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Supporting information for article:

Synergic action of an inserted carbohydrate-binding module in a glycoside hydrolase family 5 endoglucanase

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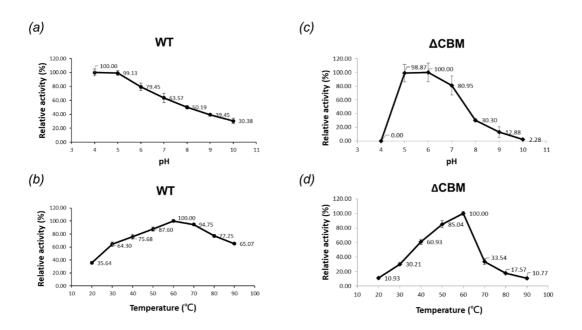
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	* PGNTSTPVPSVISG AKQEEWAEAGLIN- SMDQSVAES STEQSVAEI SKEGETVH SKEGETVH SKEGETVH SKEGETVH SKEGETVH								
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MtGlu5 : 503328546 : 503325617 : 503546037 : 501542033 : 493987263 : 501578355 : 504265218 : 568335214 : 658466377 : 428131024 : 490201866 : 500776899 :	R         PD-         PNIE         YT           R         PD-         PNIE         YT           R         PD-         PNIE         YT           T         PET         RNIE         YT           F         PRECNIE         YT         PD           V         PDD         R-IE         LT           Y         PDD         R-IE         LT           Y         PDD         R-IE         LT           V         PDD         R-IE         L           SV         PMENS         SV         R           SV         PMENS         SV         R           V         PDD         R-IE         L         SV           SV         PMENS         SV         R           V         PDD         R-IE         L         SV           L         SV         M         SV         R	320 TIDELETTICGA M TIDELETTICGAM	* 34 N PVPPTCV EG-SKNDTGT SG-SEKVIGT N PSPPVGV G PSPPVGV N PSPPVGV EG-SEKWIGR N PTPVGV EG-SEKWIGR N PTPVGV CN PRLPVGV N PRLPVGV S PRLPVGV S PTLPIGV S PTLPIGV	0 * VIR RENGAFAAGW 10 ° TT - 10 ° TT - 1	360	* EALEITYQEG	380 WAGFYLHSDA	* AGVEGYDRLAF:	400 RTSAP : 309 : 251 : 244 : 243 : 243 : 243 : 243 : 245 : 245 : 246 : 247 : 255
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MtGlu5 : 503328546 : 503325617 : 503546037 : 501542033 : 493987263 : 647306738 : 50426218 : 568355214 : 658466537 : 428131024 : 490201666 : 500776899 :	FGANGRGLLER FGANGR DMUSR FGANGR DMUSR	520 VRWTGRVRSEL KR VRWTSVARS KR ARWTSVARS KR VRWTSVARS KR VRWTSVARS KR VRWTSVARS KR VRWTSVARS KR VRWTSVRRS KR VRWTSVRRS KR VRWTSVRRS KR VRWTSVR KR VRWT KN V AC	* 54 FSW YWE AA FSW YWE AA FSY YWE AA	0 * G FGI RTTRCM G FGA PUINOT FGA PUINOT FGA PUINOT FGA PUINOT FGA PINOT FGA PINOT FGA PINOT FGA PINOT FGI RWSCA FGI RWSC	560 TILR LYPEOP DIRC LLESTS DILG LYPES LIN LIPETK NULLETT DILG LIPETK DILG LIPETK DILG VUCEK DIAT VUCEK DIAT VUCEK DIAT VUCE DIAT VUCE	*	: 470 : 350 : 392 : 333 : 335 : 346 : 301 : 335 : 3360 : 337 : 357 : 354 : 335 : 343		

MtGlu5: Miothermus taiwanensis WR220

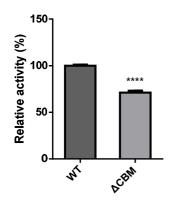
503328546: Isosphaera pallida

- 503325617: Anaerolinea thermophila
- 503546037: Mahella australiensis
- 501542033: Dictyoglomus thermophilum
- 493987263: Caldithrix abyssi
- 647306738: Dictyoglomus turgidum
- 501578355: Caldicoprobacter oshimai
- 504265218: Fervidobacterium pennivorans
- 568335214: Thermotoga maritima
- 658466537: Thermotogae bacterium JGI 0000106-011
- 428131024: Fervidobacterium gondwanense
- 490201866: Thermosipho africanus
- 500776899: Fervidobacterium nodosum

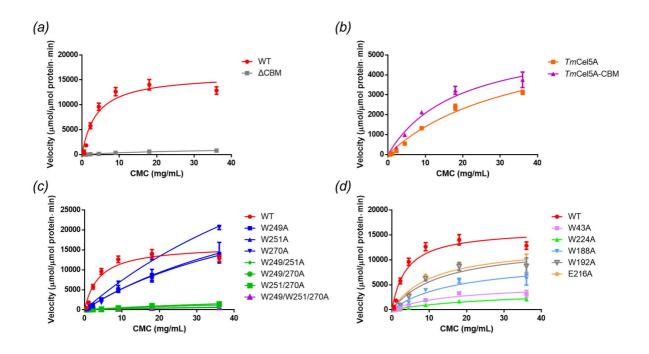
Figure S1 Multiple-sequence alignment of *Mt*Glu5 compared with other GH5 cellulases.



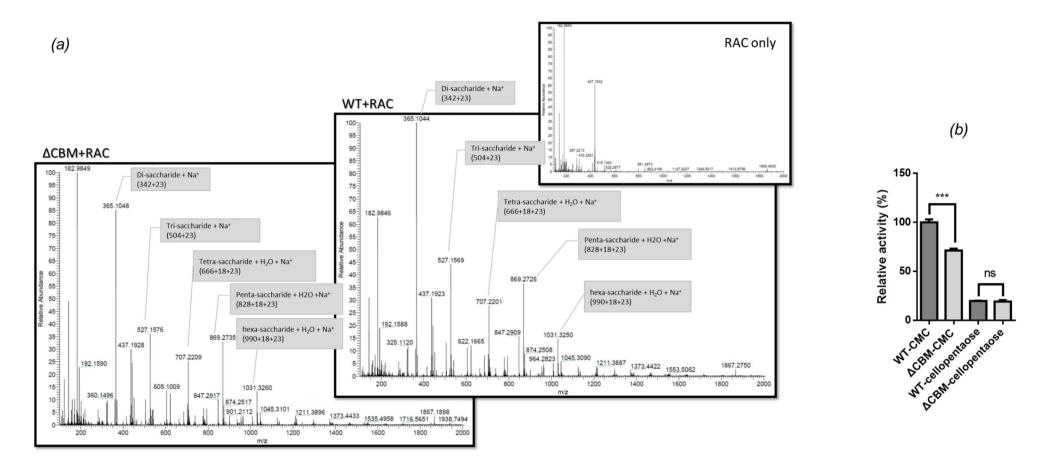
**Figure S2** The effect of reaction pH and temperature on enzyme activity on CMC. Data are shown as the means  $\pm$  SD of more than three replicates. Both WT (*a*, *b*) and  $\Delta$ CBM (*c*, *d*) present an optimum condition at 60 °C and pH 5.



**Figure S3** The activity comparison between MtGlu5 (WT) and  $\Delta$ CBM toward CMC. The specific activities of MtGlu5 and  $\Delta$ CBM toward CMC are  $385 \pm 5$  IU and  $275 \pm 8$  IU. Data are exhibited as the means  $\pm$  SD of more than three replicates. \*\*\*\* indicates statistically significant at the level of P < 0.0001 compared to WT.

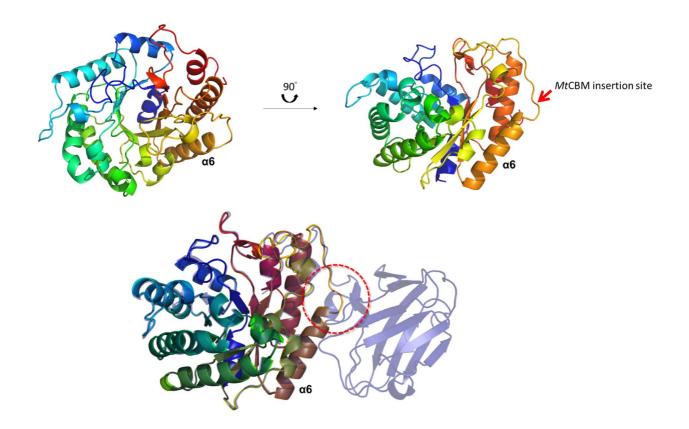


**Figure S4** Reaction kinetics curves toward CMC. Comparison of the reaction kinetics curves among *Mt*Glu5 (WT) with  $\Delta$ CBM (*a*), *Tm*Cel5A with *Tm*Cel5A-CBM (*b*), WT with Trp mutagenesis in CBM (*c*), and WT with Trp mutagenesis in the catalytic domain (*d*), respectively.

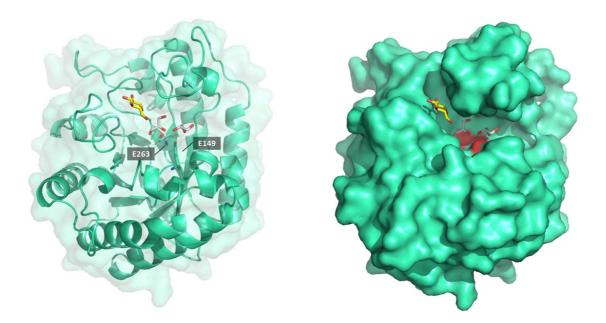


**Figure S5** The catalytic centre integrity of WT and  $\triangle$ CBM. *(a)* Hydrolyzed products are analyzed by mass spectrometry. Both WT and  $\triangle$ CBM produce the same products after hydrolysis. m/z 182.98 and m/z 437.19 are considered contaminants in the background. Two enzymes were mixed with RAC and reacted for 24 hrs. The reactions were stopped by boiling. Then the pellet was removed, and hydrolyzed products remained in the supernatant fraction. Desalting was necessary before MS analysis. *(b)* Enzyme activities toward CMC and cellopentaose. Catalysis that would not use the CBM domain is demonstrated by using

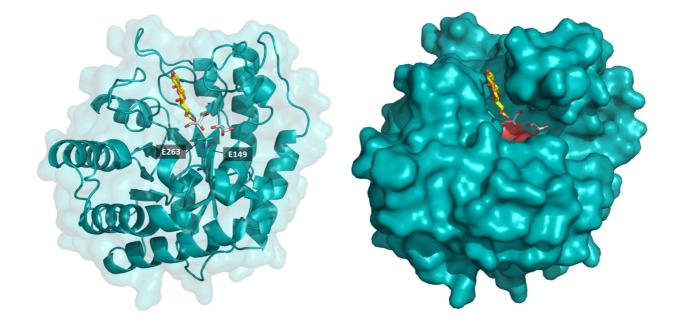
cellopentaose as a substrate. The data show that the enzyme activity of MtGlu5 (WT) and  $\Delta$ CBM toward cellopentaose are almost the same, although lower than the activity toward CMC, indicating CDs are not perturbed by the CBM truncation. (Left-half panel) Enzyme activity toward CMC, \*\*\* indicates statistically significant at the level of P < 0.001 compared to MtGlu5. (Right-half panel) Enzyme activity toward cellopentaose, ns represents no statistical significance as compared to MtGlu



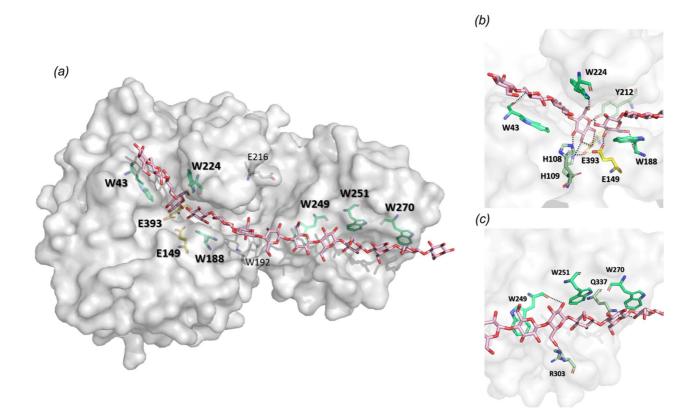
**Figure S6** The structure of  $\triangle$ CBM ramped from the N-terminus (blue) to the C-terminus (red) (PDB: 7VT5). The apo-form  $\triangle$ CBM structure displayed an intact TIM-barrel fold. The red arrow indicates the location of *Mt*CBM insertion site (a.a 235 of *Mt*Glu5). Superimposition of *Mt*Glu5 and  $\triangle$ CBM highlights the slight change in helix  $\alpha 6$ .



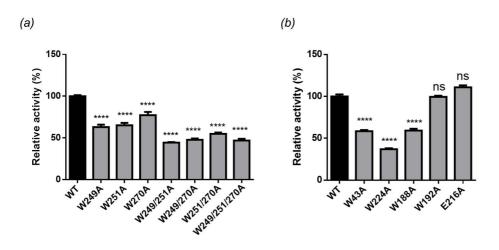
**Figure S7** The structure of  $\triangle$ CBM in complex with glucose (PDB: 7VT6). Glucose is shown in a stick form (yellow) whereas two glycerol molecules that occupied the catalytic site are shown in white sticks.



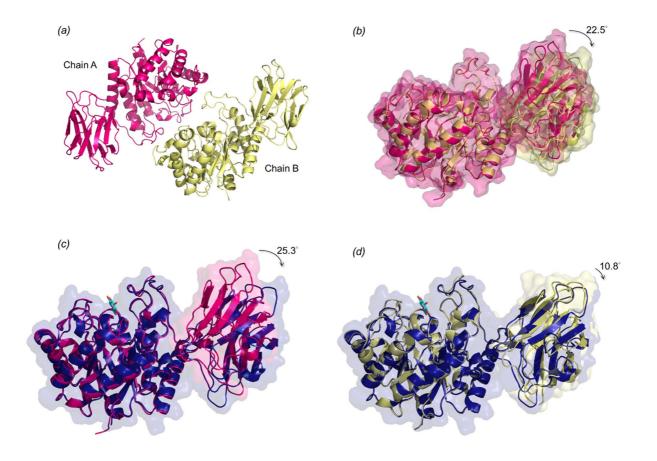
**Figure S8** The structure of  $\triangle$ CBM in complex with cellobiose (PDB: 7VT7). Cellobiose is shown in a stick form (yellow) whereas two glycerol molecules that occupied the catalytic site are shown in white sticks.



**Figure S9** Substrate binding model of MtGlu5 depicted in surface structure. (a) A 14-sugar-long single polysaccharide chain, shown in stick format (pink), is bound along the putative binding groove full of Trp residues (lime green) on the cleft surface. In this model, the polysaccharide chain is stabilized by the arrangement of the aromatic residues in the cleft of MtGlu5. The catalytic residues, E149 and E393, are shown in yellow and stick representation while the less essential residues, W192 and E216, are shown in white. The binding model is superimposed onto TmCel5A in complex with cellotetraose (PDB: 3azt) and CBM29-2 in complex with cellohexaose (PDB: 1W8T) then 14-polysaccharides are put in by *coot*. Energy minimization is simulated by the software YASARA Energy Minimization server. Snapshots of hydrogen bonds (black dashed lines) at catalytic pocket (*b*) and CBM binding surface (*c*). The surface Trp residues are shown as described and the other active-site residues are shown in olive green. (*b*) In the catalytic pocket, W43, W224, and W188 show not only steaking interaction but a direct hydrogen bond to the substrate. (*c*) W249, R303, and Q337 residues make direct interactions with the substrate in the same way as CBM29-2.



**Figure S10** The activity comparison between MtGlu5 (WT) and Trp mutants on CMC. Data are exhibited as the means  $\pm$  SD of more than three replicates. \*\*\*\* indicates statistical significance at the level of P < 0.0001 compared to WT, and *ns* is no significant differences compared to WT, respectively. (*a*) Activity comparison among WT and CBM mutants. (*b*) Activity comparison among WT and CD mutants.



**Figure S11** Structure comparison of *iMt*Glu5 (PDB: 7VT4) with *Mt*Glu5 (PDB: 7VT8). *(a)* crystal structure of *iMt*Glu5 shows dimer in the asymmetric unit. *(b)* By overlaying two chains of *iMt*Glu5, the CBMs of the two *iMt*Glu5 molecules differ by a rigid-body rotation of 22.5°. *(c)* Superimposition of glucose-bound *Mt*Glu5 and *iMt*Glu5 chain A and chain B *(d)*.

## Table S1 List of primers.

Construct	Primers (5' to 3')			
MtGlu5	F1: TATGATGGGCTGCCAATCCACC			
	F2: TGATGGGCTGCCAATCCACC			
	R1: AGCTTCGGTTGCTCCGGTACAA			
	R2: TCGGTTGCTCCGGTACAA			
i <i>Mt</i> Glu5	F1: TCGTGGGGCAGTTCGGAGCCTAC			
ΔCBM	F2: CCTACGAGAAGGGCGATCT			
	R1: CTCCGAACTGCCCCACGAAGA			
	R2: AGATGGGGCGCCGGTTCT			
W249A	F1: CAGAACGCATCCTGGGGCAGCCGG			
iW249A	F2: AGCCGGGTCGGTTTTGTGGG			
	R1: GCCCCAGGATGCGTTCTGCCAGCCCG			
	R2: CCAGCCCGCCGCGAAGGC			
W251A	F1: TGGTCCGCAGGCAGCCGGGTCG			
iW251A	F2: GTCGGTTTTGTGGGTGAAGCT			
	R1: CCGGCTGCCTGCGGACCAGT			
	R2: GTTCTGCCAGCCCGCCGC			
W270A	F1: AAGGCGCAGCCGGGTTCTATTTG			
iW270A	F2: ATTTGCACTCGGACGCTGGG			
	R1: AGAACCCGGCTGCGCCTTCCTGGTA			
	R2: CCTGGTAGGTGATCTCTAAAGCT			
W249/251A	F1: TCCGCAGGCAGCCGGGTCGGTTTTGT			
iW249/251A	F2: GTCGGTTTTGTGGGTGAAGCTT			
	R1: CCGGCTGCCTGCGGATGCGTTCTGCCAGC			
	R2: TGCGTTCTGCCAGCCCGCCGCGAA			
W249/270A	Same as W249A using W270 as a template			
iW249/270A				
W251/270A	Same as W251A using W270 as a template			

iW251/270A	
W249/251/270A	Same as W249/251A using W270 as a template
iW249/251/270A	
W43A	F1: GGGGCCGCAGGGGTCAGGCT
iW43A	F2: CTGGAGGAAGGGTTTTTCGAGCTG
	R1: CCTGACCCCTGCGGCCCCTTCCC
	R2: TTCCCAGGGGGCCTCGAGGGC
W188A	F1: GTGGGTGCCAACTCGCTGTGGCGGCTG
iW188A	F2: TGGCGGCTGTCCGAGCTGCG
	R1: CAGCGAGTTGGCACCCACCGGCCCC
	R2: CGGCCCCACAATCACCGCCC
W192A	F1: TCGCTGGCCCGGCTGTCCGAGCTG
iW192A	F2: GAGCTGCGGCTGCCGGACGAT
	R1: GGACAGCCGGGCCAGCGAGTTCCAAC
	R2: GTTCCAACCCACCGGCCCCAC
E216A	F1: CCCCTGGCGTTCACCCACCAGG
iE216A	F2: CAGGGGGCTGAGTGGCTC
	R1: GTGGGTGAACGCCAGGGGATCGTA
	R2: ATCGTAGTAGTGGAAGGTAACGA
W224A	F1: GCTGAGGCTCTCAACCCCGTTCCACC
iW224A	F2: GTTCCACCTACCGGTGTGGTCTGG
	R1: GGGGTTGAGAGCCTCAGCCCCCTG
	R2: CCCCTGGTGGGTGAACTCCAGGGG
ΔCBM	F1: CACCAGCAAAACGCCATAGC
	F2: GCCCAGGCCATGGAGTTT
	R1: TATGGCGTTTTGCTGGTGCCAGACCACACCGGTAGG
	R2: CCAGACCACACCGGTAGG
g <i>Mt</i> CBM	F1: TATGTCCCCTATACTAGGTTATTGG
	F2: TGTCCCCTATACTAGGTTATTGG
	R1: AGCTTGGTCAGCAAAGCCAGCGG

	R2: TGGTCAGCAAAGCCAGCGG			
	GST-F: TGTCCCCTATACTAGGTTATTGG			
	GST-R: CACCCTGGTTCTCACGGGATCCACGCGGAACCAG			
	CBM-F: CGTGAGAACCAGGGTGCGTTC			
	CBM-R: TGGTCAGCAAAGCCAGCGG			
GST	F1: TATGATGTCCCCTATACTAGGTT			
	F2: TGATGTCCCCTATACTAGGTT			
	R1: AGCTTTTTTGGAGGATGGTCGCC			
	R2: TTTTTGGAGGATGGTCGCC			
TmCel5A	F1: TATGATGGGTGTGGATCCGTTC			
	F2: TGATGGGTGTGGATCCGTTC			
	R1: AGCTTTTCGATGCTGTCGCCACC			
	R2: TTTCGATGCTGTCGCCACC			
TmCel5A-CBM	F1: TATGATGGGTGTGGATCCGTTC			
	F2: TGATGGGTGTGGATCCGTTC			
	R1: AGCTTTTCGATGCTGTCGCCACC			
	R2: TTTCGATGCTGTCGCCACC			
	Cel5A-F1: TGATGGGTGTGGATCCGTTC			
	Cel5A-R1: GCCCCATTTACGACCCAGCCACTTCTC			
	CBM-F: CTGGGTCGTAAATGGGGGCCGTGAGAACCAGGGTGCGTT			
	CBM-R: CTTTTGATCGTCCGGGCTGGTCAGCAAAGCCAGCGG			
	Cel5A-F2: AGCCCGGACGATCAAAAGCACCTGA			
	Cel5A-R2: TTTCGATGCTGTCGCCACC			
iTmCel5A	F1: ATCGGCCAATTCGGCGCGTATC			
iTmCel5A-CBM	F2: TATCGTAAGGCGGATCTGGAAAGCC			
	R1: CGCGCCGAATTGGCCGATGTAAATC			
	R2: GTAAATCGGACGTTTGTTTTCTTGC			

		Reducing	Reducing sugar (%)	
	Enzyme	Supernatant	Insoluble	Sol./Insol.
RAC	MtGlu5	72.6	27.4	2.65
RAC	ΔCBM	62.5	37.5	1.67

Table S2	The processivity	of MtGlu5	towards RAC.
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The processivity is presented as the ratio of the reducing ends in the soluble part to the one in the insoluble part after hydrolysis. We tested the classic methods to verify the processivity by determining the ratio of soluble reducing ends to insoluble reducing sugar ends products using both RAC and filter paper (Data not shown) as substrates, both *Mt*Glu5 and  $\Delta$ CBM present the ratio of around 2.65 and 1.67, by the criterion exhibited in the paper (Irwin *et al.*, 1993). Therefore *Mt*Glu5 is clearly an endo-glucanase.