significant information on the physicochemical and functional aspects of protein extracted from *Amygdalus pedunculatus* seed. Response surface methodology results gave the optimum value of temperature, extraction time and liquid: solid ratio at 39.49 °C, 80 min and 15.88:1, respectively. The amino acid composition of the *Amygdalus pedunculatus* seed protein was also determined that the high glutamic acid content (25.24 %) and total amino acids (94.75 %) were obtained. Finally, *Amygdalus pedunculatus* seed protein demonstrated lower protein solubility, but higher oil absorption capacity and surface hydrophobicity *etc.* The data obtained from this study could provide basic information for the food application of *Amygdalus pedunculatus* seed protein which has not been explored earlier for its functional property, such as it can be used for sausages, puddings, baked food and creams.

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REFERENCES

1. L.Q. Shen, X.Y. Wang, Z.Y. Wang, Y.F. Wu and J.S. Chen, *Food Chem.*, **107**, 929 (2008).
2. G.K. Chandi and D.S. Sogi, *J. Food Eng.*, **79**, 592 (2007).
3. S. Gorinstein, E. Pawelzik, E.D. Licon, R. Haruenkit, M. Weisz and S. Trakhtenberg, *J. Sci. Food Agric.*, **82**, 886 (2002).
4. A. Rangel, G.B. Domont, C. Pedrosa and S.T. Ferriera, *J. Agric. Food Chem.*, **51**, 5792 (2003).
5. D.S. Sogi, S.K. Garg and A.S. Bawa, *J. Food Sci.*, **67**, 2997 (2002).
6. H. Tomotake, I. Shimaoka, J. Kayashita, M. Nakajoh and N. Kato, *J. Agric. Food Chem.*, **50**, 2125 (2002).

7. M. Wang, N.S. Hettiarachchy, M. Qi, W. Burks and T. Siebenmorgen, *J. Agric. Food Chem.*, **47**, 411 (1999).
8. S.H. Khan, M.S. Butt, M.K. Sharif, A. Sameen, S. Mumtaz and M.T. Sultan, *J. Agric. Food Chem.*, **59**, 2416 (2011).
9. C. Cardoso, B. Ribeiro and R. Mendes, *Eur. Food Res. Technol.*, **234**, 935 (2012).
10. J. Zheng, H.T. Wu, B.W. Zhu, X.P. Dong, M.M. Zhang and Y.L. Li, *Eur. Food Res. Technol.*, **234**, 895 (2012).
11. M.M.M. Oliveira, D.F. Brugnera, J.A. Nascimento, N.N. Batista and R.H. Piccoli, *Eur. Food Res. Technol.*, **234**, 821 (2012).
12. X.H. Cao, H.B. Wen, C.J. Li and Z.X. Gu, *J. Cereal Sci.*, **50**, 184 (2009).
13. P.J. Shand, H. Ya, Z. Pietrasik and P.K.J.P.D. Wanasundara, *Food Chem.*, **83**, 1119 (2007).
14. M.J. Lin, E.S. Humbert and F.W. Sosulski, *J. Food Sci.*, **39**, 368 (1974).
15. M. Naczki, L.L. Diosady and L.J. Rubin, *J. Food Sci.*, **50**, 1685 (1985).
16. S.K. Sathe and D.K. Salunkhe, *J. Food Sci.*, **46**, 71 (1981).
17. K.O. Adebawale and O.S. Lawal, *Food Chem.*, **83**, 237 (2003).
18. U. Kalapathy, N.S. Hettiarachchy and K.C. Rhee, *J. Am. Oil. Chem. Soc.*, **74**, 195 (2006).
19. J.I. Boye, S. Aksay, S. Roufik, S. Ribereau, M. Mondor, E. Farnworth and S.H. Rajamohamed, *Food Res. Int.*, **43**, 537 (2010).
20. T.A. El-Adawy and K.M. Taha, *J. Agric. Food Chem.*, **4**, 1253 (2001).
21. I.Y.S. Rustom, M.H. Lopez-Leiva and B.M. Nair, *J. Food Sci.*, **56**, 1660 (1991).
22. R. Horax, N. Hettiarachchy, A. Kannan and P.Y. Chen, *Food Chem.*, **124**, 545 (2011).
23. J.Y. Cheng, S.H. Zhou, D. Wu, J.C. Chen, D.H. Liu and X.Q. Ye, *Food Chem.*, **112**, 469 (2009).
24. O. Aletor, A.A. Oshodi and K. Ipinmoroti, *Food Chem.*, **78**, 63 (2002).
25. S. Damodaran, In ed.: S. Damodaran, *Food Proteins and their Applications*, Institute National de la Recherche Agronomique Centre de Recherches de Tours Nouzilly, France; Marcel Dekker, Inc: New York (1997).
26. R.E. Aluko and R.Y. Yada, *Food Chem.*, **53**, 259 (1995).
27. E.K. Khalid, E.E. Babiker and A.H. El-Tinay, *Food Chem.*, **82**, 361 (2003).
28. S. Nakai, *J. Agric. Food Chem.*, **31**, 676 (1983).
29. D.D. Crenwelge, C.W. Dill, P.T. Tybor and W.A.A. Landmann, *J. Food Sci.*, **39**, 175 (1974).
30. S.C. Hung and J.F. Zayas, *J. Food Sci.*, **56**, 12163 (1991).
31. S. Jiamyangyuen, W.J. Harper, V. Srijsdaruk and K. Kumthonglang, *Milchwissenschaft*, **60**, 192 (2005).
32. S.H. Guzmán-Maldonado and O. Paredes-López, In ed.: O. Paredes-López, *Molecular Biotechnology for Plant Food Production*, Technomic Publishing, Lancaster (1999).