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Supporting information for article:

A DNA structural alphabet provides a new insight into DNA flexibility

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Methods

Retrieval of sequentially non-redundant structures

Selection of sequentially non-redundant set of structures. To avoid using sequentially redundant structural data and reduce statistical bias, we compared structures sequentially. Before the sequential alignment, we excluded N-terminal and C-terminal tags, which were presumed not to interact with DNA. ClustalX (Larkin *et al.*, 2007) was used to divide protein sequences into two groups, sequentially unique and sequentially redundant. Proteins were labeled as identical in case of their 100% sequence identity. In the next step, we compared DNA sequences in complexes containing proteins marked as sequentially redundant and those with differed DNA sequences were kept. Several complexes with the same or similar protein sequence but different DNAs provide unique protein-DNA interfaces and contribute to the dataset by structural information.

DNA sequences were termed sequentially unique when their sequences shorter than 24 nucleotides differed by at least two nucleotides and sequences longer than 24 nucleotides sequences by at least 3 nucleotides. DNA sequences in structures of the histone core particle were aligned and sorted into groups, in which DNA differed by less than 11 nucleotides. There were nine such groups and in each, we selected one structure deemed the best. The best structure in a group of structures tagged as sequentially redundant was selected based on crystallographic resolution or MolProbity Score (Davis *et al.*, 2007): a structure was selected when its crystallographic resolution was better than in the redundant group by at least 0.2 Å, in case a few structures tied, the best was selected based on the MolProbity Score (Davis *et al.*, 2007).

Methods: Enhancement of the original k-NN protocol and discovery of new *NtC* classes.

Table S1. Definitions of the 44 identified *NtC* conformer classes: a) *NtC* annotations, the averages of torsion angle values, and occurrences of *NtC* in the 1,802 analyzed structures; b) estimated standard deviations of the torsion averages of *NtC*. PDB formatted coordinates of representative examples of all *NtC* can be found in a separate zip file and at dnatco.org.

Annotation	CANAs	<i>NtC</i>	1	2	3	4	5	6	7	8	9	Numbers of Steps in				golden set
			δ	ϵ	ζ	$\alpha 1$	$\beta 1$	$\gamma 1$	$\delta 1$	χ	$\chi 1$	prot/DNA		naked DNA		
			#	%	#	%	#	%	#	%	#	%	#	%		
the most frequent, "canonical" A-DNA (identical to A-RNA)	AAA	AA00	83	206	287	293	174	55	83	199	202	1 173	2.3	866	11.9	296
A-DNA with $\alpha 1/\gamma 1$ crank (150/180)	AAA	AA01	82	194	291	149	193	182	87	205	188	163	0.3	88	1.2	55
A-DNA with BI-like X/X1	AAA	AA02	88	203	275	294	161	54	88	244	245	554	1.1	63	0.9	168
A-DNA similar to AA01, high $\alpha 1$	AAA	AA03	83	214	273	330	165	26	86	202	215	16	0.0	14	0.2	9
A-DNA similar to AA01, low $\alpha 1$	AAA	AA04	80	198	301	255	175	91	83	202	192	36	0.1	32	0.4	14
A-to-B: δ C3'-endo, $\delta 1$ C2'-endo, X/X1 have A/B pattern	A-B	AB01	86	193	283	299	180	55	142	221	255	2 190	4.3	347	4.8	100
A-to-B: $\delta \sim$ O4'-endo, extremely low ϵ , X/X1 are B-like	A-B	AB02	93	59	56	208	187	65	133	240	252	116	0.2	1	0.0	18
A-to-B: similar to AB01 with high $\alpha 1$ and low ζ , $\beta 1$, $\gamma 1$	A-B	AB03	96	195	253	344	151	33	131	220	250	87	0.2	20	0.3	11
BI-to-A complex conformer with $\delta 1$ from C3' to C4'-exo, X/X1 are B-like	B-A	BA01	134	189	258	293	169	51	88	248	229	749	1.5	142	1.9	54
BI-to-A complex conformer with $\delta 1$ from C3' to O4'-endo, X/X1 are B-like	B-A	BA05	130	184	266	295	170	53	102	249	235	2083	4.1	394	5.4	107
B-to-A: $\beta 1 \sim$ 120, X1 A-like	B-A	BA08	141	212	188	302	129	55	85	271	211	92	0.2	34	0.5	12
BI-to-A: $\beta 1 \sim$ 60, $\alpha 1/\gamma 1$ crank (250/170), X1 A-like	B-A	BA09	135	199	287	254	71	168	86	265	187	133	0.3	3	0.0	13
B-to-A complex cluster: $\alpha 1 \sim$ 100, $\gamma 1 \sim$ 180, X1 A-like	B-A	BA10	135	200	218	105	229	194	88	258	195	203	0.4	6	0.1	43
BI-to-A complex cluster: $\alpha 1 \sim$ 60, $\gamma 1 \sim$ 180, X1 A-like	B-A	BA13	142	231	196	74	232	196	89	266	199	153	0.3	8	0.1	57
BI-to-A: X1 A-like	B-A	BA16	145	255	187	63	226	196	87	260	201	11	0.0	1	0.0	7
BI-to-A: $\delta 1 \sim$ O4'-endo, $\beta 1 \sim$ 120	B-A	BA17	148	255	177	294	130	44	96	272	234	106	0.2	28	0.4	27
the most frequent DNA conformer, "canonical" B form, also called BI	BBB	BB00	138	183	259	303	180	44	138	252	258	17 513	34.5	1 561	21.4	1 958
less populated variant of BI form	2B1	BB01	132	181	265	301	177	49	121	248	244	3 076	6.1	486	6.7	142
BI with $\alpha 1/\gamma 1$ crank (30/300)	3B1	BB02	140	194	246	32	194	297	150	252	253	2 369	4.7	47	0.6	134
BI with $\alpha 1/\gamma 1$ crank (170/170)	3B1	BB03	145	176	276	166	164	173	146	239	232	327	0.6	5	0.1	31
BI-to-BI conformer	B12	BB04	140	201	214	315	153	46	140	263	253	3 179	6.3	211	2.9	243
BI-to-BI with $\alpha 1/\gamma 1$ crank (70/230), X1 A-like	B12	BB05	143	219	202	68	232	220	124	269	209	107	0.2	2	0.0	20
BI form, typical by ϵ/ζ switch compared to BI	BB2	BB07	144	245	171	297	141	46	141	270	259	2 415	4.8	598	8.2	371
BI form, variant, $\alpha 1/\gamma 1$ crank (60/210)	BB2	BB08	144	249	195	63	229	211	144	261	231	106	0.2	5	0.1	19
B form with extremely low values of $\alpha 1$, $\beta 1$, $\gamma 1$	miB	BB10	139	195	191	23	106	19	129	257	258	877	1.7	15	0.2	49
B form with $\alpha 1/\gamma 1$ crank (120/180)	miB	BB11	144	200	199	121	226	189	143	258	222	238	0.5	2	0.0	22
BI with $\alpha 1/\gamma 1$ crank (250/170), low $\beta 1$ 70, X1 A-like	miB	BB12	140	196	287	248	73	172	144	263	212	414	0.8	3	0.0	18
BI with $\alpha 1/\gamma 1$ crank (210/160), low $\beta 1$ 100, X1 A-like	miB	BB13	143	186	291	216	104	161	147	252	219	179	0.4	3	0.0	17
B-form with extremely low ϵ	miB	BB14	121	104	303	229	260	73	132	264	263	109	0.2	0	0.0	8
BI with high $\alpha 1$ and $\gamma 1$ near 0	miB	BB15	149	186	262	340	194	354	149	251	262	313	0.6	31	0.4	17
complex cluster of B-like conformers: A-like X, bases may be unstacked to incorporate intercalated drug, occurs where backbone accommodates deformation (metal ion near, strand crossing in Holliday junctions, ends of duplexes)	miB	BB16	145	227	281	288	173	51	142	197	267	480	0.9	138	1.9	117
A-to-B-like but X1 syn, mostly in duplexes	SQX	AB15	91	214	280	295	176	56	139	238	67	17	0.0	0	0.0	14
BI-like but X syn, may be in duplex but many G-G in quadruplexes	SQX	BB51	146	189	275	294	174	52	135	62	261	145	0.3	158	2.2	47
G-G in quadruplexes, unusual $\beta 1$, $\gamma 1$, X1 syn	SQX	BB15	140	202	282	307	258	304	151	236	65	4	0.0	21	0.3	17
BI-like but X1 syn, mostly G-G in quadruplexes, $\alpha 1$ g+, $\gamma 1$ g-	SQX	BB25	137	196	225	33	187	295	145	257	70	7	0.0	8	0.1	8
part. unstacked T-G-G-T in quadruplexes, X1 syn, unusual ζ , $\alpha 1$, $\gamma 1$	SQX	NS15	143	206	61	82	204	192	146	242	68	5	0.0	15	0.2	13
start of loop in quadruplex or hairpin, G-X, untypical ζ , $\alpha 1$, $\gamma 1$	SQX	NS02	145	225	67	74	189	191	137	264	258	11	0.0	21	0.3	14
part. unstacked T-T in quadruplexes, unusual combination of ϵ , ζ , $\alpha 1$, $\gamma 1$, X1	SQX	NS03	143	294	111	153	197	53	151	262	185	5	0.0	11	0.2	13
unstacked, in 4-way junction, ϵ , ζ high values	SQX	NS04	140	275	279	304	191	56	150	266	210	2	0.0	14	0.2	12
5'-end of dsDNA, base open (unstacked), ζ , $\alpha 1 \sim$ 60	SQX	NS05	154	242	77	63	177	64	137	237	249	18	0.0	0	0.0	10
Z form, Y-R step	ZZZ	ZZ15	148	264	76	66	186	179	95	205	60	65	0.1	73	1.0	20
Z form, Y-R step, $\delta 1$ C2'-endo	ZZZ	ZZ25	141	263	71	78	179	185	148	208	77	33	0.1	38	0.5	41
ZI form, R-Y step	ZZZ	ZZ51	97	242	294	209	230	55	144	63	205	49	0.1	52	0.7	48
ZII form, R-Y step	ZZZ	ZZ52	95	186	63	169	162	44	144	58	213	7	0.0	12	0.2	17
Unassigned conformations	NAN	NANT	x	x	x	x	x	x	x	x	x	10 839	21.3	1 718	23.6	0
			50 774	100.0	7 295	100.0										4 431
Annotation	CANAs	<i>NtC</i>	δ	ϵ	ζ	$\alpha 1$	$\beta 1$	$\gamma 1$	$\delta 1$	χ	$\chi 1$	#	%	#	%	golden set
			1	2	3	4	5	6	7	8	9	prot/DNA		naked		set

Table S1. b) estimated standard deviations of the torsion averages of *NtC*.

		1	2	3	4	5	6	7	8	9	Quality of Golden set (*)	
CANAs	NtCs	esd(δ)	esd(ϵ)	esd(ζ)	esd(α 1)	esd(β 1)	esd(γ 1)	esd(δ 1)	esd(χ)	esd(χ 1)	rmsd of 9 torsions [°]	cartesian rmsd [Å]
		AAA	AA00	4.8	7.7	5.9	6.2	8.0	6.1	4.4	7.3	6.9
AAA	AA01	3.7	7.6	5.9	8.3	6.6	5.8	3.7	4.7	4.6	12.0	0.33
AAA	AA02	5.9	8.9	9.9	6.9	8.2	4.9	5.5	7.8	7.4	12.7	0.63
AAA	AA03	6.2	5.9	4.3	4.4	6.3	7.1	5.6	8.7	7.0	8.9	0.33
AAA	AA04	8.7	9.3	6.1	6.1	10.6	6.2	7.2	7.2	7.9	11.3	0.39
A-B	AB01	3.8	12.6	7.9	7.7	7.9	6.0	4.4	11.9	8.6	15.5	0.53
A-B	AB02	8.7	8.9	9.1	11.8	9.5	8.0	7.9	10.4	8.1	13.1	0.43
A-B	AB03	7.8	6.2	6.3	6.2	7.7	7.2	9.1	3.6	6.5	8.8	0.44
B-A	BA01	6.8	4.4	7.8	10.6	12.3	4.6	3.3	11.1	6.7	6.0	0.16
B-A	BA05	8.4	7.6	7.0	7.6	5.5	7.2	7.8	8.7	7.4	14.1	0.42
B-A	BA08	6.6	6.8	4.9	4.8	6.0	5.8	5.5	5.5	3.1	13.1	0.50
B-A	BA09	4.4	5.7	5.2	5.7	10.0	6.0	4.6	4.7	4.7	9.9	0.27
B-A	BA10	10.7	10.2	20.1	19.6	16.1	12.3	6.4	12.3	6.5	7.6	0.26
B-A	BA13	6.1	11.7	11.5	10.6	8.3	10.1	6.9	8.5	7.0	20.4	0.65
B-A	BA16	5.1	11.4	4.4	8.2	6.9	2.8	3.2	3.3	3.2	14.0	0.43
B-A	BA17	8.1	8.6	10.1	9.5	9.0	7.4	7.8	9.4	8.1	8.8	0.30
BBB	BB00	5.8	9.3	10.2	9.2	10.5	6.0	5.1	9.0	9.9	12.0	0.52
2B1	BB01	8.1	6.9	6.0	7.7	5.6	6.4	3.8	8.6	6.0	16.8	0.80
3B1	BB02	5.6	7.0	7.8	8.8	14.2	8.5	4.8	8.9	9.3	10.3	0.43
3B1	BB03	5.6	7.4	8.2	14.5	12.1	8.6	6.5	12.1	9.1	15.8	0.47
B12	BB04	4.4	9.5	9.0	8.1	9.0	6.4	4.5	8.9	8.6	14.5	0.57
B12	BB05	6.0	7.8	13.4	10.8	7.9	12.4	8.0	10.6	9.5	12.2	0.44
BB2	BB07	5.1	12.3	9.6	8.8	7.6	6.0	5.5	9.7	11.2	13.8	0.35
BB2	BB08	7.5	13.7	13.0	7.4	9.9	8.3	9.3	10.0	9.2	15.3	0.50
miB	BB10	5.2	10.3	9.2	11.4	9.0	9.6	7.5	10.3	9.6	13.2	0.47
miB	BB11	6.2	9.6	9.0	12.5	11.3	10.2	7.0	10.0	9.9	12.8	0.46
miB	BB12	4.6	6.0	11.7	13.7	3.8	4.2	4.4	8.0	9.0	13.5	0.52
miB	BB13	7.3	7.9	11.4	9.4	10.5	7.0	7.9	7.9	7.5	11.5	0.43
miB	BB14	6.0	6.8	5.6	7.2	4.2	8.6	4.6	8.7	5.9	11.9	0.46
miB	BB15	4.9	6.0	10.1	10.2	10.0	12.9	5.0	9.6	7.2	6.9	0.35
miB	BB16	7.4	12.8	11.2	8.7	10.8	8.0	5.3	9.3	18.4	13.3	0.44
SQX	AB1S	2.4	3.3	4.4	4.6	4.4	3.6	4.9	4.2	3.7	16.0	0.73
SQX	BB51	5.1	8.7	7.7	9.6	6.9	7.7	6.8	9.4	10.8	9.2	0.30
SQX	BB1S	6.6	9.3	6.8	5.6	6.7	4.9	3.5	12.7	3.9	5.5	0.17
SQX	BB2S	2.6	3.1	2.3	6.4	3.7	3.1	1.9	4.8	2.6	14.2	0.45
SQX	NS1S	2.5	6.3	7.0	7.9	6.0	6.1	4.8	4.5	4.7	12.2	0.80
SQX	NS02	2.8	9.3	6.6	12.1	7.9	7.7	9.1	12.5	8.3	8.1	0.22
SQX	NS03	3.2	3.9	6.2	8.5	4.7	6.4	4.4	5.9	2.3	8.7	0.28
SQX	NS04	3.5	6.2	7.3	9.3	7.9	7.1	2.6	6.4	3.9	6.7	0.13
SQX	NS05	2.1	2.0	4.0	5.3	2.2	4.2	1.3	4.7	3.1	7.5	0.27
ZZZ	ZZ1S	3.5	5.3	4.0	3.8	4.4	3.5	5.0	5.5	6.1	7.0	0.18
ZZZ	ZZ2S	4.5	5.5	6.2	7.0	6.7	4.7	6.6	6.8	4.7	10.1	0.30
ZZZ	ZZS1	5.8	7.2	8.7	6.9	10.1	5.8	4.9	4.9	6.4	12.3	0.31
ZZZ	ZZS2	2.0	5.6	6.4	6.0	6.6	5.1	2.9	4.3	4.6	6.8	0.19
NAN	NANT	x	x	x	x	x	x	x	x	x		
CANAs	NtCs	esd(δ)	esd(ϵ)	esd(ζ)	esd(α 1)	esd(β 1)	esd(γ 1)	esd(δ 1)	esd(χ)	esd(χ 1)		
		1	2	3	4	5	6	7	8	9		

(*) "rmsd of 9 torsions" is rmsd measured in torsion space between the mean and most distant member of the golden set; "cartesian rmsd" is rmsd between 18 atoms of the dinucleotide that define the nine torsions

Conformational assignment of a dinucleotide

Algorithm assigning the *NtC* conformational classes is schematically shown in a flowchart in Figure S1. The assignment process is summarized below.

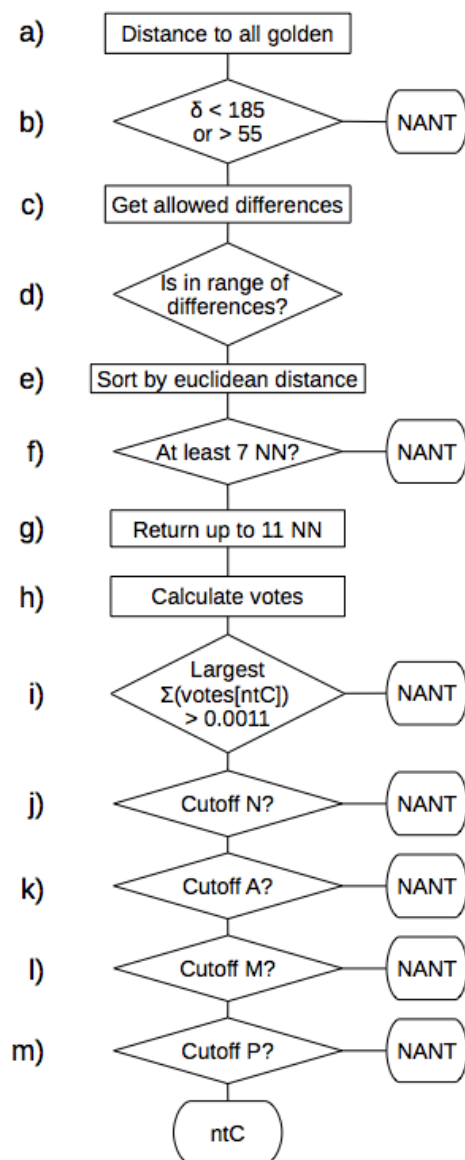


Figure S1. Schema of the algorithm assigning *NtC* classes to DNA dinucleotides.

a) In the initial step, distances are calculated between candidate dinucleotide and all dinucleotides in the golden set. All the distance, angular average and estimated standard deviation (ESD) formulas take into account the circularity (0-360°) of the data.

b) Steps with delta torsions outside the physically possible region are marked as unassigned conformer (NANT).

- c) The allowed difference from the average is calculated as 5 times the ESD of the respective torsion within the known *NtC* class (ESD and class average are calculated for the golden set using functions from the R package <http://www.R-project.org/>).
- d) The distances between the to-be-assigned dinucleotide and the golden set members are then tested if they are within the allowed limit.
- e) Sort the *NtC* classes satisfying condition d) by their Euclidean distances from the to-be-assigned dinucleotide.
- f) First up to 11 nearest neighbors are stored.
- g) At least 7 neighbors are necessary to assign a *NtC* conformer.
- h) For each *NtC* class a sum of votes is calculated (as $1/(\text{Euclidean distance})^2$).
- i) If the maximal sum exceeds 0.0011 ($1/(30^2)$), the step is a candidate for the conformer class, subject to the following tests for final classification.
- j) Each torsion must lie within 28° cutoff of a corresponding torsion of the nearest neighbor.
- k) Each torsion must lie within 28° cutoff of an average of corresponding torsion over all nearest neighbors (7 to 11 neighbors).
- l) The sum of (signed) differences from the candidate cluster average for first seven torsions (δ to δ_7) as well as sum of torsions 8 and 9 (both χ torsions) must lie within $\pm 60^\circ$. The value of $\pm 60^\circ$ was selected based on the analysis of sums of torsions from the data assigned without this test (Figure S3) in order to minimize false assignment of steps with systematically shifted torsion angles.

Verification of dinucleotide membership to the golden set. The data in the *golden set* were verified for self-consistency by comparing each member of the original *golden set* to the rest; ESD and averages of remaining data were calculated on the fly. If the step was assigned to a different *NtC* class than to the class it obtained in the original *golden set*, it was removed. The procedure was repeated until self-consistent golden set was obtained. Both cutoffs used in steps j) and k) above were set to a more stringent value of 20° for this purpose.

To summarize, the definition of the *NtC* classes and the algorithm of their assignment is not a result of a standard statistical procedure. All nine dinucleotide torsions have non-normal overlapping distributions that must be analyzed simultaneously in a complex 9-D space using noisy experimental data. The data are often of limited quality and quantity, even when we make use of virtually the complete set of non-redundant DNA structures available at the date of retrieval. The complexity of the 9-D torsional space required combining formal mathematical and statistical tools with knowledge-based empirical rules and involved human intervention at several critical steps.

Numbers of dinucleotide steps in the golden set as a function of the crystallographic resolution

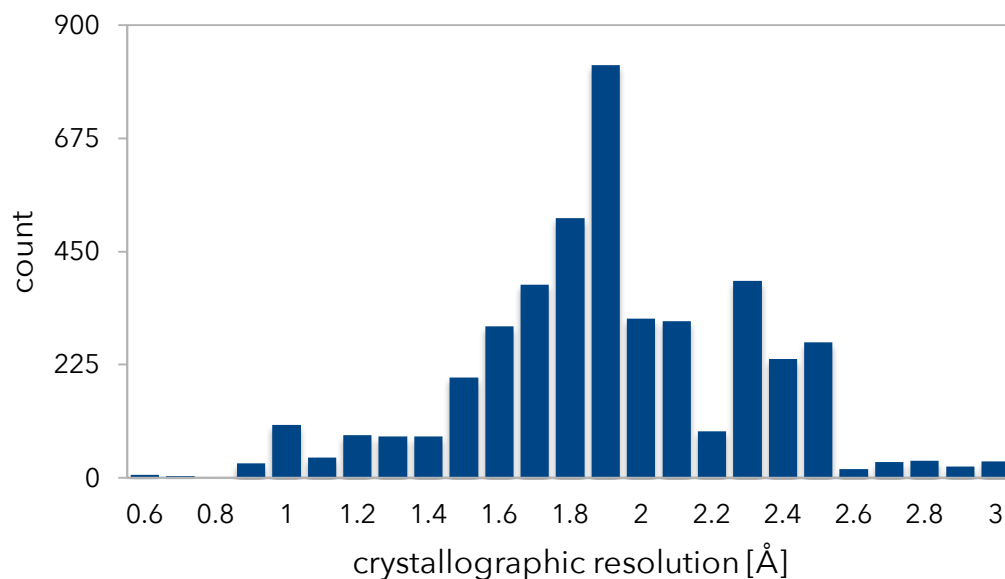


Figure S2. Histogram of numbers of dinucleotide steps in the *golden set* as a function of crystallographic resolution.

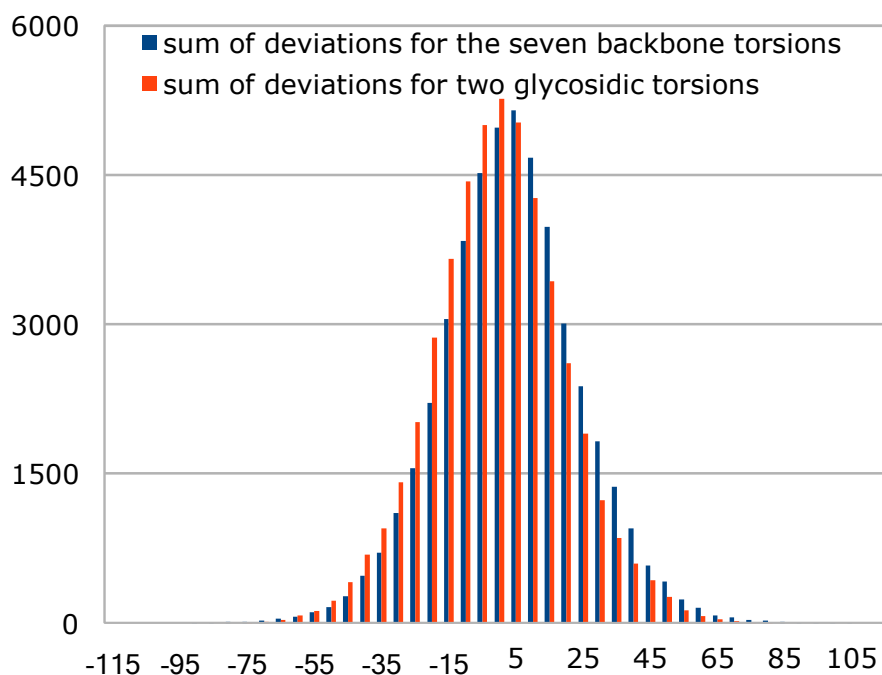


Figure S3. Histogram of sums of differences between the average of a particular *NtC* and the actual torsion values for dinucleotides assigned to this *NtC* in the *golden set*. Blue bars show differences of the backbone torsions $\delta, \epsilon, \zeta, \alpha_1, \beta_1, \gamma_1, \delta_1$, the red ones of both glycosidic torsions χ and χ_1 . Both distributions are Gaussian and have about the same spread. Numbers of dinucleotides with deviations larger than $\pm 60^\circ$ is negligible.

Results

AAA: A-form. The main conformational feature of the A-form is the sugar pucker in the C3'-*endo* region and the corresponding δ torsion values 80°-90°. The most frequent A-DNA conformer described here as *NtC AA00* is also characterized by a low *anti* glycosidic torsion χ near 200°. It is structurally identical to the A-RNA. *NtC AA01* differs from “the canonical” *AA00* by a typical “crankshaft motion” switch of torsion angles α and γ from values 300°/60° to 150°/180°. *AA02* is close to the canonical *AA00*, but its glycosidic torsions χ have B-like values near 250°.

A-B: conformers bridging A- to B-form. The main A-B conformer, *AB01*, has been described previously (Svozil et al., 2008) and is characterized by an A-like sugar pucker C3'-*endo* region ($\delta \sim 85^\circ$) and by values of the glycosidic torsion χ near 230° in its first (5'-) nucleotide, and B-like values of torsions α_1 , β_1 , γ_1 , δ_1 , and χ_1 in the second nucleotide. *AB01* is complemented by a newly characterized conformer *AB02*. It is interesting by its extremely low value of torsion ϵ near 60°, which is compensated by a switch of the following torsion ζ from a typical value near 300° to 60°. Despite the exotic value of ϵ , *AB02* is observed relatively frequently, 164 times in more than 100 structures, mostly in double helices of naked or complexed DNA, but also in hairpin stems (3dsd_C_DC17_DT18 (Williams et al., 2008)) and in unpaired strand ends (4gz2_C_DC1_DA2 (Schellenberg et al., 2012)).

Analysis of the stability of conformer AB02. The unprecedentedly low value of ϵ torsion near 60° prompted us to investigate whether this conformation is energetically stable as an isolated molecule or whether it is stabilized by intermolecular interactions in the crystal phase. We therefore performed quantum mechanical calculations of the internal potential energy of this conformer by scanning the ϵ torsion of the dinucleotide DG105 - DG106 of chain P from the high-resolution structure 4fj7 (Xia et al., 2013) between 30° and 80°. The results of the computations, which are detailed in Table S2 and the main text Figure 4, showed a near-harmonic potential with minimal energy near ϵ of 55° demonstrating that the *AB02* conformer is locally stable as an isolated molecule.

Table S2. Torsion values (in degrees) in the *NtC AB02*, a representative dinucleotide from structure 4fj7 (Xia et al., 2013), and after quantum mechanical optimization by DFT.

	δ	ϵ	ζ	α	β	γ	δ_1	χ	χ_1
<i>AB02</i> average	92	61	59	208	185	64	133	237	253
4FJ7_P_DG105_DG106	91	60	48	209	189	68	136	242	251
energy-optimized torsions	83	55	69	210	174	61	129	242	251

The hydrogens were added using *openbabel* and their positions optimized while keeping all the positions of heavy atoms frozen resulting in a dinucleotide with the overall charge of -2 (-1 at each phosphate group). During the scans, the position of heavy atoms of both guanine bases and C1' atoms were forced frozen, while positions of the remaining heavy atoms of the sugar phosphate backbone and of all hydrogen atoms were optimized. The solvation effects

were included using the COSMO model with a value of 78.4 for the solvent permittivity. To compute intramolecular energy of the step, we used dispersion corrected DFT functional TPSS (Tao *et al.*, 2003) and TZVP (Schafer *et al.*, 1994) augmented with empirical dispersion correction (Jurecka *et al.*, 2007) employing the TurboMole package of programs (Ahlich *et al.*, 1989).

B1A and B2A: conformers bridging B- to A-form. B-to-A conformers represent a structural counterpart of the previous *CANA* letter, *A-B*. The *B1A* and *B2A* letters are however represented by a much more diverse and numerous group of conformers than the *A-B* letter. The eight B-A conformers have the first sugar in the C2'-*endo* pucker and ϵ/ζ combination typical of either BI (180°/260°, *B1A* letter) or BII conformation (250°/170°, *B2A*). The sugar pucker of the second nucleotide are either in a typical C3'-*endo* or, in minority, O4'-*endo* pucker with $\delta 1$ near 100°. Conformationally soft glycosidic torsions correlate with the sugar pucker as expected: while χ is B-like, $\chi 1$ is lower and more A-like. Some of the B-to-A conformers have less common combinations of torsions ζ , $\alpha 1$, $\beta 1$, and $\gamma 1$ including $\alpha 1/\gamma 1$ crankshaft in *BA10* and *BA13*, or an extremely small value of β in *BA09*. All B-to-A conformers occur in duplexes but they can also occur in quadruplexes (*BA13* in 3sc8, 3sc8_A_DG14_DG15 (Collie *et al.*, 2012)) or joints of Holliday junctions (*BA05* in 1p4e, 1p4e_J_DT15_DT16 (Chen & Rice, 2003)).

Conformers of the B-form. What is usually described as “B-DNA form” should be seen as a family of conformations rather than a single, firmly defined structure. Neither do the two traditionally recognized B forms, BI and BII, suffice to describe the conformational diversity of the B-like backbone. In the structural alphabet *CANA*, we describe the B-form by three letters for BI conformers, one letter for BII conformers, and one letter for transitional BI-to-BII conformers. In addition, we grouped numerically minor but structurally diverse conformers that still bear some features of the B-form, under the letter *miB*.

BBB, 2B1, and 3B1: BI conformers. The dominant DNA conformer is a BI *NtC BB00*. It alone constitutes letter *BBB* because it represents about one third of all analyzed steps and should be viewed as the canonical B-form. It has both sugars in the C2'-*endo* pucker, values of both glycosidic torsions χ high *anti* near 250°, and displays a characteristic pattern of ϵ , ζ , $\alpha 1$, $\beta 1$, $\gamma 1$: t/g-/g-/t/g+ (180°, 260°, 300°, 180°, 40°). The related BI conformer *BB01* represents a slight but distinct and highly populated variation of *BB00*. Less populated conformers *BB02* and *BB03* (*CANA 3B1*) are characterized by a switch of the $\alpha 1/\gamma 1$ values from the prevailing 300°/40° to 30°/300° and 170°/170°, respectively.

Dinucleotides whose conformations belong to the letters *BBB*, *2B1*, and *3B1* form the core of the Watson-Crick-paired double helices, but they can occur in almost any structural context including quadruplexes, or joints of Holliday junctions. This adaptability to various local constraints further demonstrates the role of BI as the main B-DNA conformation.

BB2 and B12: BII and BI-to-BII conformers. The main characteristics telling apart BI and BII forms is a switch of values of ϵ and ζ torsions from 180°/260° in BI to about 250°/180° in BII. The most frequent conformer epitomizing BII form is *BB07*, also typical by a low value of torsion $\beta 1$, 140°. The other BII conformer, *BB08*, in addition has $\alpha 1/\gamma 1$ values switched to ~60°/210°. The main BI-to-BII conformer, *BB04*, has torsion angle values indeed half-way between those of BI *BB00* and BII *BB07*.

A compensatory switch of $\alpha 1$ and $\gamma 1$ torsions from their most frequently observed combination of 300°/50° is a common local variation of the backbone. Observed are combinations (i) 30°/300° in highly populated *NtC BB02*; (ii) 60°/210° in BII conformer *BB08* and BI-to-BII *BB05*; (iii) 150°/180° in the “classical” crankshaft motion A-DNA modification in *AA01* and in BI *BB03*. In all these cases, the low α value correlates with β

well above the usual value of 180°.

miB: various “minor” B-forms. B-forms exist in several distinct conformers that share typical conformational features of the B-form: both sugars in the C2'-*endo* pucker and both glycosidic torsions χ above 200°. Some of these ‘minor’ forms are actually fairly common: *BB10* represents 1.6 %, *BB12* 0.8 %, and *BB16* 1.3 % of the analyzed steps. The structural features of these conformers can be quite exotic, but they do not disrupt base stacking and are compatible with incorporation into a double helix. Conformer *BB10* has extremely low values of α , β , γ : 20°/100°/20°, but it occurs quite frequently in both naked DNA (458d_A_DC1_DG2 (Aymami *et al.*, 1999)) and in ~300 protein complexes, e.g. 3cro_A_DA5_DC6 (Mondragon & Harrison, 1991), and in 4kgc (4kgc_I_DA-31_DA-30 (Adhireksan *et al.*, 2014)), where it was detected 36 times. Even more extreme γ values near 0° are observed in *BB15*, but even this *NtC* does not disrupt the double helical architecture and participates in W-C pairing in naked DNA (3gqc_G_DT10_DA11 (Swan *et al.*, 2009)), DNA with a drug bound in the minor groove (netropsin in 1d86, 1d86_B_DT20_DC21 (Sriram *et al.*, 1992)), as well as in a protein-bound DNA (3lz1_J_DC18_DC19 (Vasudevan *et al.*, 2010)). *BB15* also occurs in the central region of guanine quadruplexes (1k8p_B_DG22_DG23 (Parkinson *et al.*, 2002)). *BB14* is similar to *AB02*, which was discussed above, by its extremely low value of ϵ torsion, and is fully compatible with the W-C pairing. Contrary to *AB02*, it is frequently found in nucleosomes.

NtC BB16 is a complex B-like conformer without any extreme torsion value, which is, however, important by the ability to accommodate a large local deformation of the strand. A particular combination of torsions attributable to this conformer can lead to unstacking of the bases to incorporate an intercalated drug (1nab_A_DC1_DG2 (Temperini *et al.*, 2003)), 308d_A_DC1_DG2 (Gao *et al.*, 1997)), or to stack a base between two bases of the complementary strand (1g38_C_DT705_DC706 (Goedecke *et al.*, 2001)). *BB16* is also often found at or near strand crossing in Holliday junctions (114j_D_DC37_DC38 (Thorpe *et al.*, 2003)), near a metal cation (e.g. mutually close 3 Mg⁺⁺ in 1mow_B_DC312_DA313 (Chevalier *et al.*, 2002)), at bends of histone complexes (5f99_J_DA-70_DA-69 (Frouws *et al.*, 2016)), or in a hairpin loop (2cdm_B_DT23_DC24 (Boer *et al.*, 2006)).

SQX: conformers occurring mainly in non-duplex DNA. Conformers from this structurally variable group are rare but important for the architecture of folded forms of DNA, because dinucleotides in these conformations are found in unpaired or mismatched parts of DNA duplexes or in non-duplex DNA structures. *NtC ABIS*, *BBS1*, *BB2S*, and *NSIS* occur in guanine tetraplexes, *BBIS* and *BB2S* are actually observed only in their G-G steps (1jpb_A_DT1008_DG1009 (Haider *et al.*, 2002)). The structural similarity of *NSIS* and *NS02* explains their similar structural role: both form a bridge between the loop and the G-quartet of a tetraplex. *NSIS* is found almost exclusively entering the G-quartet by the TG sequence (1jpb_A_DT1008_DG1009), and *NS02* leaving it by GT (4fxm_A_DG10_DT11 (Nicoludis *et al.*, 2012)). *NS02* can, however, be found in other than tetraplex structures such as a hairpin loop (3c2p_D_DA16_DA17 (Gleghorn *et al.*, 2008)), or in a topologically complex self-folding loop in 4f4y_P_DG13_DC14 (Wilson *et al.*, 2013). *BBS1* also forms the G-quartet core of tetraplex structures (1jpb_A_DG1003_DG1004), but its structural role is much wider. It accommodates a purine-purine mismatch (178d_B_8OG21_DG22 (McAuley-Hecht *et al.*, 1994)), occurs in loops of complicated self-folding ssDNA hairpin structures (2vju_C_DA-20_DG-19 (Barabas *et al.*, 2008)), or in an unusual duplex linking Z-DNA ends with T-T mismatches in the duplex midpoint (4l25_B_DG4_DA5 (Kondo *et al.*, 2014)). *BBS1*, together with another unusual conformer, *ABIS*, is involved in forming non-Watson-Crick C-G pairs in TATA box DNA bound to the TBP protein (e.g. 1qn7_E_DG210_DG211, 1qn3_E_DC209_DG210 (Patikoglou *et al.*, 1999)). The DNA duplex resembles A-form in

these complexes but, as can be shown by a detailed conformational analysis at dnatco.org, it is severely deformed at several places. Suboptimal hydrogen bonding by the guanine Hoogsteen edge to the cytosine C-H edge then probably releases strain of the deformed duplex. *ABIS* may accommodate steric strain in duplexes caused by mismatched pairs such as G-G mismatch in 1nk4 (1nk4_B_DG28_DG29 (Johnson & Beese, 2004)), or at strands opposing an abasic site of DNA lesions (3cvv (Glas *et al.*, 2009)). The last conformer belonging to the *SQX* letter, *NS05*, displays RNA-like “open” conformation with completely unstacked bases. However, the general validity of this conformer should be taken with caution as all known examples are TC steps originating from DNA complexes with DNA polymerase *rb69* (3nci_T_DT1_DC2 (Wang *et al.*, 2011)).

ZZZ: Z-form conformers. The four conformers building the left handed Z-DNA duplex are so called ZI (*ZZS1*) and ZII (*ZZS2*) forms of the purine-pyrimidine steps. The alternating pyrimidine-purine steps are also described by two conformers, the major *ZZIS*, and the minor *ZZS2* with $\delta 1$ in the C2'-*endo* pucker, which can occur only at the strand ends. In all *ZZZ* conformers, a base in the *syn* orientation is always purine.

NAN: non-assigned conformers. About 18 % of the analyzed steps end up as unassigned to any of the existing *NtC*. Although the assignment process can be tuned to classify a higher number of dinucleotides by loosening parameters of the currently used protocol, we consider it representing a realistic compromise between the assignment accuracy and the complex nature of the DNA conformational space.

Annotation of selected prototypical DNA structures.

Table S3. The assignment of the *NtC* classes to selected crystal, NMR, and MD dodecamer structures. a) *NtC* classes of structures appearing in Figure 6 in the main text. The assignment is also available at the web site dnatco.org.

CANA assignment for structures shown in Figure 6

Conner, B.N.: JMB 174: 663-695, '84		Drew, H.R.: PNAS 78: 2179-2183, '81		Harper, A.: Acta D 54: 1273-1284, '98		Chen, L.: BCH 33: 13540-13546, '94	
step	ntC	step	ntC	step	ntC	step	ntC
1ana_A_C381_DC2	AA00	1bna_A_DC1_DG2	BB04	362d_A_DT1_DG2	ZZ1S	190d_A_DC1_DC2	AA02
1ana_A_DC2_DG3	AA00	1bna_A_DG2_DC3	BA05	362d_A_DG2_DC3	ZZS2	190d_A_DC2_DC3	NANT
1ana_A_DG3_DG4	AB01	1bna_A_DC3_DG4	AB01	362d_A_DC3_DG4	ZZ1S	190d_A_DC3_DC4	NANT
1ana_B_C385_DC6	AA00	1bna_A_DG4_DA5	BB04	362d_A_DG4_DC5	ZZS1	190d_B_DC5_DC6	NANT
1ana_B_DC6_DG7	AA00	1bna_A_DA5_DA6	BB01	362d_A_DC5_DA6	ZZ1S	190d_B_DC6_DC7	NANT
1ana_B_DG7_DG8	AA00	1bna_A_DA6_DT7	BA05	362d_B_DT7_DG8	ZZ1S	190d_B_DC7_DC8	NANT
		1bna_A_DT7_DT8	BA05	362d_B_DG8_DC9	ZZS2	190d_C_DC9_DC10	NANT
		1bna_A_DT8_DC9	BB01	362d_B_DC9_DG10	ZZ1S	190d_C_DC10_DC11	AA02
		1bna_A_DC9_DG10	BB00	362d_B_DG10_DC11	ZZS1	190d_C_DC11_DC12	NANT
		1bna_A_DG10_DC11	BB07	362d_B_DC11_DA12	ZZS2	190d_D_DC13_DC14	AA02
		1bna_A_DC11_DG12	BA05			190d_D_DC14_DC15	AA02
		1bna_B_DC13_DG14	BB00			190d_D_DC15_DC16	AB01
		1bna_B_DG14_DC15	BA05				
		1bna_B_DC15_DG16	AB01				
		1bna_B_DG16_DA17	BB00				
		1bna_B_DA17_DA18	BB00				
		1bna_B_DA18_DT19	BA05				
		1bna_B_DT19_DT20	BB01				
		1bna_B_DT20_DC21	BB01				
		1bna_B_DC21_DG22	BB00				
		1bna_B_DG22_DC23	BA17				
		1bna_B_DC23_DG24	BA05				
Haider, S.: JMB 320: 189-200, '02		Eichman, B.F.: PNAS 97: 3971-3976, '00					
step	ntC	step	ntC	step	ntC		
1jppq_A_DG1001_DG1002	BBS1	1dcw_A_DC1_DC2	BB16	1dcw_C_DC1_DC2	BB16		
1jppq_A_DG1002_DG1003	BB2S	1dcw_A_DC2_DG3	BB07	1dcw_C_DC2_DG3	BB07		
1jppq_A_DG1003_DG1004	BBS1	1dcw_A_DG3_DG4	BB07	1dcw_C_DG3_DG4	BB07		
1jppq_A_DG1004_BRU1005	BA05	1dcw_A_DG4_DT5	BB00	1dcw_C_DG4_DT5	BB00		
1jppq_A_BRU1005_DT1006	AB01	1dcw_A_DT5_DA6	NANT	1dcw_C_DT5_DA6	BB04		
1jppq_A_DT1006_DT1007	NS03	1dcw_A_DA6_DC7	BB07	1dcw_C_DA6_DC7	BB07		
1jppq_A_DT1007_DT1008	BB16	1dcw_A_DC7_DC8	BB00	1dcw_C_DC7_DC8	BB00		
1jppq_A_DT1008_DG1009	NS1S	1dcw_A_DC8_DG9	BB07	1dcw_C_DC8_DG9	BB07		
1jppq_A_DG1009_DG1010	BBS1	1dcw_A_DG9_DG10	BB04	1dcw_C_DG9_DG10	BB04		
1jppq_A_DG1010_DG1011	BB1S	1dcw_B_DC1_DC2	BB16	1dcw_D_DC1_DC2	BB16		
1jppq_A_DG1011_DG1012	BBS1	1dcw_B_DC2_DG3	BB07	1dcw_D_DC2_DG3	BB07		
1jppq_B_DG2001_DG2002	BBS1	1dcw_B_DG3_DG4	BB07	1dcw_D_DG3_DG4	BB07		
1jppq_B_DG2002_DG2003	BB2S	1dcw_B_DG4_DT5	BB00	1dcw_D_DG4_DT5	BB00		
1jppq_B_DG2003_DG2004	BBS1	1dcw_B_DT5_DA6	BB04	1dcw_D_DT5_DA6	BB04		
1jppq_B_DG2004_BRU2005	BA05	1dcw_B_DA6_DC7	NS04	1dcw_D_DA6_DC7	NS04		
1jppq_B_BRU2005_DT2006	AB01	1dcw_B_DC7_DC8	BB16	1dcw_D_DC7_DC8	BB16		
1jppq_B_DT2006_DT2007	NS03	1dcw_B_DC8_DG9	BB07	1dcw_D_DC8_DG9	BB07		
1jppq_B_DT2007_DT2008	BB16	1dcw_B_DG9_DG10	BB07	1dcw_D_DG9_DG10	BB07		
1jppq_B_DT2008_DG2009	NS1S						
1jppq_B_DG2009_DG2010	NANT						
1jppq_B_DG2010_DG2011	BB1S						
1jppq_B_DG2011_DG2012	BBS1						

Table S3. The assignment of the *NtC* classes to selected DNA structures. b) *NtC* classes of crystal, NMR, and MD structures of oligonucleotides with sequence of the Dickerson-Drew dodecamer. The assignment is also available at the web site dnatco.org.

Selected X-ray structures

Drew, H.R., PNAS 78: 2179-2183, '81		Drew, H.R., PNAS 79: 4040-4044, '82		Holbrook, S.R., Acta B 41: 255-262, '85		Westhof, E., JBSD 5: 581-600, '87	
step	Ddnc	step	Ddnc	step	Ddnc	step	Ddnc
1bna_A_DC1_DG2	BB04	2bna_A_DC1_DG2	BB04	7bna_A_DC1_DG2	NANT	9bna_A_DC1_DG2	BB01
1bna_A_DG2_DC3	BA05	2bna_A_DG2_DC3	BA01	7bna_A_DG2_DC3	BA05	9bna_A_DG2_DC3	BA01
1bna_A_DC3_DG4	AB01	2bna_A_DC3_DG4	AB01	7bna_A_DC3_DG4	AB01	9bna_A_DC3_DG4	AB01
1bna_A_DG4_DA5	BB04	2bna_A_DG4_DA5	BB04	7bna_A_DG4_DA5	BB04	9bna_A_DG4_DA5	BB00
1bna_A_DA5_DA6	BB01	2bna_A_DA5_DA6	BB00	7bna_A_DA5_DA6	BB01	9bna_A_DA5_DA6	BB01
1bna_A_DA6_DT7	BA05	2bna_A_DA6_DT7	BA05	7bna_A_DA6_DT7	BB01	9bna_A_DA6_DT7	BA05
1bna_A_DT7_DT8	BA05	2bna_A_DT7_DT8	BB01	7bna_A_DT7_DT8	BB01	9bna_A_DT7_DT8	BB01
1bna_A_DT8_DC9	BB01	2bna_A_DT8_DC9	BB01	7bna_A_DT8_DC9	BB00	9bna_A_DT8_DC9	BB00
1bna_A_DC9_DG10	BB00	2bna_A_DC9_DG10	BB04	7bna_A_DC9_DG10	BB15	9bna_A_DC9_DG10	BB00
1bna_A_DG10_DC11	BB07	2bna_A_DG10_DC11	BB07	7bna_A_DG10_DC11	BB07	9bna_A_DG10_DC11	NANT
1bna_A_DC11_DG12	BA05	2bna_A_DC11_DG12	BA05	7bna_A_DC11_DG12	BA05	9bna_A_DC11_DG12	BB00
1bna_B_DC13_DG14	BB00	2bna_B_DC13_DG14	BB04	7bna_B_DC13_DG14	BB00	9bna_B_DC13_DG14	BB00
1bna_B_DG14_DC15	BA05	2bna_B_DG14_DC15	BA05	7bna_B_DG14_DC15	BA05	9bna_B_DG14_DC15	BA05
1bna_B_DC15_DG16	AB01	2bna_B_DC15_DG16	AB01	7bna_B_DC15_DG16	AB01	9bna_B_DC15_DG16	AB01
1bna_B_DG16_DA17	BB00	2bna_B_DG16_DA17	BB00	7bna_B_DG16_DA17	BB00	9bna_B_DG16_DA17	BB00
1bna_B_DA17_DA18	BB00	2bna_B_DA17_DA18	BB00	7bna_B_DA17_DA18	BB00	9bna_B_DA17_DA18	BA05
1bna_B_DA18_DT19	BA05	2bna_B_DA18_DT19	BB01	7bna_B_DA18_DT19	BB01	9bna_B_DA18_DT19	BA05
1bna_B_DT19_DT20	BB01	2bna_B_DT19_DT20	BB00	7bna_B_DT19_DT20	BB01	9bna_B_DT19_DT20	BB01
1bna_B_DT20_DC21	BB01	2bna_B_DT20_DC21	BB01	7bna_B_DT20_DC21	BB01	9bna_B_DT20_DC21	BB01
1bna_B_DC21_DG22	BB00	2bna_B_DC21_DG22	BB00	7bna_B_DC21_DG22	BB00	9bna_B_DC21_DG22	BB00
1bna_B_DG22_DC23	BA17	2bna_B_DG22_DC23	BA17	7bna_B_DG22_DC23	BB07	9bna_B_DG22_DC23	BB07
1bna_B_DC23_DG24	BA05	2bna_B_DC23_DG24	AA00	7bna_B_DC23_DG24	BA01	9bna_B_DC23_DG24	BB01
Aymami, J., PNAS 87: 2526-2530, '90		Aymami, J., PNAS 87: 2526-2530, '90		Chiu, T.K., JMB 301: 915-945, '00		Chiu, T.K., JMB 301: 915-945, '00	
step	Ddnc	step	Ddnc	step	Ddnc	step	Ddnc
1vte_A_DC1_DG2	NANT	1ndn_A_DC1_DG2	NANT	1en3_A_DC1_DC2	BB00	1en8_A_DC1_DC2	AB01
1vte_A_DG2_DC3	NANT	1ndn_A_DG2_DC3	NANT	1en3_A_DC2_DA3	BB07	1en8_A_DC2_DA3	BB07
1vte_A_DC3_DG4	NANT	1ndn_A_DC3_DG4	BB10	1en3_A_DA3_DA4	BA05	1en8_A_DA3_DA4	BB00
1vte_A_DG4_DA5	NANT	1ndn_A_DG4_DA5	BB10	1en3_A_DA4_DC5	BB01	1en8_A_DA4_DC5	BB00
1vte_A_DA5_DA6	NANT	1ndn_A_DA5_DA6	NANT	1en3_A_DC5_DG6	BB00	1en8_A_DC5_DG6	BB07
1vte_A_DA6_DA7	NANT	1ndn_A_DA6_DA7	NANT	1en3_A_DG6_DT7	BB01	1en8_A_DG6_DT7	BB01
1vte_A_DA7_DA8	BB00	1ndn_A_DA7_DA8	NANT	1en3_A_DT7_DT8	BB01	1en8_A_DT7_DT8	BB00
1vte_A_DA8_DC9	BB02	1ndn_A_DA8_DC9	NANT	1en3_A_DT8_DG9	BB07	1en8_A_DT8_DG9	BB07
1vte_A_DC9_DG10	NANT	1ndn_A_DC9_DG10	NANT	1en3_A_DG9_DG10	BB00	1en8_A_DG9_DG10	BA05
1vte_A_DG10_DC11	AB01	1ndn_A_DG10_DC11	NANT				
1vte_A_DC11_DG12	NANT	1ndn_A_DC11_DG12	AA02				
1vte_B_DC13_DG14	NANT	1ndn_B_DC13_DG14	NANT	1en3_A_DC1_DC2	BB00	1en8_A_DC1_DC2	AB01
1vte_B_DG14_DC15	NANT	1ndn_B_DG14_DC15	NANT	1en3_A_DC2_DA3	BB07	1en8_A_DC2_DA3	BB07
1vte_B_DC15_DG16	NANT	1ndn_B_DC15_DG16	NANT	1en3_A_DA3_DA4	BA05	1en8_A_DA3_DA4	BB00
1vte_B_DG16_DT17	NANT	1ndn_B_DG16_DT17	NANT	1en3_A_DA4_DC5	BB01	1en8_A_DA4_DC5	BB00
1vte_B_DT17_DT18	NANT	1ndn_B_DT17_DT18	NANT	1en3_A_DC5_DG6	BB00	1en8_A_DC5_DG6	BB07
				1en3_A_DG6_DT7	BB01	1en8_A_DG6_DT7	BB01
1vte_C_DT19_DT20	NANT	1ndn_C_DT19_DT20	NANT	1en3_A_DT7_DT8	BB01	1en8_A_DT7_DT8	BB00
1vte_C_DT20_DC21	AA01	1ndn_C_DT20_DC21	BB02	1en3_A_DT8_DG9	BB07	1en8_A_DT8_DG9	BB07
1vte_C_DC21_DG22	NANT	1ndn_C_DC21_DG22	NANT	1en3_A_DG9_DG10	BB00	1en8_A_DG9_DG10	BA05
1vte_C_DG22_DC23	NANT	1ndn_C_DG22_DC23	NANT				
1vte_C_DC23_DG24	NANT	1ndn_C_DC23_DG24	NANT				

Chiu, T.K., JMB 301: 915-945, '00		Chiu, T.K., JMB 301: 915-945, '00		Sines, C.C., JACS 122: 11048, '00		Liu, J., JBC 274: 24749-24752, '99	
step	Ddnc	step	Ddnc	step	Ddnc	step	Ddnc
1en9_A_DC1_DC2	BB00	1ene_A_DC1_DC2	BB00	1fq2_A_DC1_DG2	BB00		
1en9_A_DC2_DA3	BB07	1ene_A_DC2_DA3	BB07	1fq2_A_DG2_DC3	NANT	463d_A_DG2_DC3	BA01
1en9_A_DA3_DG4	BB00	1ene_A_DA3_DG4	BB00	1fq2_A_DC3_DG4	AB01	463d_A_DC3_DG4	AB01
1en9_A_DG4_DC5	BB07	1ene_A_DG4_DC5	BB07	1fq2_A_DG4_DA5	BB00	463d_A_DG4_DA5	BB00
1en9_A_DC5_DG6	BB00	1ene_A_DC5_DG6	BB00	1fq2_A_DA5_DA6	BB01	463d_A_DA5_DA6	BB00
1en9_A_DG6_DC7	BA05	1ene_A_DG6_DC7	BA05	1fq2_A_DA6_DT7	BA05	463d_A_DA6_DT7	BB01
1en9_A_DC7_DT8	AB01	1ene_A_DC7_DT8	AB01	1fq2_A_DT7_DT8	BB01	463d_A_DT7_DT8	BA05
1en9_A_DT8_DG9	BB07	1ene_A_DT8_DG9	BB07	1fq2_A_DT8_DC9	BB01	463d_A_DT8_DC9	BB01
1en9_A_DG9_DG10	BB00	1ene_A_DG9_DG10	BB00	1fq2_A_DC9_DG10	BB00	463d_A_DC9_DG10	NANT
				1fq2_A_DG10_DC11	BB07	463d_A_DG10_DC11	BB07
				1fq2_A_DC11_DG12	NANT	463d_A_DC11_DG12	NANT
				1fq2_B_DC1_DG2	BA05		
1en9_A_DC1_DC2	BB00	1ene_A_DC1_DC2	BB00	1fq2_B_DG2_DC3	BA05	463d_B_DG14_DC15	BB00
1en9_A_DC2_DA3	BB07	1ene_A_DC2_DA3	BB07	1fq2_B_DC3_DG4	AB01	463d_B_DC15_DG16	BB00
1en9_A_DA3_DG4	BB00	1ene_A_DA3_DG4	BB00	1fq2_B_DG4_DA5	BB00	463d_B_DG16_DA17	BB00
1en9_A_DG4_DC5	BB07	1ene_A_DG4_DC5	BB07	1fq2_B_DA5_DA6	BA05	463d_B_DA17_DA18	BA05
1en9_A_DC5_DG6	BB00	1ene_A_DC5_DG6	BB00	1fq2_B_DA6_DT7	BB01	463d_B_DA18_DT19	BB01
1en9_A_DG6_DC7	BA05	1ene_A_DG6_DC7	BA05	1fq2_B_DT7_DT8	BB01	463d_B_DT19_DT20	BB01
1en9_A_DC7_DT8	AB01	1ene_A_DC7_DT8	AB01	1fq2_B_DT8_DC9	BA05	463d_B_DT20_DC21	BA05
1en9_A_DT8_DG9	BB07	1ene_A_DT8_DG9	BB07	1fq2_B_DC9_DG10	AB01	463d_B_DC21_DG22	AB01
1en9_A_DG9_DG10	BB00	1ene_A_DG9_DG10	BB00	1fq2_B_DG10_DC11	BA08	463d_B_DG22_DC23	BB07
				1fq2_B_DC11_DG12	AA00	463d_B_DC23_DG24	NANT
Tsunoda, M., Acta D 57: 345-348, '01		Tsunoda, M., Acta D 57: 345-348, '01		Tsunoda, M., Acta D 57: 345-348, '01		Tsunoda, M., Acta D 57: 345-348, '01	
step	Ddnc	step	Ddnc	step	Ddnc	step	Ddnc
1g75_A_DC1_DG2	BB00	1g8n_A_DC1_DG2	BB00	1g8u_A_DC1_DG2	BA10	1g8v_A_DC1_DG2	BB04
1g75_A_DG2_DC3	BA01	1g8n_A_DG2_DC3	BA01	1g8u_A_DG2_DC3	AA00	1g8v_A_DG2_DC3	BA05
1g75_A_DC3_DG4	AB01	1g8n_A_DC3_DG4	AB01	1g8u_A_DC3_DG4	AB01	1g8v_A_DC3_DG4	AB01
1g75_A_DG4_DA5	BB00	1g8n_A_DG4_DA5	BB01	1g8u_A_DG4_DA5	BB01	1g8v_A_DG4_DA5	BB01
1g75_A_DA5_DA6	BB01	1g8n_A_DA5_DA6	BB01	1g8u_A_DA5_DA6	BB01	1g8v_A_DA5_DA6	BB00
1g75_A_DA6_DT7	BA05	1g8n_A_DA6_DT7	BA05	1g8u_A_DA6_DT7	BB01	1g8v_A_DA6_DT7	BB01
1g75_A_DC9_DG10	BB00	1g8n_A_DC9_DG10	BB00	1g8u_A_DC9_DG10	BB00	1g8v_A_DC9_DG10	BB00
1g75_A_DG10_DC11	BB07	1g8n_A_DG10_DC11	BB07	1g8u_A_DG10_DC11	BB07	1g8v_A_DG10_DC11	BB07
1g75_A_DC11_DG12	BA05	1g8n_A_DC11_DG12	BA05	1g8u_A_DC11_DG12	BA05	1g8v_A_DC11_DG12	BA05
1g75_B_DC13_DG14	BB00	1g8n_B_DC13_DG14	BA05	1g8u_B_DC13_DG14	BB04	1g8v_B_DC13_DG14	BB07
1g75_B_DG14_DC15	BA05	1g8n_B_DG14_DC15	BA05	1g8u_B_DG14_DC15	BA05	1g8v_B_DG14_DC15	BA01
1g75_B_DC15_DG16	AB01	1g8n_B_DC15_DG16	AB01	1g8u_B_DC15_DG16	AB01	1g8v_B_DC15_DG16	AB01
1g75_B_DG16_DA17	BB00	1g8n_B_DG16_DA17	BB00	1g8u_B_DG16_DA17	BB00	1g8v_B_DG16_DA17	BB00
1g75_B_DA17_DA18	BB01	1g8n_B_DA17_DA18	BB01	1g8u_B_DA17_DA18	BB01	1g8v_B_DA17_DA18	BB01
1g75_B_DA18_DT19	BA05	1g8n_B_DA18_DT19	BA05	1g8u_B_DA18_DT19	BB01	1g8v_B_DA18_DT19	BB01
1g75_B_DC21_DG22	BB00	1g8n_B_DC21_DG22	BB00	1g8u_B_DC21_DG22	BB00	1g8v_B_DC21_DG22	BB00
1g75_B_DG22_DC23	BA08	1g8n_B_DG22_DC23	BA08	1g8u_B_DG22_DC23	BA17	1g8v_B_DG22_DC23	BA08
1g75_B_DC23_DG24	AA00	1g8n_B_DC23_DG24	AA00	1g8u_B_DC23_DG24	BA05	1g8v_B_DC23_DG24	AA00
Wilds, C.J., JACS 124: 13716-13721, '02		Moulaei, T., BCH 44: 7458-7468, '05		Fratini, A.V., JBC 257: 14686-14707, '82		Chiu, T.K., JMB 292: 589-608, '99	
step	Ddnc	step	Ddnc	step	Ddnc	step	Ddnc
1n1o_A_DC1_DG2	BB00	1z5t_A_DC1_DG2	BB04	3bna_A_DC1_DG2	NANT	455d_A_DC1_DG2	BB00
1n1o_A_DG2_DC3	BA01	1z5t_A_DG2_DC3	BA01	3bna_A_DG2_DC3	BA05	455d_A_DG2_DC3	BA01
1n1o_A_DC3_DG4	AB01	1z5t_A_DC3_DG4	AB01	3bna_A_DC3_DG4	BB16	455d_A_DC3_DG4	AB01
1n1o_A_DG4_DA5	BB00	1z5t_A_DG4_DA5	BB00	3bna_A_DG4_DA5	BB00	455d_A_DG4_DA5	BB00
1n1o_A_DA5_DA6	BB00	1z5t_A_DA5_DA6	BB00	3bna_A_DA5_DA6	BB01	455d_A_DA5_DA6	BB00
				3bna_A_DA6_DT7	BA05	455d_A_DA6_DT7	BB01
				3bna_A_DT7_DT8	AA02	455d_A_DT7_DT8	BB01
1n1o_A_DT8_DC9	BB00			3bna_A_DT8_CBR9	BA05	455d_A_DT8_DC9	BB00
1n1o_A_DC9_DG10	BB00	1z5t_A_DC9_DG10	BB00	3bna_A_CBR9_DG10	AB01	455d_A_DC9_DG10	BB00
1n1o_A_DG10_DC11	BB07	1z5t_A_DG10_DC11	BB07	3bna_A_DG10_DC11	BB07	455d_A_DG10_DC11	BB07
1n1o_A_DC11_DG12	BB00	1z5t_A_DC11_DG12	BA05	3bna_A_DC11_DG12	BA05	455d_A_DC11_DG12	BA05
1n1o_B_DC113_DG114	BB01	1z5t_B_DC13_DG14	BB07	3bna_B_DC13_DG14	BB04	455d_B_DC13_DG14	BB07
1n1o_B_DG114_DC115	BA05	1z5t_B_DG14_DC15	BA01	3bna_B_DG14_DC15	BA01	455d_B_DG14_DC15	BB01
1n1o_B_DC115_DG116	AB01	1z5t_B_DC15_DG16	AB01	3bna_B_DC15_DG16	AB01	455d_B_DC15_DG16	NANT
1n1o_B_DG116_DA117	BB00	1z5t_B_DG16_DA17	BB00	3bna_B_DG16_DA17	BB00	455d_B_DG16_DA17	BB00
1n1o_B_DA117_DA118	BB00	1z5t_B_DA17_DA18	BB01	3bna_B_DA17_DA18	BA05	455d_B_DA17_DA18	BB01
				3bna_B_DA18_DT19	AA00	455d_B_DA18_DT19	BB01
				3bna_B_DT19_DT20	AA02	455d_B_DT19_DT20	BB01
1n1o_B_DT120_DC121	BA05			3bna_B_DT20_CBR21	BA05	455d_B_DT20_DC21	BB00
1n1o_B_DC121_DG122	AB01	1z5t_B_DC21_DG22	BB00	3bna_B_CBR21_DG22	AB01	455d_B_DC21_DG22	BB00
1n1o_B_DG122_DC123	BA08	1z5t_B_DG22_DC23	BA17	3bna_B_DG22_DC23	BB07	455d_B_DG22_DC23	NANT
1n1o_B_DC123_DG124	AA00	1z5t_B_DC23_DG24	AA02	3bna_B_DC23_DG24	BA01	455d_B_DC23_DG24	BA05

Kowal, E.A., NAR 41: 7566-7576, '13		Szulik, M.W., BCH 54: 1294-1305, '15		Jiang, S., TBP		Theruvathu, J.A., BCH 52: 8590, '13	
step	Ddnc	step	Ddnc	step	Ddnc	step	Ddnc
4hqi_A_DG2_DC3	BA05	4i9v_A_DC1_DG2	BB00	4kw0_A_DC1_DG2	BB00	4mgw_A_DC1_DG2	BB00
		4i9v_A_DG2_DC3	BA01	4kw0_A_DG2_DC3	BA01	4mgw_A_DG4_DA5	BB00
		4i9v_A_DC3_DG4	AB01	4kw0_A_DC3_DG4	AB01	4mgw_A_DA5_DA6	BB00
		4i9v_A_DG4_DA5	BB07	4kw0_A_DG4_DA5	BB00	4mgw_A_DA6_DT7	BB01
4hqi_A_DA5_DA6	BB01	4i9v_A_DA5_DA6	BB01	4kw0_A_DA5_DA6	BB01	4mgw_A_DT7_DT8	BB01
4hqi_A_DA6_DT7	BA05	4i9v_A_DA6_DT7	BA05	4kw0_A_DA6_DT7	BA05	4mgw_A_DT8_DC9	BB00
4hqi_A_DT7_DT8	BB01	4i9v_A_DT7_DT8	BB01	4kw0_A_DT7_DT8	BB01	4mgw_A_DC9_DG10	BB00
				4kw0_A_DT8_DC9	BB00		
4hqi_A_DG10_DC11	BB04	4i9v_A_DG10_DC11	BB07			4mgw_A_DG10_DC11	BB07
4hqi_A_DC11_DG12	NANT	4i9v_A_DC11_DG12	BB00	4kw0_A_DC11_DG12	BB00	4mgw_A_DC11_DG12	BA05
		4i9v_B_DC13_DG14	BA05	4kw0_B_DC13_DG14	BB01	4mgw_B_DC13_DG14	BB07
4hqi_B_DG14_DC15	BB01	4i9v_B_DG14_DC15	BA05	4kw0_B_DG14_DC15	BA05		
		4i9v_B_DC15_DG16	AB01	4kw0_B_DC15_DG16	AB01		
		4i9v_B_DG16_DA17	BB00	4kw0_B_DG16_DA17	BB00	4mgw_B_DG16_DA17	BB00
4hqi_B_DA17_DA18	BB07	4i9v_B_DA17_DA18	BB01	4kw0_B_DA17_DA18	BB01	4mgw_B_DA17_DA18	BB00
4hqi_B_DA18_DT19	BA05	4i9v_B_DA18_DT19	BA05	4kw0_B_DA18_DT19	BB01	4mgw_B_DA18_DT19	BB01
4hqi_B_DT19_DT20	AB01	4i9v_B_DT19_DT20	BB01	4kw0_B_DT19_DT20	BB01	4mgw_B_DT19_DT20	BB01
				4kw0_B_DT20_DC21	BB01	4mgw_B_DT20_DC21	BB00
						4mgw_B_DC21_DG22	BB00
4hqi_B_DG22_DC23	BB04	4i9v_B_DG22_DC23	BA08			4mgw_B_DG22_DC23	BA08
4hqi_B_DC23_DG24	NANT	4i9v_B_DC23_DG24	AA00	4kw0_B_DC23_DG24	BA01	4mgw_B_DC23_DG24	AA00

X-ray structures with drugs bound in the minor groove

Wood, A.A., NAR 23: 3678-3684, '95		Quintana, J.R., BCH 30: 10294, '91		Quintana, J.R., BCH 30: 10294, '91		Quintana, J.R., BCH 30: 10294, '91	
step	Ddnc	step	Ddnc	step	Ddnc	step	Ddnc
109d_A_DC1_DG2	BB00	1d43_A_DC1_DG2	BB00	1d44_A_DC1_DG2	BB00	1d45_A_DC1_DG2	BB00
109d_A_DG2_DC3	BB00	1d43_A_DG2_DC3	BA01	1d44_A_DG2_DC3	BA01	1d45_A_DG2_DC3	BA01
109d_A_DC3_DG4	BB04	1d43_A_DC3_DG4	AB01	1d44_A_DC3_DG4	AB01	1d45_A_DC3_DG4	AB01
109d_A_DG4_DA5	BB04	1d43_A_DG4_DA5	BB04	1d44_A_DG4_DA5	BB00	1d45_A_DG4_DA5	BB00
109d_A_DA5_DA6	BB00	1d43_A_DA5_DA6	BB00	1d44_A_DA5_DA6	BB01	1d45_A_DA5_DA6	BB00
109d_A_DA6_DT7	BB01	1d43_A_DA6_DT7	BA05	1d44_A_DA6_DT7	BA05	1d45_A_DA6_DT7	BA05
109d_A_DT7_DT8	BB00	1d43_A_DT7_DT8	NANT	1d44_A_DT7_DT8	NANT	1d45_A_DT7_DT8	NANT
109d_A_DT8_DC9	BB00	1d43_A_DT8_DC9	NANT	1d44_A_DT8_DC9	NANT	1d45_A_DT8_DC9	NANT
109d_A_DC9_DG10	BB00	1d43_A_DC9_DG10	BB04	1d44_A_DC9_DG10	BB04	1d45_A_DC9_DG10	NANT
109d_A_DG10_DC11	BB07	1d43_A_DG10_DC11	BB07	1d44_A_DG10_DC11	BB07	1d45_A_DG10_DC11	NANT
109d_A_DC11_DG12	BB00	1d43_A_DC11_DG12	NANT	1d44_A_DC11_DG12	NANT	1d45_A_DC11_DG12	BB00
109d_B_DC13_DG14	BB07	1d43_B_DC13_DG14	NANT	1d44_B_DC13_DG14	NANT	1d45_B_DC13_DG14	BB07
109d_B_DG14_DC15	BB00	1d43_B_DG14_DC15	BA05	1d44_B_DG14_DC15	AA02	1d45_B_DG14_DC15	BA05
109d_B_DC15_DG16	BB04	1d43_B_DC15_DG16	AB01	1d44_B_DC15_DG16	AB01	1d45_B_DC15_DG16	AB01
109d_B_DG16_DA17	BB00	1d43_B_DG16_DA17	NANT	1d44_B_DG16_DA17	NANT	1d45_B_DG16_DA17	BB04
109d_B_DA17_DA18	BB00	1d43_B_DA17_DA18	BB04	1d44_B_DA17_DA18	BB04	1d45_B_DA17_DA18	BB00
109d_B_DA18_DT19	BB00	1d43_B_DA18_DT19	BA05	1d44_B_DA18_DT19	BB00	1d45_B_DA18_DT19	BA01
109d_B_DT19_DT20	BB00	1d43_B_DT19_DT20	BB01	1d44_B_DT19_DT20	BB01	1d45_B_DT19_DT20	AA02
109d_B_DT20_DC21	BB01	1d43_B_DT20_DC21	BB01	1d44_B_DT20_DC21	BB01	1d45_B_DT20_DC21	NANT
109d_B_DC21_DG22	BB00	1d43_B_DC21_DG22	BB00	1d44_B_DC21_DG22	BB00	1d45_B_DC21_DG22	NANT
109d_B_DG22_DC23	BB07	1d43_B_DG22_DC23	NANT	1d44_B_DG22_DC23	NANT	1d45_B_DG22_DC23	BA17
109d_B_DC23_DG24	BB02	1d43_B_DC23_DG24	NANT	1d44_B_DC23_DG24	AA03	1d45_B_DC23_DG24	AA04
Quintana, J.R., BCH 30: 10294, '91		Coll, M., BCH 28: 310-320, '89		Campbell, N.H., BMCL 16: 15-19, '06		Neidle, S., TBP	
step	Ddnc	step	Ddnc	step	Ddnc	step	Ddnc
1d46_A_DC1_DG2	NANT	1dne_A_DC1_DG2	NANT	2b3e_A_DC1_DG2	BB00	2gvr_A_DC1_DG2	BB00
1d46_A_DG2_DC3	NANT	1dne_A_DG2_DC3	NANT	2b3e_A_DG2_DC3	BA01	2gvr_A_DG2_DC3	BA01
1d46_A_DC3_DG4	NANT	1dne_A_DC3_DG4	NANT	2b3e_A_DC3_DG4	AB01	2gvr_A_DC3_DG4	AB01
1d46_A_DG4_DA5	BB07	1dne_A_DG4_DA5	NANT	2b3e_A_DG4_DA5	BB00	2gvr_A_DG4_DA5	BB00
1d46_A_DA5_DA6	BA05	1dne_A_DA5_DA6	NANT	2b3e_A_DA5_DA6	BB01	2gvr_A_DA5_DA6	BB01
1d46_A_DA6_DT7	NANT	1dne_A_DT6_DA7	NANT	2b3e_A_DA6_DT7	BA05	2gvr_A_DA6_DT7	BB01
1d46_A_DT7_DT8	NANT	1dne_A_DA7_DT8	NANT	2b3e_A_DT7_DT8	BB01	2gvr_A_DT7_DT8	BB01
1d46_A_DT8_DC9	NANT	1dne_A_DT8_DC9	NANT	2b3e_A_DT8_DC9	BB01	2gvr_A_DT8_DC9	BB01
1d46_A_DC9_DG10	BB04	1dne_A_DC9_DG10	NANT	2b3e_A_DC9_DG10	BB00	2gvr_A_DC9_DG10	NANT
1d46_A_DG10_DC11	BB07	1dne_A_DG10_DC11	NANT	2b3e_A_DG10_DC11	BB07	2gvr_A_DG10_DC11	BB07
1d46_A_DC11_DG12	NANT	1dne_A_DC11_DG12	NANT	2b3e_A_DC11_DG12	BA05	2gvr_A_DC11_DG12	BA05
1d46_B_DC13_DG14	BA17	1dne_B_DC13_DG14	BB13	2b3e_B_DC13_DG14	BB07	2gvr_B_DC13_DG14	BB07
1d46_B_DG14_DC15	NANT	1dne_B_DG14_DC15	BA05	2b3e_B_DG14_DC15	BA05	2gvr_B_DG14_DC15	BA05
1d46_B_DC15_DG16	AB01	1dne_B_DC15_DG16	NANT	2b3e_B_DC15_DG16	AB01	2gvr_B_DC15_DG16	AB01
1d46_B_DG16_DA17	NANT	1dne_B_DG16_DA17	NANT	2b3e_B_DG16_DA17	BB00	2gvr_B_DG16_DA17	BB00
1d46_B_DA17_DA18	BB00	1dne_B_DA17_DT18	NANT	2b3e_B_DA17_DA18	BB00	2gvr_B_DA17_DA18	BB01
1d46_B_DA18_DT19	NANT	1dne_B_DT18_DA19	NANT	2b3e_B_DA18_DT19	BB01	2gvr_B_DA18_DT19	BB01
1d46_B_DT19_DT20	NANT	1dne_B_DA19_DT20	NANT	2b3e_B_DT19_DT20	BB01	2gvr_B_DT19_DT20	BB01
1d46_B_DT20_DC21	BB01	1dne_B_DT20_DC21	BB00	2b3e_B_DT20_DC21	BA05	2gvr_B_DT20_DC21	BB01
1d46_B_DC21_DG22	BB04	1dne_B_DC21_DG22	NANT	2b3e_B_DC21_DG22	BB00	2gvr_B_DC21_DG22	BB00
1d46_B_DG22_DC23	NANT	1dne_B_DG22_DC23	NANT	2b3e_B_DG22_DC23	BB07	2gvr_B_DG22_DC23	BB07
1d46_B_DC23_DG24	BA01	1dne_B_DC23_DG24	NANT	2b3e_B_DC23_DG24	BA05	2gvr_B_DC23_DG24	BA05

Selected NMR structures

Tjandra, N., JACS 122: 6190-6200, '00		Kuszewski, J., JACS 123: 3903-3918, '01		Wu, Z., JBN 26: 297-315, '03		Denisov, A.Y., JBSD 16: 547-568, '98	
step	Ddnc	step	Ddnc	step	Ddnc	step	Ddnc
1duf_A_DC1_DG2	BB00	1gip_A_DC1_DG2	BB00	1naj_A_DC1_DG2	BB00	2dau_A_DC1_DG2	BB00
1duf_A_DG2_DC3	BB01	1gip_A_DG2_DC3	BB01	1naj_A_DG2_DC3	BB01	2dau_A_DG2_DC3	BB00
1duf_A_DC3_DG4	BB00	1gip_A_DC3_DG4	BB00	1naj_A_DC3_DG4	BB00	2dau_A_DC3_DG4	BB00
1duf_A_DG4_DA5	BB00	1gip_A_DG4_DA5	BB00	1naj_A_DG4_DA5	BB00	2dau_A_DG4_DA5	BB00
1duf_A_DA5_DA6	BB00	1gip_A_DA5_DA6	BB00	1naj_A_DA5_DA6	BB01	2dau_A_DA5_DA6	BB00
1duf_A_DA6_DT7	BB01	1gip_A_DA6_DT7	BB01	1naj_A_DA6_DT7	BA05	2dau_A_DA6_DT7	BA05
1duf_A_DT7_DT8	BB01	1gip_A_DT7_DT8	BB01	1naj_A_DT7_DT8	BB01	2dau_A_DT7_DT8	BA05
1duf_A_DT8_DC9	BA05	1gip_A_DT8_DC9	BA05	1naj_A_DT8_DC9	BB00	2dau_A_DT8_DC9	BB01
1duf_A_DC9_DG10	BB01	1gip_A_DC9_DG10	BB00	1naj_A_DC9_DG10	BB00	2dau_A_DC9_DG10	BB01
1duf_A_DG10_DC11	BA05	1gip_A_DG10_DC11	BB01	1naj_A_DG10_DC11	BB01	2dau_A_DG10_DC11	BB00
1duf_A_DC11_DG12	BB01	1gip_A_DC11_DG12	BB01	1naj_A_DC11_DG12	BB01	2dau_A_DC11_DG12	BB01
1duf_B_DC13_DG14	BB00	1gip_B_DC13_DG14	BB00	1naj_B_DC13_DG14	BB00	2dau_B_DC13_DG14	BB00
1duf_B_DG14_DC15	BB01	1gip_B_DG14_DC15	BB01	1naj_B_DG14_DC15	BB01	2dau_B_DG14_DC15	BB00
1duf_B_DC15_DG16	BB00	1gip_B_DC15_DG16	BB00	1naj_B_DC15_DG16	BB00	2dau_B_DC15_DG16	BB00
1duf_B_DG16_DA17	BB00	1gip_B_DG16_DA17	BB00	1naj_B_DG16_DA17	BB00	2dau_B_DG16_DA17	BB00
1duf_B_DA17_DA18	BB00	1gip_B_DA17_DA18	BB00	1naj_B_DA17_DA18	BB01	2dau_B_DA17_DA18	BB00
1duf_B_DA18_DT19	BB01	1gip_B_DA18_DT19	BB01	1naj_B_DA18_DT19	BA05	2dau_B_DA18_DT19	BA05
1duf_B_DT19_DT20	BB01	1gip_B_DT19_DT20	BB01	1naj_B_DT19_DT20	BB01	2dau_B_DT19_DT20	BA05
1duf_B_DT20_DC21	BA05	1gip_B_DT20_DC21	BA05	1naj_B_DT20_DC21	BB00	2dau_B_DT20_DC21	BB01
1duf_B_DC21_DG22	BB01	1gip_B_DC21_DG22	BB00	1naj_B_DC21_DG22	BB00	2dau_B_DC21_DG22	BB01
1duf_B_DG22_DC23	BA05	1gip_B_DG22_DC23	BB01	1naj_B_DG22_DC23	BB01	2dau_B_DG22_DC23	BB00
1duf_B_DC23_DG24	BB01	1gip_B_DC23_DG24	BB01	1naj_B_DC23_DG24	BB01	2dau_B_DC23_DG24	BB01

Fiber models from AmberTools

AmberTools "Arnot"		AmberTools "Langridge"		AmberTools "Average B"	
step	Ddnc	step	Ddnc	step	Ddnc
arnot-fd-bdna_X_DC1_DG2	BB00	lang-bdna_X_DC1_DG2	BB00	aver-bdna_A_DC1_DG2	NANT
arnot-fd-bdna_X_DG2_DC3	BB00	lang-bdna_X_DG2_DC3	BB00	aver-bdna_A_DG2_DC3	NANT
arnot-fd-bdna_X_DC3_DG4	BB00	lang-bdna_X_DC3_DG4	BB00	aver-bdna_A_DC3_DG4	NANT
arnot-fd-bdna_X_DG4_DA5	BB00	lang-bdna_X_DG4_DA5	BB00	aver-bdna_A_DG4_DA5	NANT
arnot-fd-bdna_X_DA5_DA6	BB00	lang-bdna_X_DA5_DA6	BB00	aver-bdna_A_DA5_DA6	BB15
arnot-fd-bdna_X_DA6_DT7	BB00	lang-bdna_X_DA6_DT7	BB00	aver-bdna_A_DA6_DT7	BB15
arnot-fd-bdna_X_DT7_DT8	BB00	lang-bdna_X_DT7_DT8	BB00	aver-bdna_A_DT7_DT8	BB15
arnot-fd-bdna_X_DT8_DC9	BB00	lang-bdna_X_DT8_DC9	BB00	aver-bdna_A_DT8_DC9	NANT
arnot-fd-bdna_X_DC9_DG10	BB00	lang-bdna_X_DC9_DG10	BB00	aver-bdna_A_DC9_DG10	NANT
arnot-fd-bdna_X_DG10_DC11	BB00	lang-bdna_X_DG10_DC11	BB00	aver-bdna_A_DG10_DC11	NANT
arnot-fd-bdna_X_DC11_DG12	BB00	lang-bdna_X_DC11_DG12	BB00	aver-bdna_A_DC11_DG12	NANT
arnot-fd-bdna_X_DC13_DG14	BB00	lang-bdna_X_DC13_DG14	BB00	aver-bdna_B_DC1_DG2	NANT
arnot-fd-bdna_X_DG14_DC15	BB00	lang-bdna_X_DG14_DC15	BB00	aver-bdna_B_DG2_DC3	NANT
arnot-fd-bdna_X_DC15_DG16	BB00	lang-bdna_X_DC15_DG16	BB00	aver-bdna_B_DC3_DG4	NANT
arnot-fd-bdna_X_DG16_DA17	BB00	lang-bdna_X_DG16_DA17	BB00	aver-bdna_B_DG4_DA5	NANT
arnot-fd-bdna_X_DA17_DA18	BB00	lang-bdna_X_DA17_DA18	BB00	aver-bdna_B_DA5_DA6	BB15
arnot-fd-bdna_X_DA18_DT19	BB00	lang-bdna_X_DA18_DT19	BB00	aver-bdna_B_DA6_DT7	BB15
arnot-fd-bdna_X_DT19_DT20	BB00	lang-bdna_X_DT19_DT20	BB00	aver-bdna_B_DT7_DT8	NANT
arnot-fd-bdna_X_DT20_DC21	BB00	lang-bdna_X_DT20_DC21	BB00	aver-bdna_B_DT8_DC9	NANT
arnot-fd-bdna_X_DC21_DG22	BB00	lang-bdna_X_DC21_DG22	BB00	aver-bdna_B_DC9_DG10	NANT
arnot-fd-bdna_X_DG22_DC23	BB00	lang-bdna_X_DG22_DC23	BB00	aver-bdna_B_DG10_DC11	NANT
arnot-fd-bdna_X_DC23_DG24	BB00	lang-bdna_X_DC23_DG24	BB00	aver-bdna_B_DC11_DG12	NANT

Journal abbreviations

Proc. Natl. Acad. Sci. USA	PNAS
J. Biomol. Struct. Dyn.	JBSD
Nucleic Acids Res.	NAR
J. Biomol. NMR	JBN
J. Am. Chem. Soc.	JACS
Biochemistry	BCH
Acta Cryst.	Acta
J. Mol. Biol.	JMB
Chem. Commun. (Camb.)	CHC
J. Biol. Chem.	JBC
Bioorg. Med. Chem. Lett.	BMCL

Statistics for 1bna + 2bna + 7bna + 9bna

Ddnc	count	%
AA00	1	1
AB01	8	9
BA04	1	1
BA06	17	19
BA17	2	2
BB00	22	25
BB01	20	23
BB04	7	8
BB07	6	7
BB15	1	1
NANT	3	3

Table S3. The assignment of the *NtC* classes to selected crystal, NMR, and MD structures. c) *NtC* classes assigned to dodecamres related to the Dickerson-Drew dodecamer as were deposited to the PDB and after their coordinates were modified by the REDO_PDB protocol. Discussion in the main text was based on *NtC* assignment to 34 *D-Dd* structures, Table shows results for selected six structures.

Step	original confa	original NtC	redo confal	redo NtC
1bna_A_DC_1_DG_2	60	BB04	69	BB04
1bna_A_DG_2_DC_3	91	BA05	84	BA05
1bna_A_DC_3_DG_4	72	AB01	82	AB01
1bna_A_DG_4_DA_5	67	BB04	90	BB00
1bna_A_DA_5_DA_6	75	BB01	94	BB01
1bna_A_DA_6_DT_7	82	BA05	88	BA05
1bna_A_DT_7_DT_8	71	BA05	78	BB01
1bna_A_DT_8_DC_9	69	BB01	81	BB00
1bna_A_DC_9_DG_10	76	BB00	75	BB00
1bna_A_DG_10_DC_11	74	BB07	89	BB07
1bna_A_DC_11_DG_12	73	BA05	69	BB00
1bna_B_DC_13_DG_14	62	BB00	71	BB04
1bna_B_DG_14_DC_15	85	BA05	80	BA05
1bna_B_DC_15_DG_16	73	AB01	80	AB01
1bna_B_DG_16_DA_17	86	BB00	79	BB00
1bna_B_DA_17_DA_18	89	BB00	77	BB00
1bna_B_DA_18_DT_19	87	BA05	87	BA05
1bna_B_DT_19_DT_20	78	BB01	71	BB01
1bna_B_DT_20_DC_21	83	BB01	87	BB01
1bna_B_DC_21_DG_22	70	BB00	71	BB00
1bna_B_DG_22_DC_23	88	BA17	78	BA17
1bna_B_DC_23_DG_24	75	BA05	55	NANT
1d29_A_DC_1_DG_2	66	BA17	78	BB04
1d29_A_DG_2_DT_3	0	NANT	72	BA01
1d29_A_DT_3_DG_4	0	NANT	73	AB01
1d29_A_DG_4_DA_5	0	NANT	80	BB04
1d29_A_DA_5_DA_6	74	AB01	86	BB00
1d29_A_DA_6_DT_7	70	BA01	83	BB01
1d29_A_DT_7_DT_8	0	NANT	68	BB01
1d29_A_DT_8_DC_9	44	BB01	91	BB00
1d29_A_DC_9_DA_10	54	BB00	72	BB00
1d29_A_DA_10_DC_11	43	BB07	71	BB07
1d29_A_DC_11_DG_12	86	BB00	75	BA01
1d29_B_DC_13_DG_14	0	NANT	76	BB04
1d29_B_DG_14_DT_15	37	BA05	69	BA01
1d29_B_DT_15_DG_16	0	NANT	61	AB01
1d29_B_DG_16_DA_17	64	BB04	80	BB04
1d29_B_DA_17_DA_18	71	BB00	93	BB01
1d29_B_DA_18_DT_19	57	BB04	83	BB01
1d29_B_DT_19_DT_20	82	BB01	79	BB00
1d29_B_DT_20_DC_21	70	BB01	88	BB00
1d29_B_DC_21_DA_22	59	BB00	65	BB00
1d29_B_DA_22_DC_23	63	BB07	68	BB07
1d29_B_DC_23_DG_24	62	BA01	71	NANT

4i9v_A_DC_1_DG_2	89	BB00	90	BB00
4i9v_A_DG_2_DC_3	39	BA01	43	BA01
4i9v_A_DC_3_DG_4	73	AB01	76	AB01
4i9v_A_DG_4_DA_5	95	BB07	93	BB07
4i9v_A_DA_5_DA_6	79	BB01	77	BB01
4i9v_A_DA_6_DT_7	91	BA05	91	BA05
4i9v_A_DT_7_DT_8	82	BB01	81	BB01
4i9v_A_DG_10_DC_11	83	BB07	85	BB07
4i9v_A_DC_11_DG_12	67	BB00	65	BB00
4i9v_B_DC_13_DG_14	95	BA05	94	BA05
4i9v_B_DG_14_DC_15	68	BA05	68	BA05
4i9v_B_DC_15_DG_16	92	AB01	92	AB01
4i9v_B_DG_16_DA_17	77	BB00	76	BB00
4i9v_B_DA_17_DA_18	92	BB01	90	BB01
4i9v_B_DA_18_DT_19	94	BA05	92	BA05
4i9v_B_DT_19_DT_20	68	BB01	69	BB01
4i9v_B_DG_22_DC_23	97	BA08	97	BA08
4i9v_B_DC_23_DG_24	85	AA00	87	AA00
5bna_A_DC_1_DG_2	62	BB00	0	NANT
5bna_A_DG_2_DC_3	76	BA05	65	BA05
5bna_A_DC_3_DG_4	68	AB01	78	AB01
5bna_A_DG_4_DA_5	0	NANT	71	BB00
5bna_A_DA_5_DA_6	64	BB00	93	BB00
5bna_A_DA_6_DT_7	52	BB01	62	BA05
5bna_A_DT_7_DT_8	78	BA05	80	BA05
5bna_A_DT_8_DC_9	70	BA05	82	BB01
5bna_A_DC_9_DG_10	71	AB01	47	BB00
5bna_A_DG_10_DC_11	80	BB07	73	BB07
5bna_A_DC_11_DG_12	53	BB00	0	NANT
5bna_B_DC_13_DG_14	65	BB07	63	BB04
5bna_B_DG_14_DC_15	78	BA05	68	BA05
5bna_B_DC_15_DG_16	65	AB01	0	NANT
5bna_B_DG_16_DA_17	58	BB00	56	BB00
5bna_B_DA_17_DA_18	62	BB01	64	BB01
5bna_B_DA_18_DT_19	0	NANT	68	NANT
5bna_B_DT_19_DT_20	0	NANT	65	NANT
5bna_B_DT_20_DC_21	65	BB01	73	BB01
5bna_B_DC_21_DG_22	79	BB00	53	BB00
5bna_B_DG_22_DC_23	45	BA08	60	BA17
5bna_B_DC_23_DG_24	56	AA00	68	AA03

DNA models derived from fiber diffraction.

Fit of selected fiber-based models of B-DNA duplex to *NtC* classes

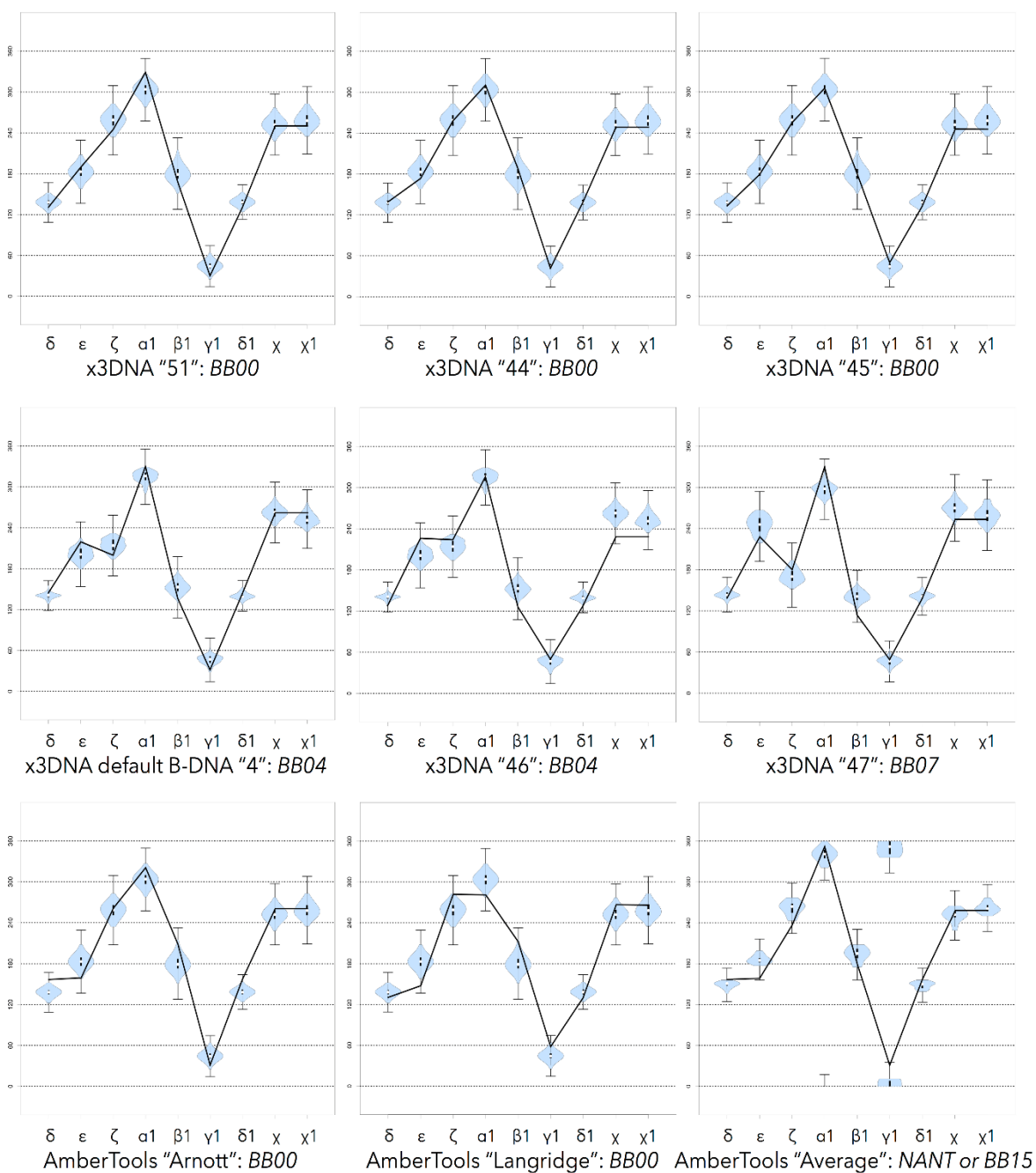


Figure S4. Assignment of *NtC* classes for B-DNA duplex models based on the fiber data. Shown are assignments of six models from x3DNA (Lu & Olson, 2008) and three models from Nucleic Acid Builder of AmberTools14 (Salomon-Ferrer *et al.*, 2013).

Table S4. The assignment of *NtC* classes to the fiber model structures, which are available in x3DNA.

id#	x3dna description of the fiber model			Assigned dinucleotide conformers								Comments
	Twist [°]	Rise [Å]	Structure description	Ddnc	%	Ddnc	%	Ddnc	%	Ddnc	%	
1	33	2.55	A-DNA (calf thymus; generic sequence: A, C, G and T)	AA00	100							default A form fiber in x3dna
2	66	5.10	A-DNA poly d(ABr5U) : poly d(ABr5U)	AA00	100							
3	0	28.03	A-DNA (calf thymus) poly d(A1T2C3G4G5A6A7T8G9G10T11) poly d(A1C2C3A4T5T6C7C8G9A10T11)	AA00	100							
4	36	3.38	B-DNA (calf thymus; generic sequence: A, C, G and T)	BB04	100							default B form fiber in x3dna
5	72	6.72	B-DNA poly d(CG) : poly d(CG)	BB07	50	BB04	50					
6	180	16.86	B-DNA (calf thymus) poly d(C1C2C3C4C5) : poly d(G6G7G8G9G10)	BB00	39	BB07	33	BB04	28			
7	39	3.31	C-DNA (calf thymus; generic sequence: A, C, G and T)	BB00	100							chain A (BB04, BB00), chain B (BB07, NANT), NANT similar to BB07 (zeta too high)
8	40	3.31	C-DNA poly d(GGT) : poly d(ACC)	BB07	100							
9	120	9.94	C-DNA poly d(G1G2T3) : poly d(A4C5C6)	BB04	30	BB07	30	BB00	20	NANT	20	
10	80	6.47	C-DNA poly d(AG) : poly d(CT)	BB00	67	NANT	33					NANT similar to BB04 (gamma too high)
11	80	6.47	C-DNA poly d(A1G2) : poly d(C3T4)	BB04	33	BB00	33	BB07	17	NANT	17	NANT similar to BB16 (zeta too low, gamma and chi too high)
12	45	3.01	D-DNA poly d(AAT) : poly d(ATT)	BB04	100							NANT similar to BB07 (alpha too low, zeta and gamma too high)
13	90	6.13	D-DNA poly d(CI) : poly d(CI)	BB04	100							
14	-90	18.50	D-DNA poly d(A1T2A3T4A5T6) : poly d(A1T2A3T4A5T6)	BB00	55	NANT	36	BB04	9			
15	-60	7.25	Z-DNA poly d(GC) : poly d(GC)	ZZS1	67	ZZ1S	33					default Z form fiber in x3dna
16	-51	7.57	Z-DNA poly d(As4T) : poly d(As4T)	ZZS1	67	ZZ1S	33					
17	0	10.20	L-DNA (calf thymus) poly d(GC) : poly d(GC)	NANT	100							weird torsions (76 171 124 298 227 221 147 26 167 and 147 276 98 82 198 180 76 167 26)
18	36	3.23	B'-DNA alpha poly d(A) : poly d(T) (H-DNA)	BB07	50	BB04	50					chain A (BB07), chain B (BB04)
19	36	3.23	B'-DNA beta2 poly d(A) : poly d(T) (H-DNA beta)	AA00	50	BB00	50					chain A (AA00), chain B (BB00)
20	33	2.81	A-RNA poly (A) : poly (U)	AA00	100							
21	30	3.00	A'-RNA poly (I) : poly (C)	AA00	100							
22	33	2.56	Hybrid poly (A) : poly d(T)	AA00	100							
23	32	2.78	Hybrid poly d(G) : poly (C)	AA00	100							
24	36	3.13	Hybrid poly d(I) : poly (C)	AA00	100							
25	33	3.06	Hybrid poly d(A) : poly (U)	BB00	50	AA00	50					chain A (BB00), chain B (AA00)
26	36	3.01	10-fold poly (X) : poly (X)	-								non-standard base, not assigned
27	33	2.52	11-fold poly (X) : poly (X)	-								non-standard base, not assigned
28	33	2.60	Poly (s2U) : poly (s2U) (symmetric base-pair)	AA00	100							
29	33	2.60	Poly (s2U) : poly (s2U) (asymmetric base-pair)	AA00	100							
30	33	3.16	Poly d(C) : poly d(I) : poly d(C)	AA00	100							
31	30	3.26	Poly d(T) : poly d(A) : poly d(T)	AA00	100							
32	33	3.04	Poly (U) : poly (A) : poly (U) (11-fold)	AA00	100							
33	30	3.04	Poly (U) : poly (A) : poly (U) (12-fold)	AA00	67	AA03	33					
34	30	3.29	Poly (I) : poly (A) : poly (I)	AA00	100							
35	31	3.41	Poly (I) : poly (I) : poly (I) : poly (I)	AA04	100							
36	60	3.16	Poly (C) or poly (mC) or poly (eC)	AA00	100							
37	36	3.20	B'-DNA beta2 Poly d(A) : poly d(U)	BB04	50	BB00	50					chain A (BB04), chain B (BB00)
38	36	3.24	B'-DNA beta1 Poly d(A) : poly d(T)	BB07	50	BB04	50					chain A (BB07), chain B (BB04)
39	72	6.48	B'-DNA beta2 Poly d(AI) : poly d(CT)	BB00	50	BB01	50					0101 within chain
40	72	6.46	B'-DNA beta1 Poly d(AI) : poly d(CT)	BB04	50	BB00	50					4040 within chain
41	144	13.54	B'-DNA Poly d(AATT) : poly d(AATT)	BB00	51	BB04	26	BB07	23			~ 0470 within chain
42	33	3.04	Poly(U) : poly d(A) : poly(U) [cf. #32]	AA00	100							
43	36	3.20	Beta Poly d(A) : Poly d(U) [cf. #37]	BB04	50	BB00	50					chain A (BB04), chain B (BB00)
44	36	3.23	Poly d(A) : poly d(T) (Ca salt)	BB00	100							
45	36	3.23	Poly d(A) : poly d(T) (Na salt)	BB00	100							
46	36	3.38	B-DNA (BI-type nucleotides; generic sequence: A, C, G and T)	BB04	100							
47	40	3.32	C-DNA (BII-type nucleotides; generic sequence: A, C, G and T)	BB07	100							default BII form fiber in x3dna
48	88	6.02	D(A)-DNA ploy d(AT) : ploy d(AT) (right-handed)	AA00	100							
49	60	7.20	S-DNA ploy d(CG) : poly d(CG) (C_BG_A, right-handed)	BA01	50	AB03	50					
50	60	7.20	S-DNA ploy d(GC) : poly d(GC) (C_AG_B, right-handed)	BA01	50	AB03	50					
51	32	3.22	B*-DNA poly d(A) : poly d(T)	BB00	100							
52	90	6.06	D(B)-DNA poly d(AT) : poly d(AT) [cf. #48]	BB04	100							
53	-39	3.29	C-DNA (generic sequence: A, C, G and T) (deprecated)	NANT	100							strange torsions (147 290.3 176.2 58.9 220.2 157.6 147 177.1 177.1)
54	33	2.56	A-DNA (generic sequence: A, C, G and T) [cf. #1]	AA00	100							
55	36	3.39	B-DNA (generic sequence: A, C, G and T) [cf. #4]	BB04	100							

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- Models #1 to #41 are based on Struther Arnott: "Polynucleotide secondary structures: an historical perspective", pp. 1-38 in "Oxford Handbook of Nucleic Acid Structure" edited by Stephen Neidle (Oxford Press, 1999).
- Models #42 and #43 are from Chandrasekaran & Arnott: "The Structures of DNA and RNA Helices in Oriented Fibers", pp 31-170 in "Landolt-Bornstein Numerical Data and Functional edited by W. Saenger (Springer-Verlag, 1990).
- Models #44-445 based on Alexeev et al., "The structure of poly(dA)... poly(dT) as revealed by an X-ray fiber diffraction". J. Biomol. Str. Dyn, 4, pp. 989-1011, 1987.
- Models #46-447 based on van Dam & Levitt, "BI nucleotides in the B and C forms of natural-sequence polymeric DNA: a new model for the C form of DNA". J. Mol. Biol., 304, pp. 541-561, 2000.
- Models #48-455 based on Premilat & Albiser, "A new D-DNA form of poly(dA-dT).poly(dA-dT): an A-DNA type structure with reversed Hoogsteen Pairing". Eur. Biophys. J., 30, pp. 404-410, 2001 (and several other publications).

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