Antimicrobial and Antiinflammatory Activity of the Hydrogels Containing Rutin Delivery

HIMESH SONI^{1,*}, JITENDER MALIK², A.K. SINGHAI² and SARVESH SHARMA²

¹Suresh Gyan Vihar University, Jaipur-302 025, India

²Lakshmi Narain College of Pharmacy, Raisen Road, Bhopal-462 021, India

*Corresponding author: E-mail: himeshsoni@rediffmail.com

(Received: 31 October 2012;

Accepted: 21 August 2013)

AJC-13953

Flavonoids are principal active constituents have been used to treatment of various human diseases. Rutin (quercetin-3-rhamnosyl glucoside) as the flavonoids display anticancer, antiviral, antiinflammatory and heart disease protective activities. Rutin by acting as antioxidants exhibited several beneficial effects, such as antiinflammatory, antiallergic, antiviral as well as an anticancer activity. The objective of present study was to formulate hydrogels containing rutin at various concentrations *i.e.*, 0.020, 0.025 and 0.030 % (w/w). Hydrogels were prepared using Carbopol 934 and a methanolic (co-solvent) dispersion of rutin. The hydrogels were subjected to antimicrobial and antiinflammatory activity. The zone of inhibition of rutin in hydrogel systems was evaluated by cup plate method against bacteria *i.e.*, *Staphylococcus aureus*, *Staphylococcus glurance* and *Eschericia coli*. The antiinflammatory activity of the hydrogel systems was also carried out by Carrageenan-induced rat paw edema model. Results of *in vivo* activity led to the conclusion that the hydrogel H2 formulation showed predominantly significant activity, which is comparable to the standard drug.

Key Words: Rutin, *Staphylococcus aureus*, *Staphylococcus glurance*, *Eschericia coli* and hydrogel.

INTRODUCTION

The hydrogel can be defined as a cross-linked polymeric network which has the capacity to hold water within its porous structure. The water holding capacity of the hydrogels arise mainly due to the presence of hydrophilic groups, *viz.* amino, carboxyl and hydroxyl groups, in the polymer chains¹. The hydrogel is a permanent or chemical gel stabilized by covalently cross-linked networks. These chemical hydrogels may be prepared either by cross linking water-soluble polymers or by converting hydrophobic polymers into hydrophilic polymers that are then cross-linked to form a network. With such a structure, hydrogels are able to swell, absorbing a large amount of water without the polymer dissolving, which gives them characteristics similar to those of soft tissue². Therefore, hydrophilic gels called hydrogels are cross-linked materials absorbing large quantities of water without dissolving. Softness, smartness and the capacity to store water make hydrogels unique materials³. Hydrogels can be designed to change properties (*e.g.* swelling/collapse or solution-to-gel transitions) in response to externally applied triggers, such as temperature, ionic strength, solvent polarity, electric/magnetic field, light, or small (bio) molecules⁴. Rutin is a flavonoid present in the plant kingdom as *allelopathic* substances. Rutin (Fig. 1) is the rhamnoglucoside of the flavonoid quercetin and found in many plants and used for treatment of various diseases related to the

vascular⁵. It is quercetin-3-rutinoside or 3,3',4', 5,7-penta-hydroxy flavones-3-rutinoside and has a chemical formula C₂₇H₃₀O₁₆.

Fig. 1. Structure of rutin

It has been reported that phenolic compounds show antimicrobial activity against a wide range of microorganisms⁶. They also possess significant antiinflammatory activity by virtue of the presence of the free phenolic groups. We already reported the formulation and characterization of the hydrogels containing rutin. An acute toxicity study in rabbits was reported by Lima *et al.*⁷ showing the innocuity of rutin after

daily oral doses of 0.01 mol kg⁻¹, through the evaluation of blood markers of toxicity. Furthermore, a critical review published in 2007 concluded that quercetin, the aglicone of rutin, does not produce adverse health effect taking into account an estimated level of dietary intake⁸. Recently, it has been demonstrated that rutin does not have genotoxic effects on HCT cells⁹. Taking all these aspects, the objective of this study was to develop a hydrogels containing rutin in varying concentration and to investigate the antiinflammatory and antimicrobial potential.

EXPERIMENTAL

Rutin was isolated from *Annona squamosa* leaves. It is identified and characterized by various analytical techniques like HPLC, FT-IR. Carbopol 934 and HPMC were purchased from Hi-media laboratories Pvt. Ltd. All the other chemicals and reagents used were of analytical grade.

Nutrient broth (NB), Nutrient agar (NA), Muller-Hinton agar, Peptone water and antibiotics gentamycin were procured from Hi-media laboratories, Mumbai, India.

Test organisms: The test organisms *Staphylococcus aureus* (MTCC 265), *Staphylococcus glurance* (MTCC 265), *Eschericia coli* (MTCC 167), were procured from Microbial Tissue Culture technology Centre, lab. Chandigarh, India.

Preparation of hydrogel systems: In separate container, the hydrogel forming polymers were dissolved in small amount of double distilled water in various proportions as shown in Table-1 and then remaining ingredients *i.e.*, glycerine and sodium benzoate were added. Now, methanolic dispersion of rutin (1 mg/mL) was added to it and make up the volume up to 100 mL. Then, sonicated (Lark probe sonicator) at 60 frequency, 20 s at 28 °C. The above formulation was allowed to stand for 24 h at room temperature. The pH of this gel preparation was maintained 6 ± 0.4 and stored in well closed container.

TABLE-1
COMPOSITION OF HYDROGEL FORMULATIONS

Components	Quantity		
	H1 0.020 % (w/w)	H2 0.025 % (w/w)	H3 0.030 % (w/w)
Carbopol 934	500 mg	500 mg	500 mg
HPMC	500 mg	500 mg	500 mg
Acaica	500 mg	500 mg	500 mg
Glycerine	2 mL	2 mL	2 mL
Sodium benzoate	100 mg	100 mg	100 mg
Rutin	3.2 mg	4 mg	4.8 mg

Antimicrobial activity by disc diffusion method

Preparation of inoculum: *E.coli*, *S. glurance* and *S.aureus* strains were used. 50 mL of nutrient broth was prepared in 100 mL conical flask. It was sterilized and then inoculated with inoculum with the help of sterile loop in laminar air flow from preserved slants¹⁰. They were then kept in incubator at 37 °C for sufficient period of time for organism to grow.

Preparation of media: 200 mL of nutrient agar media was prepared and the pH was maintained at 7.0-7.2.

Pour plate technique: 1 mL of prepared inoculums was poured in sterile Petri dish and then poured 15 mL of nutrient agar media in it and allowed to solidify.

Disc diffusion method: After solidification the disc of whatman filter paper imbibed with 20 µL of sample was carefully placed with the help of forceps at the center of the Petri dish of different doses and then kept in incubator for 24 h¹¹.

Standard: Gentamycin (50 µg/disc), was taken as the positive control.

Measurment of zones: The zone of inhibition was measured with the help of zone reader.

Antiinflammatory activity: Animals were divided into six groups with three animals in each; the first three groups served as controls: (a) un-inflamed, injected with saline; (b) inflamed, injected with carrageenan; (c) inflamed, treated with the reference product (oleogel diclofenac sodium). The other three groups were inflamed and treated with the hydrogel systems H1, H2 and H3. Carrageenan solution (1 % w/v, in normal saline) was used to induce inflammation. The animals were placed singly in observation chambers for 10 min to minimize any stress-related behavioural changes. The mice then received sub-plantar administration of 50 µL of the carrageenan solution in the left hind paw and were returned immediately to the observation chamber. The thickness (mm) of the paw was measured at 0, 1, 2, 3 and 4 h after carrageenan administration, using an electronic digital Vernier caliper¹². The hydrogel formulations and the standard were applied to the plantar surface of the left hind paw by gently rubbing 0.5 g of the formulation 50 times with the index finger. After 0.5 h, pleurisy was induced by injecting 50 µL of 1 % w/v carrageenan solution subcutaneously into the sub-plantar surface of the left paw of the mice. Control un-inflamed animals received 50 µL normal saline subcutaneously into the sub-plantar surface of the left paw.

$$\text{Inhibition of edema (\%)} = \left(1 - \frac{V_t}{V_c}\right) \times 100$$

where, V_t = volume of edema in test, V_c = volume of edema in control.

RESULTS AND DISCUSSION

The formulated hydrogels of different concentration of rutin were evaluated for antimicrobial and antiinflammatory activity. Different composition of hydrogel were tabulated in Table-1. The zone of inhibition for three different strain of bacteria *i.e.*, *S. aureus*, *S. glurance* and *E. coli* for H2 formulation was found to be 3, 9 and 5 cm, respectively (Table-2 and Fig. 2). Our other two H1 and H3 formulation also show moderate inhibition for bacterial strain. The zone of inhibition of H2 formulation signifies that minimum concentration available of drug is sufficient to prevent the growth of microbe. Antimicrobial of plant origin is effective in the treatment of several infections and important for the antimicrobial activity. The action of compounds containing phenolic (flavonoid) groups may be related to inhibition of hydrolytic enzyme or other interactions to inactivate microbial adhesions, on specific transport of carbohydrates *etc.* Phenolic compounds exhibit a wide range of antiallergenic, antiinflammatory, antimicrobial,

TABLE-2
ANTIMICROBIAL EFFECT OF HYDROGEL
SYSTEMS DRUGS BY CUP PLATE METHOD

Bacterial strains	Zone of inhibition (diameter in mm)				
	Ctl.	Std.	Formulation		
			H1	H2	H3
<i>E. coli</i>	–	22	01	05	07
<i>S. aureus</i>	–	30	Nil	03	06
<i>S. glurence</i>	–	17	Nil	09	12

Fig. 2. Observation of zone of inhibition of H2 formulation

antioxidant, antithrombotic, cardio protective and vasodilator effects. Rutin has shown significant anti microbial activity against both the gram positive and gram negative bacteria. The antiinflammatory activity of different hydrogel formulation was carried out with the standard Oleogel. The formulated hydrogels H2 decreased the paw oedema volume by 26.07 % within 4 h after administration, while standard drug decreased the paw edema volume by 41.44 % (Table-3, Fig. 3) when compared with the paw edema volume of the control.

TABLE-3
ANTIINFLAMMATORY EFFECT IN
TERM OF % INHIBITION OF EDEMA

Time (h)	H1	H2	H3	Oleogel (standard)
1	68.67	71.78	72.04	80.11
2	69.05	72.35	73.97	87.54
3	34.87	37.49	39.01	48.37
4	24.56	26.07	29.63	41.44
5	17.91	19.96	21.04	28.34

Fig. 3. Average % edema inhibition by different hydrogel formulation

Conclusion

Flavonoid is major phenolic compounds are becoming the major subject of medical research. They have been reported to possess many useful properties, including oestrogenic activity, antiinflammatory activity, enzyme inhibition, antimicrobial activity. For centuries, preparations that contain flavonoids as the principal physiologically active constituents have been used by physicians and lay healers in attempts to treat human diseases^{13,14}. Hydrogels have been used to deliver active component. Instead of conventional creams, the hydrogels have been formulated for better patient compliance. These hydrogels have moisturizing properties therefore scaling and dryness is not expected with this drug delivery system¹⁵. Many antifungal formulations have been developed as hydrogel formulation. It has shown better absorption than conventional cream formulations¹⁶. The present study revealed that the hydrogel H2 formulation showed predominantly significant activity, which is comparable to H1 and H3. Since H2 contained 0.025 % w/w amount of rutin which evoked antimicrobial and antiinflammatory activity more or less similar to H3 formulation which contained 0.030 % w/w amount of rutin.

REFERENCES

- Y. Tanaka, J.P. Gong and Y. Osada, *Progr. Polym. Sci.*, **30**, 1 (2005).
- I. Gibas and H. Janik, *Chem. Chem. Technol.*, **4**, 297 (2010).
- K. Haraguchi and T. Takehisa, *Adv. Mater.*, **14**, 1120 (2002).
- R.V. Ulijn, N. Bibi, V. Jayawarna, P.D. Thornton, S.J. Todd, R.J. Mart, A.M. Smith and J.E. Gough, *Mater. Today*, **10**, 40 (2007).
- G. Toker, S. Turkoz and N. Erdemogly, *J. Chem. Soc. Pak*, **20**, 240 (1998).
- A. Aberoumand and S.S. Deokule, *Pak. J. Nutr.*, **7**, 582 (2008).
- L. Lima, T.T. Oliveira, T.J. Nagem, A.S. Pinto, E.Q. Lima and J.F. Silva, *Acta Farm. Bonaerense*, **22**, 21 (2003).
- M. Harwood, B. Danielewska-Nikiel, J.F. Borzelleca, G.W. Flamm, G.M. Williams and T.C. Lines, *Food Chem. Toxicol.*, **45**, 2179 (2007).
- J.C. Marcarini, R.C. Luiz, L.R. Ribeiro, C.B. Ho\$mann-Campo and M.S. Mantovani, *Exp. Toxicol. Pathol.*, **63**, 450 (2011).
- A. Basile, S. Giordano, J.A. Lopez-Saez and R.C. Cobiainchi, *Phytochemistry*, **52**, 79 (1999).
- C.A. Winter, E.A. Risley and G.W. Nuss, *Proc. Soc. Exp. Biol. Med.*, **111**, 544 (1962).
- A.A. Mohammed, A.A. Khalil and H.E.S. El-Beltagi, *Grasas y Aceites*, **61**, 67 (2010).
- B. Havsteen, *Flavonoids, Biochem. Pharmacol.*, **32**, 1141 (1983).
- J.B. Harborne and H. Baxter, *The Handbook of Natural Flavonoids*, John Wiley & Sons, Chichester, UK, Vol. 2, pp. 413-15 (1999).
- N. Trookman, R. Rizer, T.J. Stephens and R. Trancik, *J. Am. Acad. Dermatol.*, **75**, 730 (2007).
- J.Y. Chang, Y.K. Oh, H.S. Kong, E.U. Kim, D.D. Jang and K.T. Nam, *J. Control. Rel.*, **82**, 39 (2002).