Analysis of Peripheral Blood Lymphocytes of Relapsing-Remitting Multiple Sclerosis

Itaru FUNAKAWA

Division of Neurology, Department of Medicine, Kawasaki Medical School, Kurashiki 701-01, Japan Accepted for publication on September 25, 1996

ABSTRACT. Peripheral lymphocyte surface antigen subsets were analyzed in 25 patients with multiple sclerosis. These patients were divided into two subgroups of patients in the stable phase and those in relapse. As compared with a normal control group and stable phase group, the relapse group showed decreased CD4+CD45RA+ cells (p<0.0001 and p< 0.05, respectively) and increased CD4+CD45RO+ cells (p<0.005 and p< 0.01, respectively). CD3+IL-2R+ cells significantly increased during relapse as compared with the stable phase (p<0.02). CD3+ γ δ T cells decreased in the relapse group as compared with the normal control group (p < 0.05). No significant changes were observed in the CD4/CD8 ratio, unlike the findings in previous reports. It was confirmed that analysis of peripheral lymphocyte subsets provides sufficient indicators for the determination of the activity of multiple sclerosis. The decreases in CD4+CD45RA+ cells and CD3+ γ δ T cells proved to be a particularly sensitive indicator for the diagnosis of patients who had had only one attack or had not yet been diagnosed, and the increases in CD4+CD45RO+ cells and CD3+IL-2R+ cells were determined to be a sensitive indicator for the prediction of relapse among patients diagnosed as having multiple sclerosis.

Key words: multiple sclerosis — naive cell - CD45 - T cell subset

Multiple sclerosis (MS) is a demyelinating disease of the central nervous system, showing repeated remission and relapse. The etiology of this disease is thought to involve some autoimmune mechanism, but still largely remains obscure. Although symptoms can be improved in many cases by high-dose steroid therapy during an acute relapsing phase,^{1,2)} its prognosis largely depends on the availability of early detection of relapse based on objective evidence. Relapse is usually predicted by reference to clinical symptoms, magnetic resonance imaging (MRI) findings, and electrophysiological test results. However, areas with abnormal signal intensity may not be demonstrated on MRI if examination is delayed or in repeatedly relapsing cases. Therefore a much simpler routine examination may be required to detect evidence of relapse in clinical cases.

A number of studies have been performed to analyze the peripheral lymphocyte surface antigen subsets in MS. Although an increased CD4/CD8 ratio once attracted keen attention,³⁻⁵⁾ subpopulations of CD4 have been studied extensively in recent years. To date, the CD45 antigen has been determined to have five isoforms. Among these isoforms, CD4+CD45RO+ T cells, called memory cells, are now thought to have a proliferative response to

140 I Funakawa

the antigen, by which the cells have been stimulated (helper inducer), whereas CD4+CD45RA+ T cells, called naive or virgin cells, are thought to be immature cells without such function (suppressor inducer). Since Rose et al⁷ and Morimoto et al⁸ reported a decrease in naive cells from peripheral lymphocytes during aggravation of MS, a number of related studies have been published, and such decrease is now thought to be a reliable indicator of the activity of MS. In addition, interleukin-2 receptor (IL-2R) and human leukocyte antigen DR (HLA-DR), expressed on activated lymphocytes, have also been studied with regard to whether they can serve as indicators for an active phase of MS. However, memory cells, another isoform of CD45 antigen, have rarely been studied in MS.⁶

In the present study, peripheral lymphocyte surface antigens were analyzed by two-color flow cytometry using various monoclonal antibodies, to determine whether an active phase of MS occurring in Japan could be identified, and which monoclonal antibodies should be selected and combined from many existing antibodies to obtain the most effective indicator.

SUBJECTS AND METHODS

Subjects

Twenty-five patients with MS of the relapsing remitting type, treated at the Division of Neurology, Kawasaki Medical School from April 1991 to March 1995, were examined. According to Poser's diagnostic criteria, 3, 18, and 4 patients were classified as having laboratory-supported definite MS, clinically definite MS, and clinically probable MS, respectively. Among these patients, 22 relapsed, 28 attacks were observed, and 13 patients were in the stable phase. The patients in relapse, 4 males and 18 females, were aged 41.3 years on average (20-70 years). Those in the stable phase, 1 male and 12 females, were aged 34.4 years on average (19-62 years). Nineteen healthy adults, 10 males and 9 females, aged 33.3 years on average (23-51 years), were used as normal controls. Each relapse of MS was defined as aggravation of MS, categorized as 1 or more on Kurtzke's expanded disability status scale (EDSS). Peripheral blood was sampled during a very early period of relapse.

Methods

Heparinized whole venous blood was reacted with various monoclonal antibodies. The actual procedures used were as follows. After 20 μ l each of two kinds of monoclonal antibodies were placed in a Falcon tube, 100 μ l of heparinized venous blood was added to the tube, gently stirred, and then incubated at room temperature in the dark for 15 minutes. FACS Lysing Solution® (Becton Dickinson Co., Ltd.), diluted 10 times, was added at 2 ml to the tube, stirred for three seconds in vortex, incubated at room temperature in the dark for 10 minutes, and then centrifuged at 1,300 rpm for five minutes at room temperature. The supernatant was removed, and 3 ml of CELL WASH® (Becton Dickinson Co., Ltd.) was added to the pellet, stirred for three seconds in vortex, and then centrifuged at 1,200 rpm for five minutes at room temperature. After the supernatant was discarded, 1 ml of CELL WASH® was added, gently stirred, and stored in a cool dark place. Measurement was performed within two hours after sample preparation; namely, 10,000

lymphocytes were counted using FACS tar[®] (Becton Dickinson Co, Ltd). The monoclonal antibodies (moAbs) used, which were all obtained from Becton Dickinson Co, Ltd, were anti-Leu 2a (CD8) moAb, anti-Leu 3a (CD4) moAb, anti-Leu 18 (CD45RA) moAb, anti-Leu 15 (CD11b) moAb, anti-Leu 4 (CD3) moAb, anti-Leu 45RO (CD45RO) moAb, anti-interleukin-2 receptor (IL-2R) (CD25) moAb, anti-HLA-DR moAb, and anti-γST-cell receptor (TCR) moAb. The combinations of the moAbs examined are shown in Table 1. Statistical analysis was performed with Mann-Whitney's U test.

TABLE 1. Combination of monoclonal antibodies

	FITC	PE	
1	HLA-DR	Leu 4	
2	Leu 2a	Leu 3a	
3	Leu 2a	Leu 15	
4	Leu 18	Leu 45RO	
5	IL-2R	Leu 4	
6	Leu 18	Leu 3a	
7	Leu 3a	Leu 45RO	
8	γ δ-TCR	Leu 4	

FITC: fluorescein isothiocyanate

PE: phycoerythrin

RESULTS

The CD4+CD45RA+/CD4+ ratios were 0.51 ± 0.10 (mean \pm SD) in the normal control (NC) group, 0.33 ± 0.11 in the relapsing MS (Re) group, and 0.45 ± 0.11 in the stable phase (S) group. These results indicate a significant decrease in the Re group as compared with the NC and S groups (p<0.0001 and p<0.05, respectively) (Fig 1). The CD4+CD45RA-/CD4+CD45RA+ ratios were 1.06 ± 0.43 in the NC group, 2.58 ± 1.90 in the Re group, and 1.63 ± 1.31 in the S group, indicating a significant increase in the Re group as compared with the NC and S groups (p<0.0001 and p<0.05, respectively). The percent of CD4+CD45RA+ cells in relation to all lymphocytes was $19.88\pm6.94\%$ in the NC group, $12.07\pm5.54\%$ in the Re group, and $17.54\pm7.95\%$ in the S group. These percentages indicated a significant decrease in the Re group as compared with the NC and S groups (p<0.001 and p<0.05, respectively).

The CD4+CD45RO+/CD4+ ratios were 0.57 ± 0.11 in the NC group, 0.68 ± 0.14 in the Re group, and 0.51 ± 0.16 in the S group. These findings showed a significant increase in the Re group as compared with the NC and S groups (p<0.01 and p<0.01, respectively). The CD4+CD45RO-/CD4+CD45RO+ ratios were 0.85 ± 0.36 in the NC group, 0.56 ± 0.42 in the Re group, and 1.15 ± 0.69 in the S group, indicating a significant decrease in the Re group as compared with the NC and S groups (p<0.005 and p<0.01, respectively) (Fig 2). There were no significant differences in the percent of CD4+CD45RO+ cells in relation to all lymphocytes among the three groups. The CD45RA+/CD45RO+ ratios were 2.23 ± 0.74 in the NC group and 1.73 ± 1.01 in the Re group. There was a significant decrease in the Re group (p<0.02).

4.0

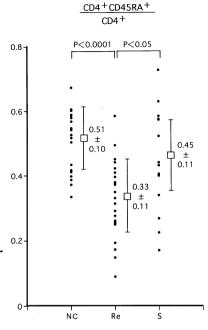


Fig 1. The mean and standard deviation are expressed as open square and vertical bars,

CD4+CD45RO CD4+CD45RO+

respectively. NC: normal control group Re: relapsing MS group S: stable MS group

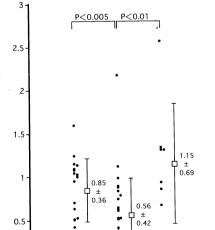


Fig 2. The mean and standard deviation are expressed as open square and vertical bars, respectively.

NC

Re

NC: normal control group Re: relapsing MS group S: stable MS group

0

The CD3+CD25+/CD3+ ratios were 0.04 ± 0.01 in the S group and 0.06 ± 0.03 in the Re group, and there was a significant increase in the Re group as compared with the S group (p<0.02). The percent of CD3+CD25+ cells in relation to all lymphocytes was $2.38\pm0.86\%$ in the S group and $4.23\pm2.36\%$ in the Re group, indicating a significant increase in the Re group (p<0.02) (Fig 3). The percent of CD3+ γ δ T cells in relation to all lymphocytes was $6.07\pm3.48\%$ in the NC group, $4.95\pm6.34\%$ in the Re group and $3.70\pm1.66\%$ in the S group. There was a significant decrease in the Re group as compared with the NC group (p<0.05) (Fig 4).

There were no significant differences in the CD4/CD8 ratio, HLA-DR⁺ cells, or the CD8/CD11b ratio between the NC, Re, and S groups (Table 2).

TABLE 2. Result

	NC	Re	S	p-value
CD3+HLA-DR+/CD3+	0.14 ± 0.07	0.22 ± 0.15	0.16 ± 0.09	ns
CD3+HLA-DR+(%)	9.39 ± 0.97	14.04 ± 10.77	10.55 ± 1.86	ns
CD4/CD8	1.55 ± 1.00	1.44 ± 0.92	1.33 ± 0.43	ns
$CD8^+CD11b^-/CD8^+CD11b^+$	$2.33\!\pm\!0.93$	3.44 ± 2.65	3.77 ± 3.41	ns

Data are mean ±S NC: normal control group

Re: relapsing MS group S: stable MS group ns: no significance

CD 3 + IL-2R+(%)

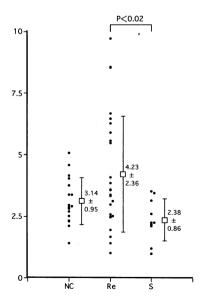


Fig 3. The mean and standard deviation are expressed as open square and vertical bars, respectively.

NC: normal control group Re: relapsing MS group S: stable MS group

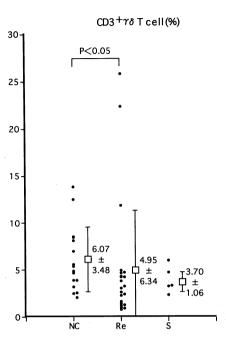


Fig 4. The mean and standard deviation are expressed as open square and vertical bars, respectively.

NC: normal control group Re: relapsing MS group S: stable MS group

DISCUSSION

Several reports on peripheral lymphocyte surface antigens in relation to MS have been published. Many investigators have reported a non-specific increase in the CD4/CD8 ratio associated with a decrease in CD8+ cells during active phases,³⁻⁵⁾ but some others have observed no such decrease.¹¹⁻¹³⁾ In recent years, it has been possible to perform more detailed analysis through fluorescent double staining of subpopulations of CD4 cells. Morimoto *et al*⁸⁾ reported a significant decrease in peripheral suppressor inducer T cells during aggravation of MS in 1987. Supplementary studies have disclosed comparable results.^{7,14-18)} From the results of these studies, it has been inferred that the non-specific decrease in CD8 cells reported previously results from a decrease in CD4+CD45RA+ cells, suppressor inducer T cells.¹⁵⁾ In the present study, I also confirmed that CD4+CD45RA+ cells decreased and CD4+CD45RO+ cells increased during a relapse of MS, as reported by Morimoto *et al*, indicating a state of strong sensitization.

In the present study, IL-2R⁺ cells, which are expressed on activated lymphocytes, were found to significantly increase in the relapsing MS group, as compared with the stable MS group. Corrigan $et\ al^{14}$ reported that IL-2R⁺ cells did not reflect the activity of the disease, whereas Selmaj $et\ al^{19}$ stated that the increase in IL-2R⁺ cells was specific to MS. Ruutiainen $et\ al^{20}$ also stated that IL-2R⁺ cells increased during aggravation phases, and decreased as the patients began to respond to treatment. Scolzzi $et\ al^{21}$ observed increases

in both HLA-DR⁺ cells and IL-2R⁺ cells in MS. In addition, it has been reported that serum soluble IL-2R increases during relapsing phases of MS.²²⁾ In the present study, IL-2R⁺ cells significantly differed in number between relapse and stable MS groups. This seems to indicate that the IL-2R⁺ cell can be useful for estimating the relapse of MS in patients diagnosed as having MS and, therefore, it can serve as a valuable indicator during follow-up of the course of stable MS.

To date, it has been clinically determined that administration of interferon (IFN)-alpha and -beta improves the symptoms of MS, but IFN-gamma worsens those symptoms.^{23,24)} It has also been reported that IFN-gamma increases about two weeks before aggravation of MS.²⁵⁾ CD4+CD45RO+ cells are major IFN-gamma-producing cells, and have been shown to stimulate the production of IL-2.^{26,27)} From these results, it can be concluded that a decrease in CD4+CD45RA+ cells, an increase in CD4+CD45RO+ cells, and an increase in IL-2R+ cells represent a series of consistent immune responses.

There have been very few studies of γ δT cells in MS. Droogan *et al*²⁸⁾ showed that, in cerebrospinal fluid (CSF), γ δT cells were significantly lower than in a non-inflammatory control, but, in peripheral blood, there were no significant differences. In the present study, the number of γ δT cells was significantly lower in the Re group. Apoptosis of γ δT cells is induced in the presence of recombinant IL-2.²⁸⁾ IL-2 and soluble IL-2R increase in MS sera.²⁹⁾ The result in the present study is compatible with these findings.

As previously noted, many investigators have reported a non-specific increase in the CD4/CD8 ratio in MS, but others have reported no such significant increase, as was the case in the present study. This may indicate that a decrease in suppressor inducer cells does not always result in a quantitative decrease in CD8 cells, but does result in a qualitative change. In addition, it has been pointed out that suppressor/cytotoxic T cells may show different CD4/CD8 ratios according to the monoclonal antibodies used. These may be the reasons for no decrease in the CD4/CD8 ratio in the present study.

HLA-DR⁺ cells did not differ among the three groups examined in the present study. This does not agree with the ability of IFN-gamma to express HLA-DR antigen on the peripheral lymphocyte surface,³²⁾ if administered, or with an increase in HLA-DR⁺ cells during relapse of MS, as reported by Scolzzi *et al*,²¹⁾ but it does agree with the results obtained by Corrigan *et al*.¹⁴⁾ Salmaggi *et al*³³⁾ reported a decrease in HLA-DR⁺ cells in MS. Therefore, changes in the peripheral HLA-DR⁺ cells in MS are still the subject of much controversy.

The effect of steroid therapy was not considered in the present study. Reports of the effects of steroid therapy on lymphocyte surface antigens have not been consistent; namely, a number of reports disclosed some changes in peripheral lymphocyte surface antigens,^{34–36)} but some others detected no such changes.²⁰⁾ This issue should be further studied.

Since an increase in cells in the CSF is small in MS, only a few reports on CSF lymphocytes have been published. Some authors compared CSF lymphocytes with peripheral lymphocytes, and found a more remarkable decrease in naive cells in CSF.^{37–39)} Scolozzi *et al*²¹⁾ found a higher expression rate of IL-2R⁺ cells in CSF as compared with peripheral blood. Further

146 I Funakawa

studies on CSF with the accumulation of more data on lymphocyte surface antigens should provide useful clinical information.

Although MRI and electrophysiological findings are useful for the prediction of relapses of MS, it is not always possible to do these examinations for various reasons. Although high-does steroid therapy is often initiated based on aggravation of EDSS in routine physical examinations before various tests provide results, it is desirable to have various parameters that can be used as indicators for relapse, even for the purpose of preventing excessive steroid administration. Although a decrease in CD4+CD45RA+ cells is not specific to MS, being also observed in active systemic erythematodes and human T lymphotrophic virus-I-associated myelopathy, 40,41) this change in peripheral lymphocyte surface antigens was found to be useful in the present study to ensure there was a relapse, and then to initiate treatment. Since there are numerous human lymphocyte surface marker antibodies, it is difficult to select a suitable combination of antibodies. From the results of the present study, it was determined that the decrease in CD4+CD45RA+ and CD3+γ δ+ peripheral lymphocytes and the increases in CD4+CD45RO+ cells and CD3+IL-2R+ cells can serve as useful indicators for relapse, and an examination limited to these antibodies also has an economic merit. It was particularly noteworthy that the decrease in CD4+CD45RA+ cells, which most significantly differed between the normal control and MS groups, is a useful indicator before definite diagnosis Increases in CD4+CD45RO+ cells and CD3+IL-2R+ cells of MS is made. should be carefully observed in patients diagnosed as having MS to detect a relapse during their clinical courses. It was concluded that for a combination of CD4 and CD45RA, the CD4+CD45RA+/CD4+ ratio or CD4+CD45RA+/ CD4+CD45RA+ ratio, and a combination of CD4 and CD45RO, the CD4+ CD45RO-/CD4+CD45RO+ ratio are more sensitive, because a significant difference may not be detected by comparing the frequencies of positive cells alone, but can be demonstrated by comparing the ratios of positive cells to negative cells.

ACKNOWLEDGMENT

I wish to express my gratitude to Prof. Akira Terao, the Division of Neurology, Kawasaki Medical School, for his guidance in this study. I also appreciate wish to thank Mss. Akiko Kita, Hisae Towaki, Narumi Kuwano, and Kimiko Takeyari of the Tissue Culture Center for their technical support.

A part of this study was supported by a grant from the Kawasaki Medical School Study Project 1992 (No4-604).

The results of the analysis of a small number of the patients examined in the present study were reported at the 33rd General Conference of the Japanese Society of Neurology (Kagoshima, 1992).

REFERENCES

1) Milligan NM, Newcombe R, Compston DAS: A double-blind controlled trial of high dose methyprednisolone in patients with multiple sclerosis: 1. Clinical effects. J Neurol Neurosurg Psychiatry 50: 511-516, 1987

 Barnes MP, Bateman DE, Cleland PG, Dick DJ, Walls TJ, Newman PK, Saunders M, Tilley PJB: Intravenous methyprednisolone for multiple sclerosis in relapse. J Neurol Neurosurg Psychiatry 48: 157-159, 1985

3) Reinherz EL, Schlossman SF: Regulation of the immune response — Inducer and suppressor T-lymphocyte subsets in human beings. N Eng J Med 303: 370-373, 1980

- 4) Compston A: Lymphocyte subpopulations in patients with multiple sclerosis. J Neurol Neurosurg Psychiatry 46: 105-114, 1983
- 5) Ghezzi A, Zaffaroni M, Caputo D, Montanini R, Cazzullo CL: Evaluation of evoked potentials and lymphocyte subsets as possible markers of multiple sclerosis: One year follow up of 30 patients. J Neurol Neurosurg Psychiatry 49: 913-919, 1986
- 6) Kasahara Y, Miyawaki T, Taniguchi N: Differentiation of CD4⁺ T cells and CD45R molecules. Clin Immunol 22: 616-624, 1990 (in Japanese)
- Rose LM, Ginsberg AH, Rothstein TL, Ledbetter JA, Clark EA: Selective loss of a subset of T helper cells in active multiple sclerosis. Proc Natl Acad Sci USA 82: 7389-7393, 1985
- 8) Morimoto C, Hafler DA, Weiner HL, Letvin NL, Hagan M, Daley J, Schlossman SF: Selective loss of the suppressor-inducer T-cell subset in progressive multiple sclerosis. Analysis with anti-2H4 monoclonal antibody. N Engl J Med 316: 67-72, 1987
- 9) Poser CM, Paty DW, Scheinberg L, McDonald WI, Davis FA, Ebers GC, Johnson KP, Sibley WA, Silberberg DH, Tourtellotte WW: New diagnostic criteria for multiple sclerosis: Guideline for research protocols. Ann Neurol 13: 227-231, 1983
- 10) Kurtzke JF: Rating neurologic impairment in multiple sclerosis: An expanded disability status scale (EDSS). Neurology 33:1444-1452, 1983
- 11) Rice GPA, Finney D, Braheny SL, Knobler RL, Sipe JC, Oldstone MBA: Disease activity markers in multiple sclerosis. Another look at suppressor cells defined by monoclonal antibodies OKT4, OKT5, and OKT8. J Neuroimmunol 6:75-84, 1984
- 12) Hirsch RL, Ordonez J, Panitch HS, Johnson KP: T8 antigen density on peripheral blood lymphocytes remains unchanged during exacerbations of multiple sclerosis. J Neuroimmunol 9:391-398, 1985
- 13) Carpa R, Marcianó N, Cotti CV, Vignolo LA, Bettinzioli M, Airó p, Cattaneo R, Gasparotti R, Moretti R: Immunological and gadolinium-DTPA MRI evaluation of relapsing remitting multiple sclerosis. Acta Neurol Scand 86: 342-345, 1992
- 14) Corrigan E, Hutchinson M, Feighery C: Fluctuations in T helper subpopulations in relapsing-remitting multiple sclerosis. Acta Neurol Scand 81:443-447, 1990
- 15) Chofflon M, Weiner HL, Morimoto C, Hafler DA: Loss of functional suppression is linked to decreases in circulating suppressor inducer (CD4+2H4+) T cells in multiple sclerosis. Ann Neurol 24: 185-191, 1988
- 16) Zaffaroni M, Rossini S, Ghezzi A, Parma R, Cazzullo CL: Decrease of CD4+CD45+ T-cells in chronic-progressive multiple sclerosis. J Neurol 237: 1-4, 1990
- Porrini AM, Gambi D, Malatesta G: Memory and naive CD4⁺ lymphocytes in multiple sclerosis. J Neurol 239: 437-440, 1992
- 18) Eoli M, Ferrarini M, Dufour A, Heltaj S, Bevilacqua L, Comi G, Cosi V, Filippini G, Martinelli V, Milanese C, La Mantia L, Salmaggi A: Presence of T-cell subset abnormalities in newly diagnosed cases of multiple sclerosis and relationship with short-term clinical activity. J Neurol **240**: 79-82, 1993
- 19) Selmaj K, Plater-Zyberk C, Rockett KA, Maini RN, Alam R, Perkin GD, Rose FC: Multiple sclerosis: Increased expression of interleukin-2 receptors on lymphocytes. Neurology 36: 1392-1395, 1986
- 20) Ruutiainen I, Salonen R, Halonen P, Panelius M, Eskola J, Salmi A: Treatment of acute exacerbations in early multiple sclerosis: cyclosporin A or prednisolone? Acta Neurol Scand 83: 52-54, 1991
- 21) Scolozzi R, Boccafogli A, Tola MR, Vicentini L, Camerani A, Degani D, Granieri E, Caniatti L, Paolino E: T-cell phenotypic profiles in the cerebrospinal fluid and peripheral blood of multiple sclerosis patients. J Neurol Sci 108:93-98, 1992
- 22) Hartung H-P, Hughes RAC, Taylor WA, Heininger K, Reiners K, Toyka KV: T cell activation in Guillain-Barré syndrome and in MS: elevated serum levels of soluble IL-2 receptors. Neurology 40: 215-218, 1990
- 23) Panitch HS, Hirsch RL, Haley AS, Johnson KP: Exacerbations of multiple sclerosis in patients treated with gamma interferon. Lancet i:893-894, 1987
- 24) Noronha A, Toscas A, Jensen MA: Contrasting effects of alpha, beta, and gamma interferons on nonspecific suppressor function in multiple sclerosis. Ann Neurol 31: 103-106, 1992
- 25) Beck J, Rondot P, Catinot L, Falcoff E, Kirchner H, Wietzerbin J: Increased production of interferon gamma and tumor necrosis factor precedes clinical manifestation in multiple sclerosis: do cytokines trigger off exacerbations? Acta Neurol Scand 78: 318-323, 1988
- 26) Hedlund G, Sandberg-Wollheim M, Sjögren HO: Increased Proportion of CD4+

200

- CDw29+CD45R-UCHL-1+ lymphocytes in the cerebrospinal fluid of both multiple sclerosis patients and healthy individuals. Cell Immunol 118: 406-412, 1989
- 27) de Jong R, Brouwer M, Miedema F, van Lier RAW: Human CD8+ T lymphocytes can be divided into CD45RA+ and CD45RO+ cells with different requirements for activation and differentiation. J Immunol 146: 2088-2094, 1991
- 28) Droogan AG, Crockard AD, Hawkins SA, McNeill TA: γ δT cell distribution in cerebrospinal fluid and peripheral blood of patients with multiple sclerosis. J Neurol Sci 126: 172-177, 1994
- 29) Gallo P, Piccinno MG, Tavolato B, Sidén A: A longitudinal study on IL-2, sIL-2R, IL-4, and IFN-g in multiple sclerosis CSF and serum. J Neurol Sci 101: 227-232, 1991
- 30) Kölmel HW, Sudau C: T-cell subsets in the cerebrospinal fluid and blood of patients with multiple sclerosis. J Neuroimmunol 20: 229-232, 1988
- 31) Thompson AJ, Brazil J, Martin EA, Hutchinson M, Whelan CA, Feighery C: Suppressor T cell changes in active multiple sclerosis: Analysis with three different monoclonal antibodies. J Neurol Neurosurg Psychiatry 48: 1062-1064, 1985
- 32) Bever CT, Panitch HS, Levy HB, McFarlin DE, Johnson KP: Gamma-interferon induction in patients with chronic progressive MS. Neurology 41:1124-1127, 1991
- 33) Salmaggi A, Baldetorp B, Milanese C, Nespolo A, Parma R, Sandberg-Wollheim M: Analysis of peripheral blood lymphocyte phenotype and function during dexamethazone treatment of progressive multiple sclerosis. Acta Neurol Scand 84:91-97, 1991
- 34) Gallo P, Chiusole M, Sanzari M, Sivieri S, Piccinno MG, Argentiero V, Rizzotti P, Tavolato B: Effect of high-dose steroid therapy on T-cell subpopulations. A longitudinal study in MS patients. Acta Neurol Scand 89:95-101, 1994
- 35) de Shazo RD, Chadha N, Morgan JE, Shortly VJ, Rangan SRS, Kalyanaraman VS, Hyslop N, Chapman Y, O'Dea S: Immunologic assessment of a cluster of asymptomatic HTLV-1-infected individuals in New Orleans. Am J Med 86: 65-70, 1989
- 36) Frequin STFM, Lamers KJB, Barkhof F, Jongen PJH, Hommes OR: T-cell subsets in the cerebrospinal fluid and peripheral blood of multiple sclerosis patients treated with high-dose intravenous methylprednisolone. Acta Neurol Scand 88: 80-86, 1993
- 37) Salonen R, Ilonen J, Jägerroos H, Syrjälä H, Nurmi T, Reunanen M: Lymphocyte subsets in the cerebrospinal fluid in active multiple sclerosis. Ann Neurol 25: 500-502, 1989
- 38) Marrosu MG: Decrease in memory (CDw29-high) cerebrospinal fluid cells from acute MS patients. Acta Neurol Scand 84:487-490, 1991
- 39) Zaffaroni M, Gallo L, Ghezzi A, Cazzullo CL: CD4⁺ lymphocyte subsets in the cerebrospinal fluid of multiple sclerosis and non-inflammatory neurological diseases. J Neurol 238: 209-211, 1991
- 40) Thomas ML: The leukocyte common antigen family. Ann Rev Immunol 7:339-369, 1989
- 41) Lal RB, Rudolph DL, Schmid DS, Lairmore MD: Concomitant augmentation of CD4+CD29+ helper inducer and diminution of CD4+CD45RA+ suppressor inducer subset in patients infected with human T cell lymphotropic virus types I or II. Clin Exp Immunol 87: 293-297, 1992