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Supporting information for article:

The structure of the complex between the arsenite oxidase from *Pseudorhizobium banfieldiae* sp. strain NT-26 and its native electron acceptor cytochrome c_{552}

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Figure S1 Proposed reaction mechanism of arsenite (As^{III}) oxidation at the Mo active site of the AioA subunit. The As^{III} [As(OH₃)] is oxidised to As^V [H₂AsO₄], reducing Mo^{VI} to Mo^{IV}. The Mo atom is re-oxidised by electron transfer to the Fe-S clusters. Adapted (Kalimuthu *et al.*, 2014).



Figure S2 Positioning of the unique cyt_{552} molecule between AioAB heterodimers presumably facilitates crystallization. (A) The AioA₂B₂ complex is comprised of chains EFK and GHL. The cyt_{552} proteins (chains K and L) are colored in salmon, the AioB subunits (chains H and F) in cyan and the AioA subunits (chains E and G) in gray. (B) The Fe-S clusters are shown as yellow and orange spheres and the Moco with oxygen and Mo atoms in red and green, respectively. The edge-edge distances between the cyt_{552} heme, the Moco and the [3Fe-4S] cluster of AioA and the [2Fe-2S] Rieske cluster of AioB are shown.



Figure S3 Packing in the AioAB/cyt c_{552} crystal. The AioAB heterodimers are colored gray. The cytc₅₅₂ molecules that are positioned at the AioAB interface (part of the 'functional' complex) are shown in salmon, and cytc552 molecules that sit between AioAB heterodimers (the 'non-functional' complexes) are colored yellow.



Figure S4 The AioAB/cyt c_{552} complex. (A) Structure of the AioAB/cyt c_{552} complex where the C_{α} trace of the heme containing cyt c_{552} is colored according to temperature factor (from blue ('low' *B*) to red ('high' *B*)). (B) Superposition of the coordinates of AioAB from the present structure, with that of the AioAB protein alone (PDB 4AAY) (Warelow *et al.*, 2013). Residues at the AioAB/cyt c_{552} interface show no major conformational changes upon complex formation. The [2Fe-2S] Rieske cluster is shown as yellow and orange spheres. AioAB and cyt c_{552} are represented in gray and salmon, respectively and the superposed structure of AioAB alone is blue.

А		66 76	
	AioB	ISELTLNEPL <mark>D</mark> VAYPD <mark>E</mark> DAAGVLLKLGT	84
	AfAioB	AKNLAANEPVSFTYPDTSSPCVAVKLGA	41
	SfSoxF	IQQVQQQIKSSGCAVYFFAYPLTDEPCFLVDLQALTGQQITEIPNPYYGKYAGPLGQI	75
	TtRp	LEELKPGDPFVLAYPMDPKTKVV-KSGEAKNTLLVARFDPE-EL	68
	SyPetC3	KGELKGNTPKGP-	35
В	cytc ₅₅₂ Rvcytc ₂ Pdcytc _{55;}	70 80 90 100 110 AGVEGFNYSPAFKAKAEEGWVWDEVHLTEYLANPKAYIKGTKMAFAGLKKPEDV GTIEGFAYSDANKNSGITWTEEVFREYIRDPKAKIPGTKMIFAGVKDEQKV AGVDGFNYSDPMKAHGGDWTPEALQEFLTNPKAVVKGTKMAFAGLPKIEDR	114 90 88

Figure S5 Secondary structure-based sequence alignment of Rieske protein and cytochrome *c* sequences related to AioAB/cyt*c*₅₅₂. **(A)** Interface residues Asp67 and Glu73 are unique to AioB (red). Residue numbers for AioB are represented in purple. **(B)** Conserved residues between the sequence of cyt*c*₅₅₂ are colored blue. Residue numbers for cyt*c*₅₅₂ are shown in red. The alignment was generated using ClustalW Omega (Sievers *et al.*, 2011). Abbreviations used are as follows: AioB, *Pseudorhizobium banfieldiae* sp. str. NT-26 (this work); *Af*AioB, AioB subunit of arsenite oxidase from *Alcaligenes faecalis*; *Sf*SoxF, Rieske protein II from *Sulfolobus acidocaldarius*; *Tt*Rp, Rieske protein from *Thermus thermophilus*; *Sy*PetC3, Rieske protein from *Synechocystis* PCC 6803; cyt*c*₅₅₂, cytochrome *c*₅₅₂ from *Pseudorhizobium banfieldiae* sp. str. NT-26 (this work); *Rv*cyt*c*₂, cytochrome c2 from *Rhodopseudomonas viridis*; *Pd*cyt*c*₅₅₂, cytochrome *c*₅₅₂ from *Paracoccus denitrificans*.



Figure S6 The heme binding site in cyt_{552} . The heme iron atom is coordinate by residues His38 and Met103. The protoporphyrin ring is covalently attached to residues Cys34 and Cys37.

Structures	AioAB/cytc ₅₅₂ ^a		SorT/SorU ^b		AxgNIR/Cytc ₅₅₁ ^c		FNR/Fd ^d		Cytochrome c ₂ :	
									reaction cen	ter ^e
Parameter	AioAB	cytc ₅₅₂	Sor T	SorU	AxgNIR	Cytc ₅₅₁	FNR	Fd	Cytochrome c_2	Reaction center
Buried surface area $(\text{\AA}^2)^{\text{f}}$	680	660	644	696	529	553	800	800	674	614
Interfacing residues ^g	25	18	31	21	10	11	25	21	21	23
Edge to edge distance between	7.5		8.2		10.5		6		8.4	
redox centres (Å)										
Hydrogen bonds	0		6		3		0		3	
Salt-bridges	2		1		0		5		0	
PDB ID	8ED4		4PW9		2ZON		1GAW		1L9J	

Table S1Comparison of the protein-protein interfaces from AioAB/cytc552

^a This work.

^b PDB code 4PW9 (McGrath *et al.*, 2015).

^c PDB code 2ZON (Nojiri et al., 2009).

^d PDB code 1GAW (Kurisu *et al.*, 2001).

^e PDB code 1L9J (Axelrod *et al.*, 2002).

^fCalculated as the average for the promoter of interest divided by the average for the entire complex structure.

^gCalculated by using PISA (Krissinel & Henrick, 2005).

Atomic packing density (ρ)	0.69
Distance (Pro122 in AioB subunit – Heme) (Å)	4.1
Average decay exponent (β)	1.4
Electron coupling (H _{DA})	1.3 X 10 ⁻³
Maximum ET rate (s ⁻¹)	1.6 X 10 ⁸

Table S2 Electron transfer parameters between AioB ([2Fe-2S]) and cytochrome c_{552} (heme)^a.

^a Calculated using HARLEM (Kurnikov, 2000).

Table S3	Kinetic parameters of enzy	mes determined with their native	electron acceptors.
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Enzyme	Electron acceptor	$K_{M(cytc)}(\mu M)$	K _{cat} (s ⁻¹)	K_{cat}/K_{M} (M ⁻¹ .s ⁻¹)
AioAB ª	Cytochrome <i>c</i> ₅₅₂	2.9 ± 0.2	390 ± 25	135.4 x 10 ⁶
SorT ^b	SorU	32 ± 5	140 ± 11	4.3 x 10 ⁶
Chicken sulfite oxidase ^c	cytochrome <i>c</i>	2.2 ± 0.05	95.0 ± 1.9	43.2 x 10 ⁶
SorAB ^d	cytochrome <i>c</i> 550	4.0	334 ± 11	8.3 x 10 ⁶

^a This work.

^b (McGrath *et al.*, 2015).

^c (Brody & Hille, 1999)

^d (Kappler & Bailey, 2005).

Protein	
AioA	AFKRHIDRLPIIPADAKKHNVTCHFCIVGCGYHAYTWPINKQGGTDPQNNIFGVDLSEQ
subunit	QQAESDAWYSPSMYNVVKQDGRDVHVVIKPDHECVVNSGLGSVRGARMAETSFSEA
	RNTQQQRLTDPLVWRYGQMQPTSWDDALDLVARVTAKIVKEKGEDALIVSAFDHGG
	AGGGYENTWGTGKLYFEAMKVKNIRIHNRPAYNSEVHGTRDMGVGELNNCYEDAEL
	ADTIVAVGTNALETQTNYFLNHWIPNLRGESLGKKKELMPEEPHEAGRIIIVDPRRTVT
	${\tt VNACEQTAGADNVLHLAINSGTDLALFNALFTYIADKGWVDRDFIDKSTLREGTARPP}$
	LYPARGVSEANPGHLSSFEDAVEGCRMSIEEAAEITGLDAAQIIKAAEWIGMPKEGGKR
	RRVMFGYEKGLIWGNDNYRTNGALVNLALATGNIGRPGGGVVRLGGHQEGYVRPSD
	AHVGRPAAYVDQLLIGGQGGVHHIWGCDHYKTTLNAHEFKRVYKKRTDMVKDAMS
	AAPYGDREAMVNAIVDAINQGGLFAVNVDIIPTKIGEACHVILPAATSGEMNLTSMNG
	${\tt ERRMRLTERYMDPPGQSMPDCLIAARLANTMERVLTEMGDVGYAAQFKGFDWQTEE}$
	${\sf DAFMDGYNKNAHGGEFVTYERLSAMGTNGFQEPATGFTDGKIEGTQRLYTDGVFSTD}$
	${\tt DGKARFMDAPWRGLQAPGKQQQKDSHKYLINNGRANVVWQSAYLDQENDFVMDRF}$
	PYPFIEMNPEDMAEAGLKEGDLVEIYNDAGATQAMAYPTPTARRGETFMLFGFPTGV
	QGNVTSAGTNELIIPNYKQTWGNIRKISDAPRNVAHLSFKSKEYQSA
AioB subunit	HHHHHHDYDIPTTENLYFQGAMGSGIQATAAAGVEYPANRLANISELTLNEPLDVA
	${\tt YPDEDAAGVLLKLGTRVEGGVGPDGDIVGFSTICPHKGFPLSYSADNKTFNCPGHFSVF}$
	DPEKGGQQVWGQATQNLPQYVLRVADNGDIFAEGVDELIYGRLSNVL
cyt <i>c</i> 552	MD ESNAEKGAVVFKKCAACHAVGDGAANKVGPELNGLIGRKVAGVEGFNYSPAFKA
	KAEEGWVWDEVHLTEYLANPKAYIKGTKMAFAGLKKPEDVADVIAYLKTFSTP <u>LEH</u>
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Table S4Complete amino-acid sequences of AioA, AioB and $cyt_{c_{552}}$.

^aBold and underlined amino-acid residues are derived from the affinity tags and cleavage sites.