

Volume 76 (2020) Supporting information for article:

Crystal structures of human ENPP1 in apo and bound forms Matthew L. Dennis, Janet Newman, Olan Dolezal, Meghan Hattarki, Regina N. Surjadi, Stewart D. Nuttall, Tam Pham, Tom Nebl, Michelle Camerino, Poh Sim Khoo, Brendon J. Monahan and Thomas S. Peat

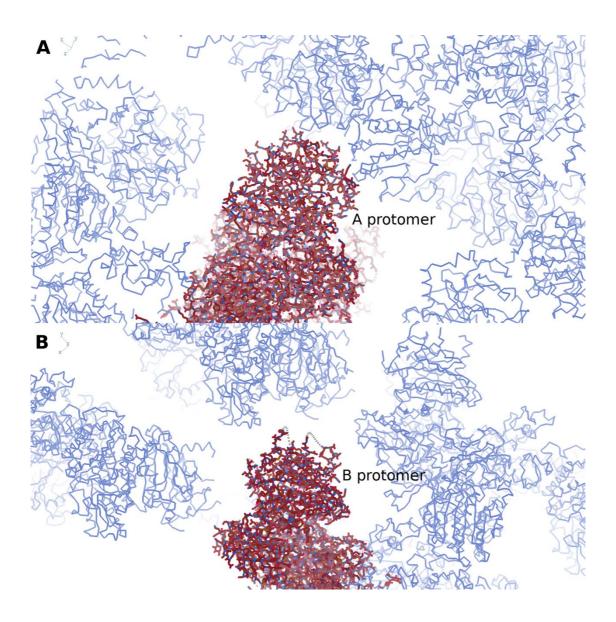
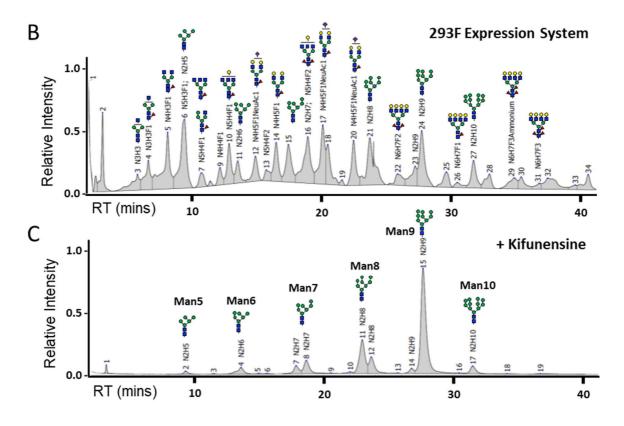


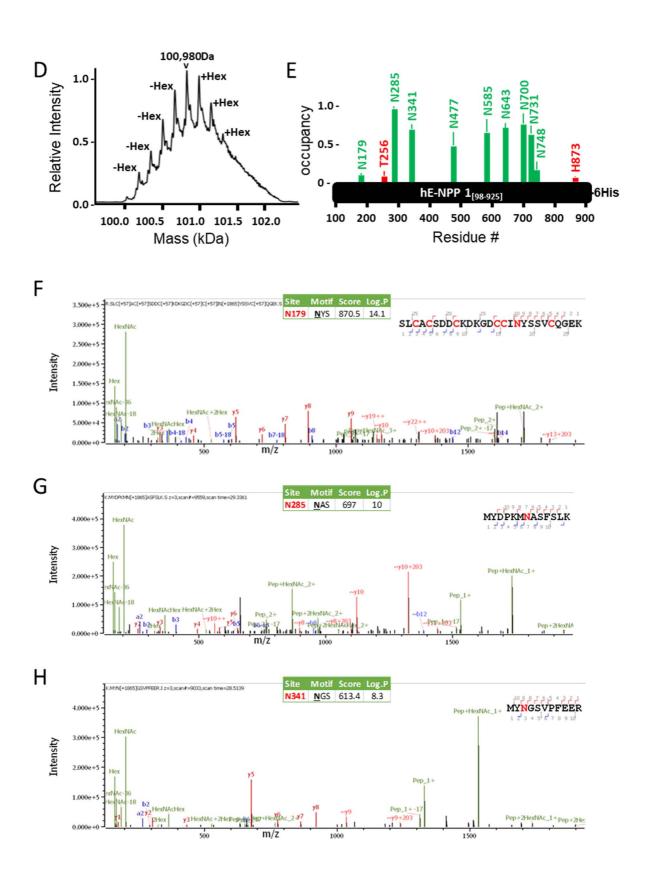
Figure S1 Crystal contacts of hENPP1 (nuclease-like domain). A: Shows the packing of the crystal around the A protomer with the nuclease-like domain in the center of the figure (in red). B: Shows the packing of the crystal around the B protomer with the nuclease-like domain in the center of the figure (in red).

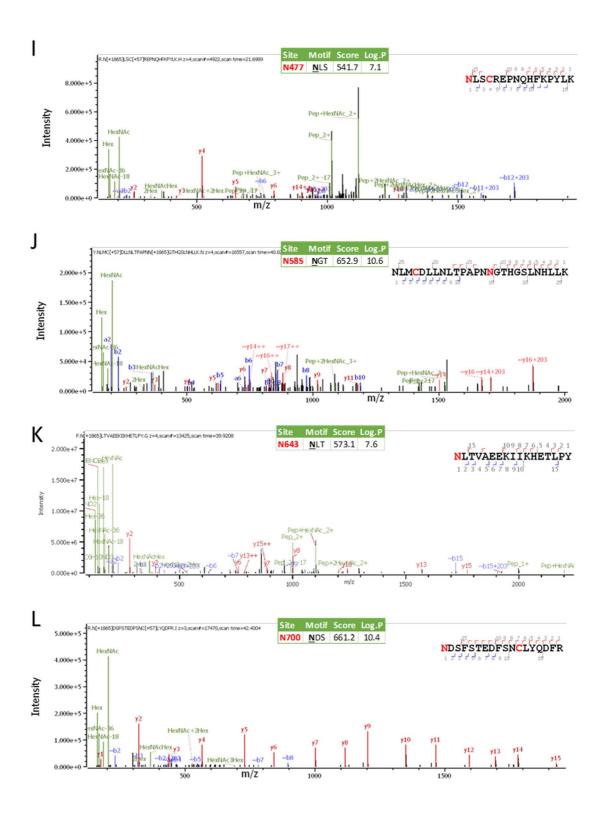
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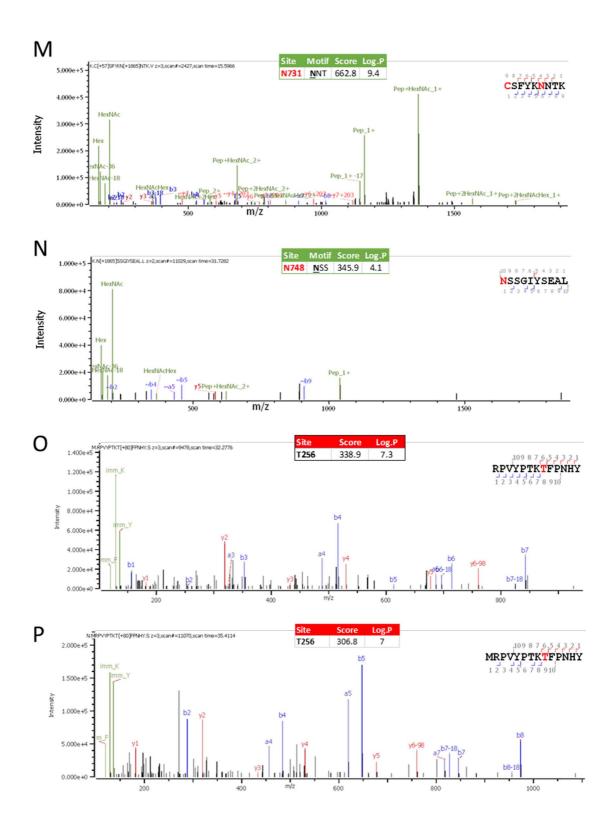
Accession	Protein	Log Prob	Peptides	% Cov.	
P22413	ENPP198-925	3757	767	96%	

10	20	30	40	50	60	70	80	90	10
MERDGCAGGG	SRGGEGGRAPRE	GPAGNGRDRGR	SHAAEAPGDP	QAAASLLAPM	OVGEEPLEKA	ARARTAKDPN	TYKVLSLVLS	VCVLTTILGC	IFGLKP
110	120	130	140	150	160	170	180	190	20
CAKEVKSCKG	RCFERTFGNCRC	DAACVELGNCC	LDYQETCIEP	EHIWTCNKFR	CGEKRLTRSL	CACSDDCKDK	GDCCINYSSV	CQGEKSWVEE	PCESIN
210		230	240	250	260	270	280	290	30
PQCPAGFETP	PTLLFSLDGFRA	EYLHTWGGLLP	VISKLKKCGT	YTKNMRPVYP	TKTFPNHYSI	VTGLYPESHG	IDNKMYDPK	MNASFSLKSK	EKFNPE
310	320	330	340	350	360	370	380	390	40
YKGEPIWVTA	KYQGLKSGTFFW	PGSDVEINGIF	PDIYKMYNGS	VPFEERILAV	LQWLQLPKDE	RPHFYTLYLE	EPDSSGHSYG	PVSSEVIKAL	QRVDGM
410		430	440	450	460	470	480	490	50
GMLMDGLKEI	NLHRCLNLILIS	DHGMEQGSCKK	YIYLNKYLGD	VKNIKVIYGP	AARLRPSDVP	DKYYSFNYEG	ARNLSCREP	NQHFKPYLKH	FLPKRL
510	-	530	540	550	560	570	580	590	60
FAKSDRIEPL	TFYLDPQWQLAI	NPSERKYCGSG	FHGSDNVFSN	MQALFVGYGP	GFKHGIEADT	FENIEVYNLM	COLLNLTPAP	NNGTHGSLNH	LLKNPV
610		630	640	650	660	670	680	690	70
TPKHPKEVHP	LVQCPFTRNPRD	NLGCSCNPSIL	PIEDFQTQFN	LTVAEEKIIK	HETLPYGRPR	VLQKENTICL	LSQHQFMSGY	SQDILMPLWT	SYTVDR
710		730	740	750	760	770	780	790	80
DSFSTEDFSN	CLYQDFRIPLSP	VHKCSFYKNNT	KVSYGFLSPP	QLNKNSSGIY:	SEALLTTNIV	PMYQSFQVIW	RYFHDTLLRK	YAEERNGVNV	VSGPVF
810		830	840	850	860	870	880	890	90
FDYDGRCDSL	ENLROKRRVIRN	QEILIPTHFFI	VLTSCKDTSQ	TPLHCENLDT	LAFILPHRTD	NSESCVHGKHI	DSSWVEELLM	LHRARITDVE	HITGLS
910	920								









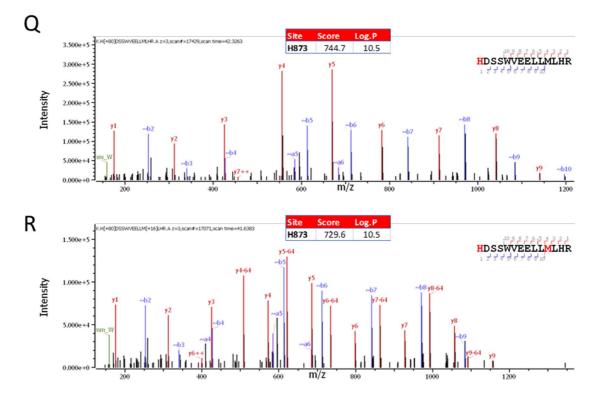


Figure S2 Mass spectrometric analysis. A: Peptide sequencing of a kifunensine-treated, affinity purified hENPP1-HIS preparation by LC-MSMS confidently identified 767 unique peptides with 96% sequence coverage matching of hENPP1 residues 98-925 (green). Nine predicted N-linked glycosylation sites of hENPP1 are highlighted in yellow, based on UniProt entry P22413. B: N-linked glycoprofiling of hENPP1 expressed in Freestyle 293 cells identified over 30 different glycan structures, including predominantly high-mannose and complex, core-fucosylated glycan structures. C: Kifunensine treatment significantly reduced N-glycan heterogeneity and resulted in hENPP1 decorated with high-mannose-type glycan structures of Man9 > Man8 > Man7 type. **D:** Intact mass determination of a kifunensinetreated hENPP1 154-925 construct is consistent with 13.8-15.0kDa N-linked glycan mass additions (+/- Hex). The major observed mass of 100,980Da matches the expected protein mass (MW~86,556.5Da) decorated with 4xMan8 + 4xMan9 N-linked glycans. E: A diagram depicting the relative occupancy of N-linked glycosylation sites (green) or phosphorylation sites (red) of hENPP1 based on spectrum counting (e.g. fraction of modified versus total peptide spectra matches per site; height of bars and whiskers correspond to average +/standard error of three independent biological replicates). F-N: Highly confident LC-MSMS evidence spectra showing Man9-modified glycopeptide sequences for glycosylation sites N179, N285, N341, N477, N585, N700, N731 and N48. O-R: LC-MSMS evidence for peptide sequences containing phosphorylated T256 or H873.

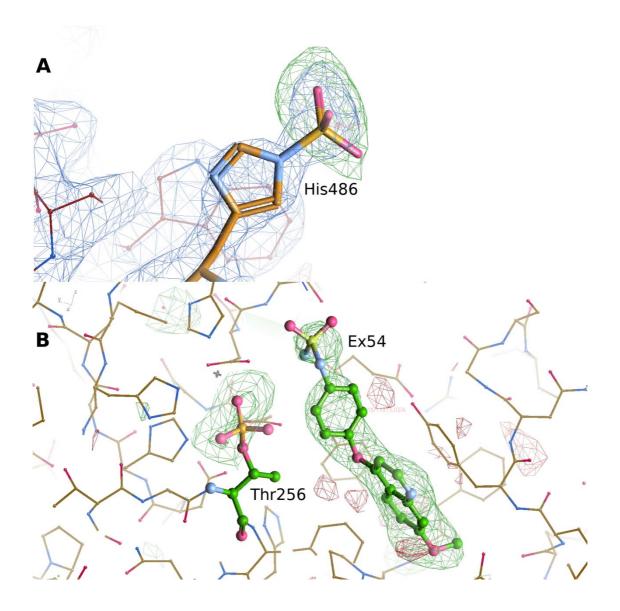


Figure S3 Difference density for phosphorylation sites A: Difference density map (green) set at 3 σ for His486 showing significant difference density in the original MR map generated (difference density for the phosphate is still seen at 7 σ). The blue density is set at 1.5 σ . B: Difference density map set at 3 σ for both Thr256 and Ex54 in the catalytic site of hENPP1. Figure was generated using Coot and The Gimp.

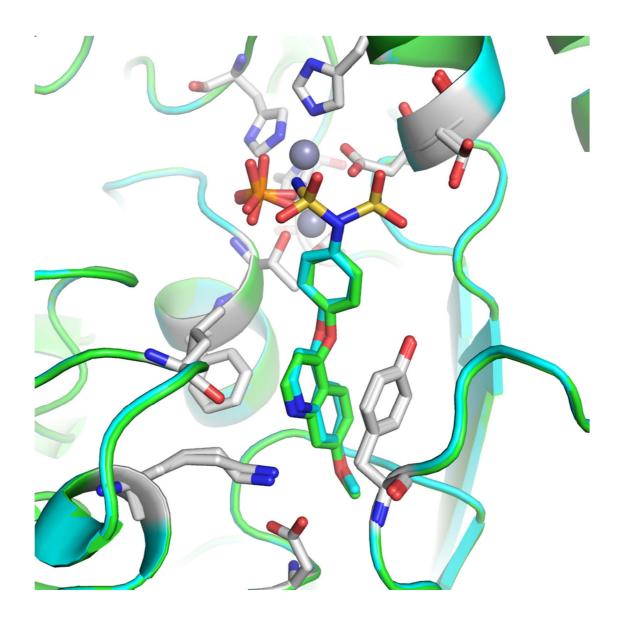
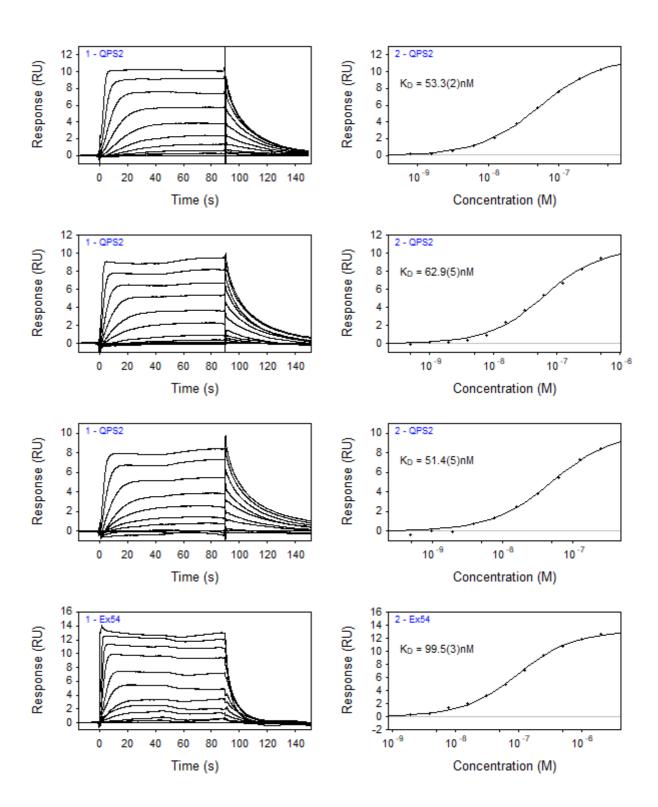
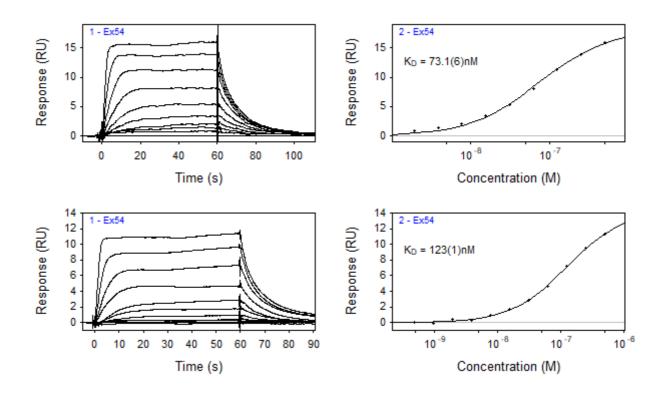
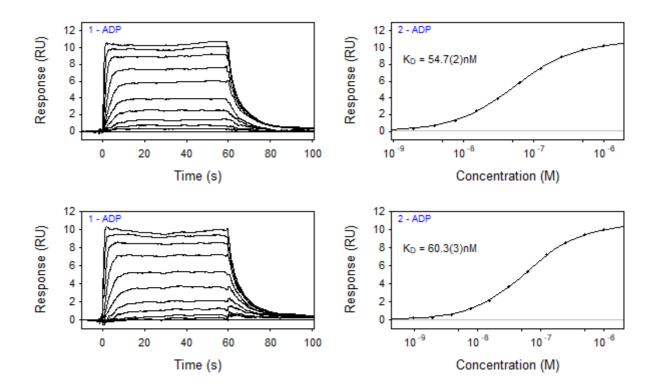
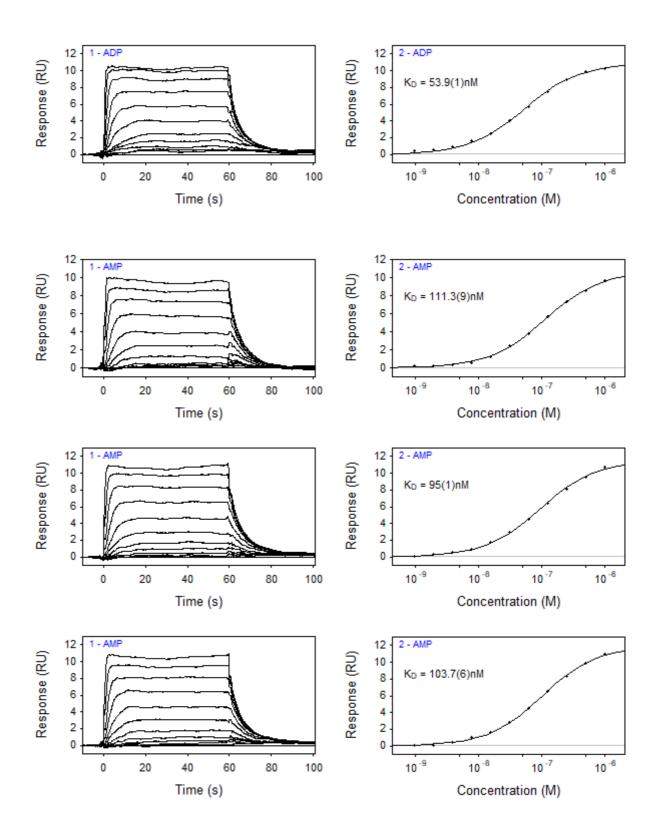


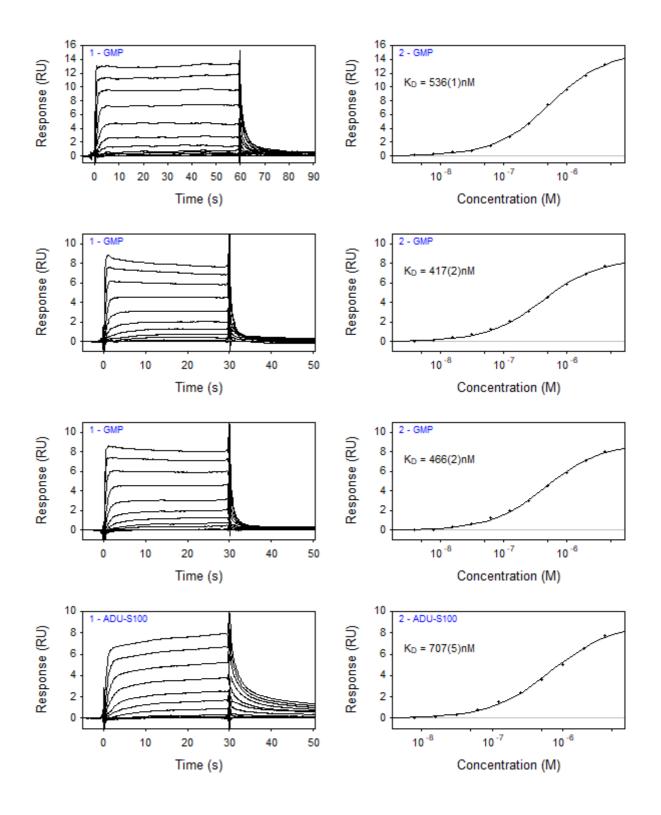
Figure S4 Ex54 sulfamide movement. Variable sulfamide position of Ex54. The A and B protomers were superposed using the SSM algorithm in Coot. There is little variation between these active sites, but one can see that the sulfamide position of the two Ex54 molecules is quite different. Figure was generated with PyMol.

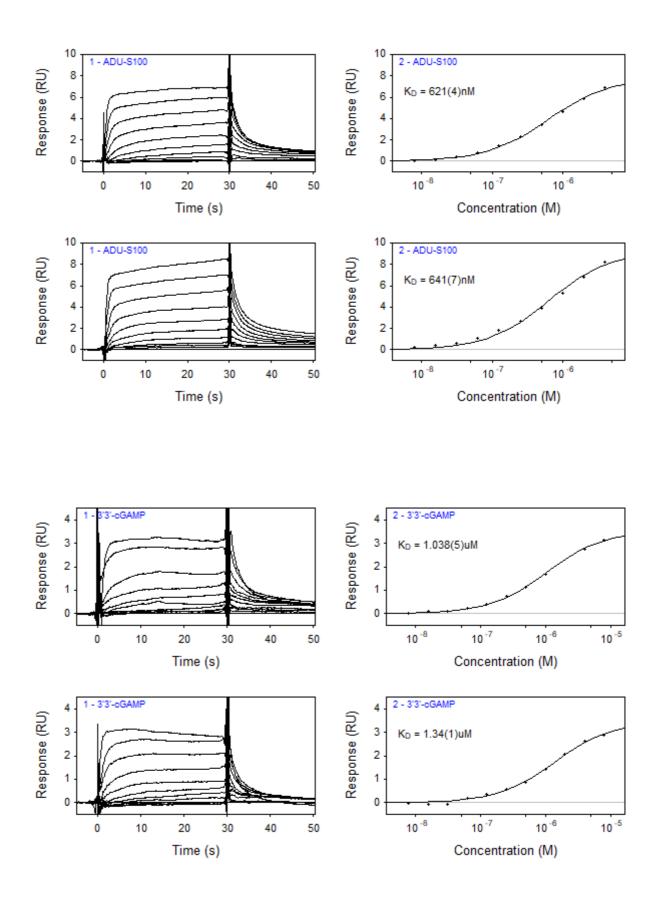


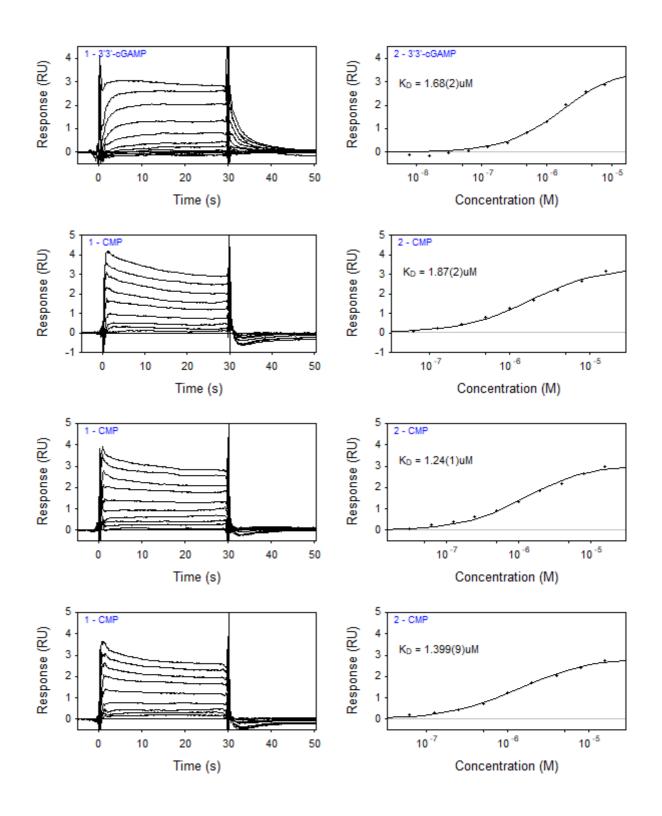


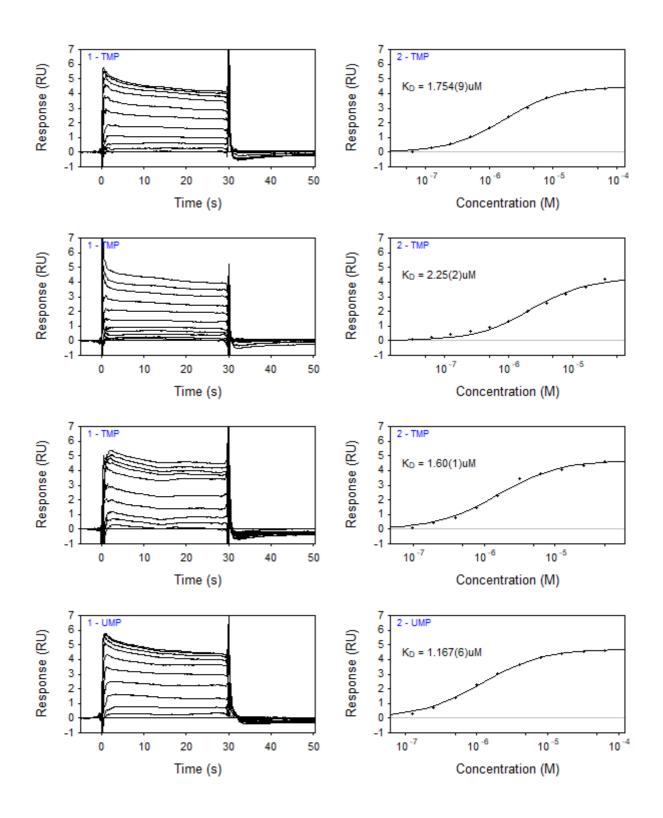












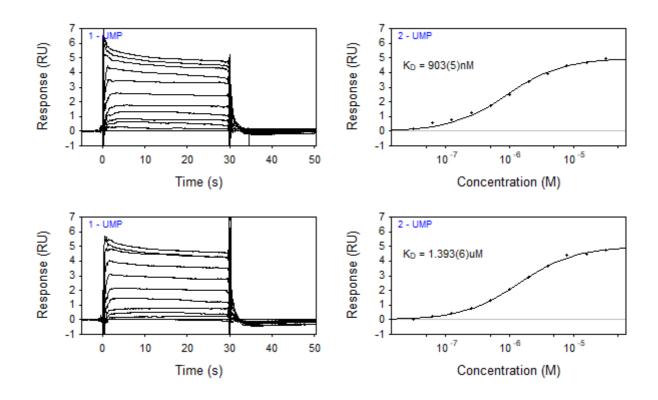
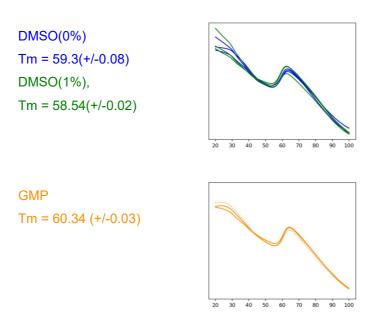
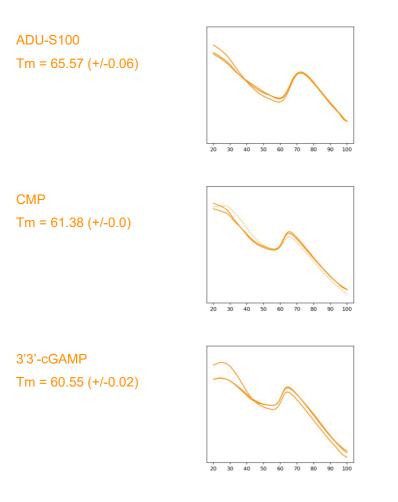


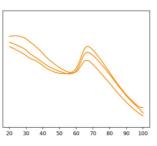
Figure S5 SPR sensorgrams and binding isotherms.



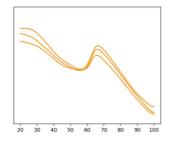


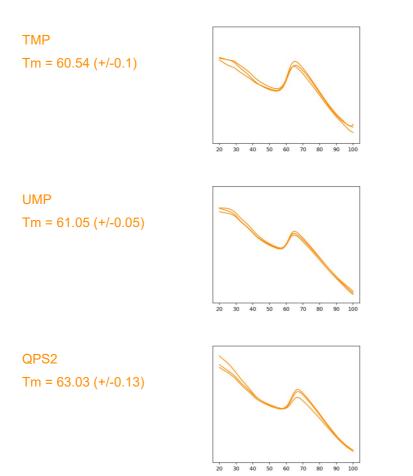
ADP

Tm = 62.6 (+/-0.08)

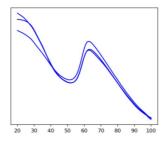


AMP Tm = 62.01 (+/-0.1)





DMSO(1%) Tm = 58.37(+/-0.07)



Ex54 Tm = 62.15 (+/-0.03)

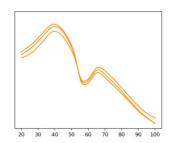


Figure S6 DSF melt curves. The x-axis presents temperature (°C) and the y-axis presents normalised RFU (relative fluorescence units).