

Supporting Information

Preparation and X-ray analysis of 2,3-dichlorophenylglucosinolate

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Table contents

1. Synthesis	
1.1. General methods.....	S2
1.2. 2,3-Dichlorobenzaldehyde oxime 2	S2
1.3. 2,3-Dichlorobenzohydroxymoyl chloride 3	S3
1.4. 2,3,4,6-Tetra-O-acetyl- β -D-glucopyranosyl thiol 8	S3
2. NMR spectrum	

Compound Number	Page	¹ H NMR Spectrum	¹³ C NMR Spectrum	ORTEP Plot
2	S5	√	√	
3	S6	√	√	
8	S7	√	√	
9	S8	√	√	
10	S9	√	√	
11	S10	√	√	
9	S11			√
11	S12			√

1. Synthesis

1.1. General Procedures

Melting points (mp) were recorded on a hot stage apparatus and are uncorrected. Optical rotations were measured at the stated temperatures in the stated solvent on a polarimeter at the sodium d-line (589 nm); $[\alpha]_D$ values are given in $10^{-1} \text{ deg cm}^2 \text{ g}^{-1}$. Infrared spectra (ν_{max}) were recorded on a FT-IR spectrometer. Samples were analyzed as KBr drift (for solids) or as thin films on NaCl plates (for liquids/oils). Unless otherwise specified, proton (^1H) and carbon (^{13}C) NMR spectra were recorded on a 300 MHz spectrometer operating at 300 MHz for proton and 75 MHz for carbon nuclei. Chemical shifts were recorded as δ values in parts per million (ppm). Spectra were acquired in deuterated chloroform (CDCl_3) at 300 K unless otherwise stated. For ^1H NMR spectra recorded in CDCl_3 , the peak due to residual CHCl_3 (δ_{H} 7.24) was used as the internal reference, while the central peak (δ_{C} 77.0) of the CDCl_3 triplet was used as the reference for proton-decoupled ^{13}C NMR spectra. Low-resolution mass spectra were measured on a mass spectrometer at 300 °C and scan rate of 5500 m/z /second using either water/methanol/acetic acid in a ratio of 0/99/1 or 50/50/1 as a mobile phase. Accurate mass measurement was by mass spectrometry with a heated electrospray ionisation (HESI) source. The mass spectrometer was operated with full scan (50-1000 amu) in positive or negative FT mode (at a resolution of 100,000). The analyte was dissolved in water/methanol/acetic acid in a ratio of 0/99/1 or 50/50/1 and infused via syringe pump at a rate of 5 $\mu\text{L}/\text{min}$. The heated capillary was maintained at 320 °C with a source heater temperature of 350 °C and the sheath, auxiliary and sweep gases were at 40, 15 and 8 units respectively. Source voltage was set to 4.2 kV. Solvents were dried over standard drying agents and freshly distilled before use. Ethyl acetate and hexane used for chromatography were distilled prior to use. All solvents were purified by distillation. Reactions were monitored by TLC on silica gel 60 F₂₅₄ plates with detection by UV fluorescence or charring with a basic potassium permanganate stain. Flash column chromatography was performed on silica gel 60 particle size 0.040-0.063 μm (230–400 mesh).

1.2. 2,3-Dichlorobenzaldehyde oxime 2.

Hydroxylamine hydrochloride (2.50 g, 36 mmol) was added to a solution of the aldehyde **1** (5.25 g, 30 mmol) in MeOH (60 mL) followed by pyridine (3.0 mL, 30 mmol). The reaction

was stirred at rt for 2.5 h. After the MeOH was removed *in vacuo*, the residue was suspended in DCM (120 mL) and washed with 1 M HCl solution (3 × 30 mL), H₂O (3 × 30 mL) and brine solution. The organic phase was dried (Na₂SO₄) and then concentrated at reduced pressure. The compound **2** was obtained by re-crystallization with hexane/EtOAc as white crystals (5.64 g, 99%). *R*_f 0.61 in hexane/ethyl acetate (7:3), mp 121-122 °C. δ_H (300 MHz, CDCl₃, 300 K) 8.57 (s, 1H, CH=N), 8.13 (s, 1H, OH), 7.73 (dd, *J*_{4,5} 8.1, 1H, H4), 7.49 (dd, *J*_{5,6} 7.8, 1H, H6), 7.20 (dd, *J*_{5,6} 7.8, *J*_{4,5} 8.1, 1H, H5). δ_C (75 MHz, CDCl₃, 300 K) 147.2 (C=N), 133.4 (C-3), 131.8 (C-2), 131.5 (C-4), 131.2 (C-1), 127.1 (C-6), 125.0 (C-5). HRMS (ESI) *m/z* for C₇H₅Cl₂NO [M+H]⁺, calcd 189.9821, found 189.9813.

1.3. 2,3-Dichlorobenzohydroxymoyl chloride **3**

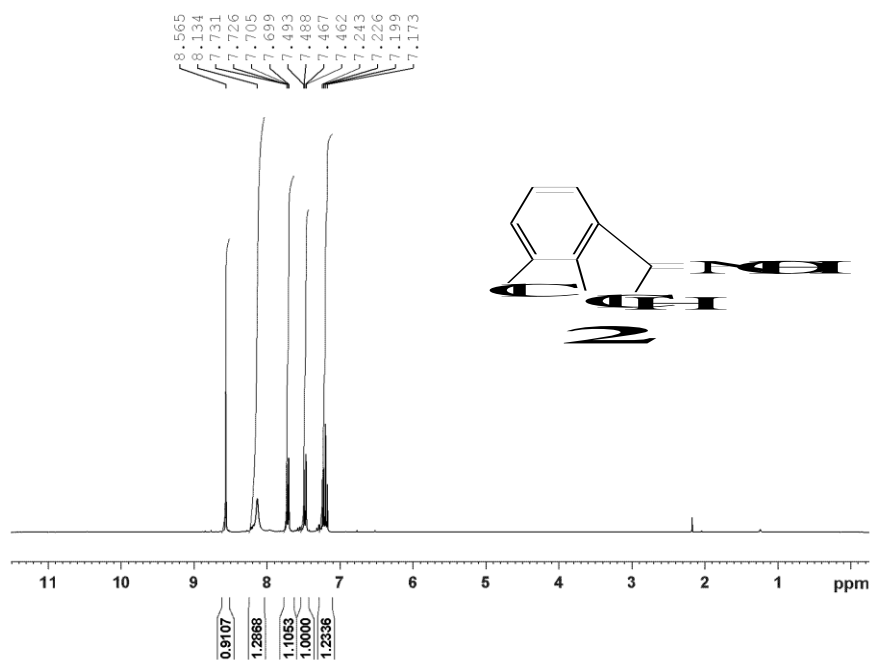
One-fifth of *N*-chlorosuccinimide (4.46 g, 33.4 mmol) (CAUTION: induction period) was added initially to a solution of the oxime **2** (5.90 g, 31.4 mmol) in DMF (60 ml). The reaction was cooled and stirred in an ice bath (for about 30 minutes) for the reaction happened (as indicated by a slight temperature rise). The reaction temperature was kept under 35 °C, while the addition of the NCS was repeated until all the rest of NCS was added. The reaction mixture was allowed to reach rt over 4 h. After that, the reaction mixture was poured into ice-water and extracted with Et₂O (3 × 20 ml). The organic layers were washed with water, dried over MgSO₄ and concentrated under reduced pressure. The pure hydroxymoyl chloride **3** obtained by chromatograph eluting with 80% hexane/EtOAc as white crystals (6.04 g, 87%). *R*_f 0.63 in hexane/ethyl acetate (2:3), mp 112-113 °C. ν_{max} (NaCl)/cm⁻¹ 3392 (OH), 1629 (C=N), 1462, 1409, 1242. δ_H (300 MHz, CDCl₃, 300 K) 8.76 (s, 1H, OH), 7.55 (d, *J* 8.1, 1H, H4), 7.36 (d, *J* 7.8, 1H, H6), 7.27 (dd, *J*_{4,5} 8.1, *J*_{5,6} 7.8, 1H, H5). δ_C (75 MHz, CDCl₃, 300 K) 136.4 (C=N), 134.2 (C-3), 133.8 (C-2), 131.7 (C-4), 131.6 (C-1), 128.8 (C-6), 127.1 (C-5). HRMS (ESI) *m/z* for C₇H₅Cl₃NO [M+H]⁺, calcd 223.9431, found 223.9421.

1.4.2,3,4,6-Tetra-O-acetyl-β-D-glucopyranosyl thiol **8**

To a solution of D-glucose **4** (3.00 g, 16.6 mmol) in dry pyridine (33 mL) at 0 °C under a nitrogen atmosphere was slowly added acetic anhydride (31.5 mL, 333 mmol). The reaction mixture was stirred at 0 °C for 1 h before a catalytic amount of DMAP (200 mg, 1.67 mmol) was added. As the reaction mixture was allowed to reach rt, it becomes slightly exothermic.

After 6 h, the clear yellow mixture was slowly poured into rapidly stirred ice-water (125 mL), giving a sticky solid. After EtOAc extraction (3×45 mL), evaporation of the solvent and co-evaporations with dry toluene (3×20 mL), peracetylated glucose was obtained as a yellow solid (5.84 g, 90%). A solution of pentaacetyl-D-glucopyranose **5** (2.00 g, 5.1 mmol) in DCM (20 mL) was stirred in an ice bath while HBr/HOAc (6 mL, 45 wt%) was added drop-wise. After an hour, the solution was washed with ice-water and cold saturated NaHCO₃ solution, dried over MgSO₄ and concentrated to leave the glucosyl bromide **6** as a pale yellow oil (1.83 g). The oil was dissolved in dry acetone (20 mL), and the solution was added to freshly activated 4Å molecular sieves (2 g) and thiourea (500 mg, 6.6 mmol). The mixture was maintained at reflux temperature (60 °C) under a nitrogen atmosphere for 2.5 h, cooled and filtered through Celite. Solvent removal and trituration of the syrupy residue with hexane (3×20 mL) gave the isothiuronium bromide **7** as a colorless amorphous power. The crude product was dissolved in DCM (20 mL), a solution of Na₂S₂O₅ (2.00 g) in water (20 mL) was added, and the mixture was maintained at reflux under a nitrogen atmosphere for an hour. After cooling, the organic layer was separated and washed with water, saturated NaCl solution, dried over Na₂SO₄, filtered and concentrated under reduced pressure to give a residue. Pure **8** was obtained by flash column chromatography on silica gel eluting with 0-3% MeOH/DCM as a solid (1.60 g, 86%). *R*_f 0.3 in 50% hexane/EtOAc, mp 114–115 °C (Lit. 113–114 °C)(Fujihira *et al.*, 2003). $[\alpha]_{\text{D}}^{20} +10.5$ (*c* 1.0, CHCl₃) (Lit. +11)(Fujihira *et al.*, 2003). δ_{H} (300 MHz, CDCl₃, 300 K) 5.02–5.18 (m, 2H, H3 and H4), 4.93 (dd, *J*_{1,2} 9.6, *J*_{2,3} 9.3, 1H, H2), 4.51 (dd, *J*_{1,2} 9.6, *J*_{1,SH} 9.9, 1H, H1), 4.23 (dd, *J*_{5,6b} 4.8, *J*_{6a,6b} 12.3, 1H, H6b), 4.11 (dd, *J*_{5,6a} 2.4, *J*_{6a,6b} 12.3, 1H, H6a), 3.66–3.17 (m, 1H, H5), 2.29 (d, *J*_{1,SH} 9.9, 1H, SH), 1.96–2.08 (4 × br s, 12H, CH₃COO). δ_{C} (75 MHz, CDCl₃, 300 K) 170.2, 169.7, 169.2, 168.9 (4 × CH₃COO), 87.3 (C-1), 75.9 (C-3), 73.2 (C-2, C-3), 67.7 (C-4), 61.6 (C-6), 20.6, 20.3(2), 20.2 (4 × CH₃COO). HRMS (ESI) *m/z* for C₁₄H₁₉O₉S [M-H][−], calcd 363.0755, found 363.0746.

h1-qv-01-29-01

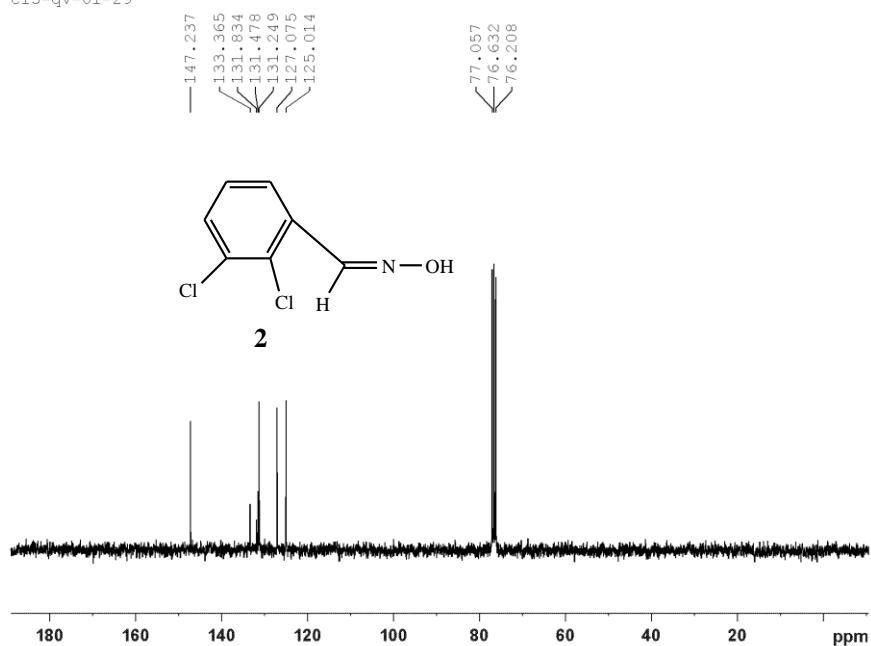


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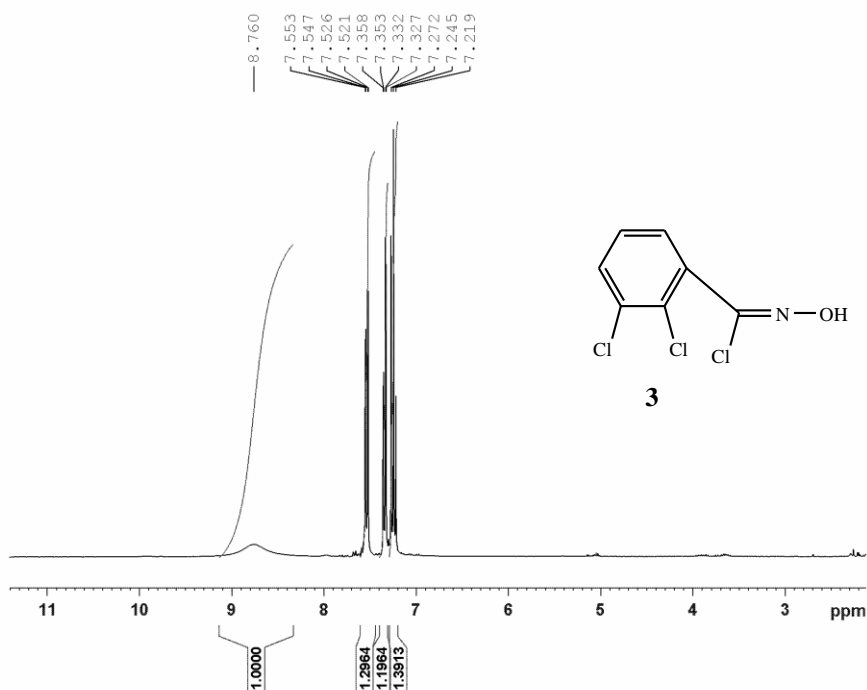
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D11           0.03000000 sec
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PL1           -1.00 dB
SFO1          75.4745111 MHz

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QV-01-30



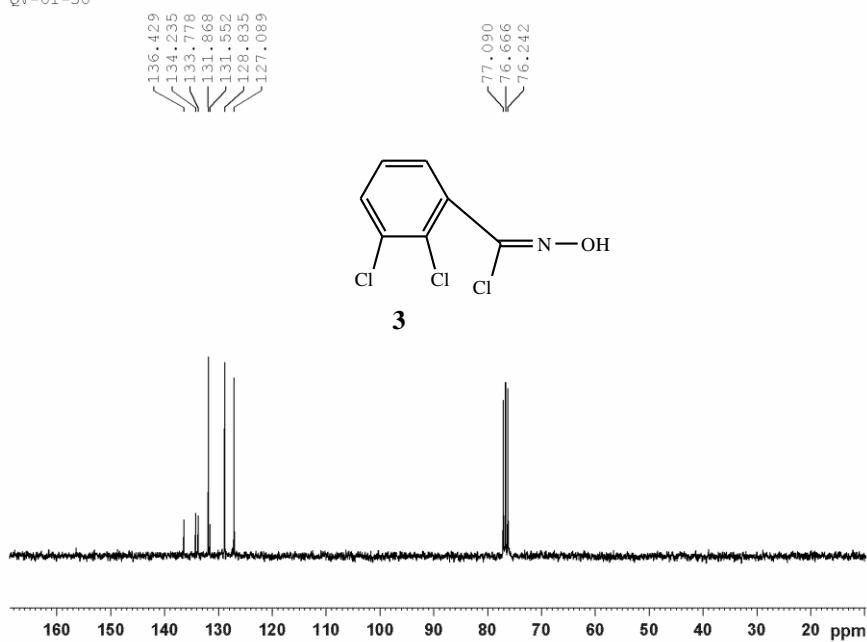
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QV-01-30



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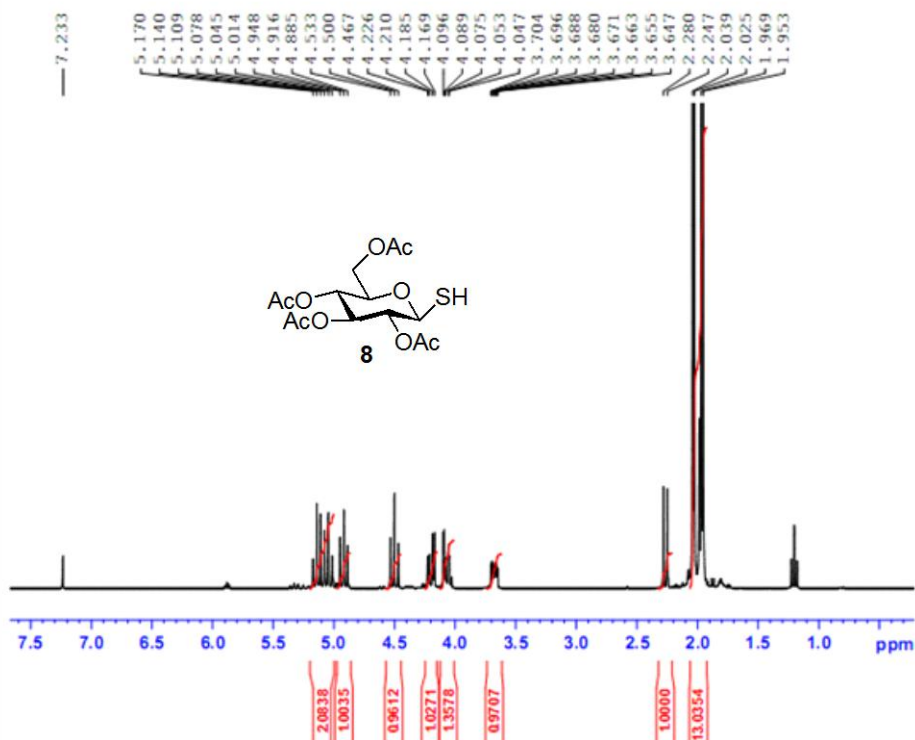
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RG            14596.5
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DE            6.00 usec
TE            300.0 K
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D11           0.03000000 sec
TD0           1

===== CHANNEL f1 =====
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SFO1          75.4745111 MHz

===== CHANNEL f2 =====
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PCPD2         77.00 usec
PL2           -1.00 dB
PL12          17.00 dB
PL13          17.00 dB
SFO2          300.1317000 MHz
SI            16384
SF            75.4677770 MHz
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GB            0
PC            1.40

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QV-01-04-02-08-11 thio glucose CDCl3

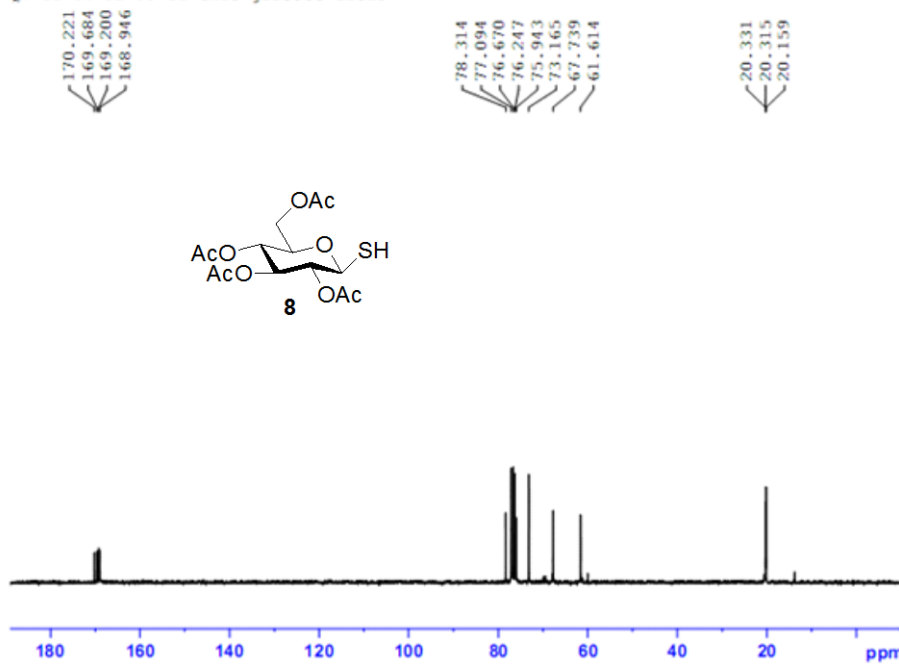


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QV-01-04-02-08-11 thio glucose CDCl3



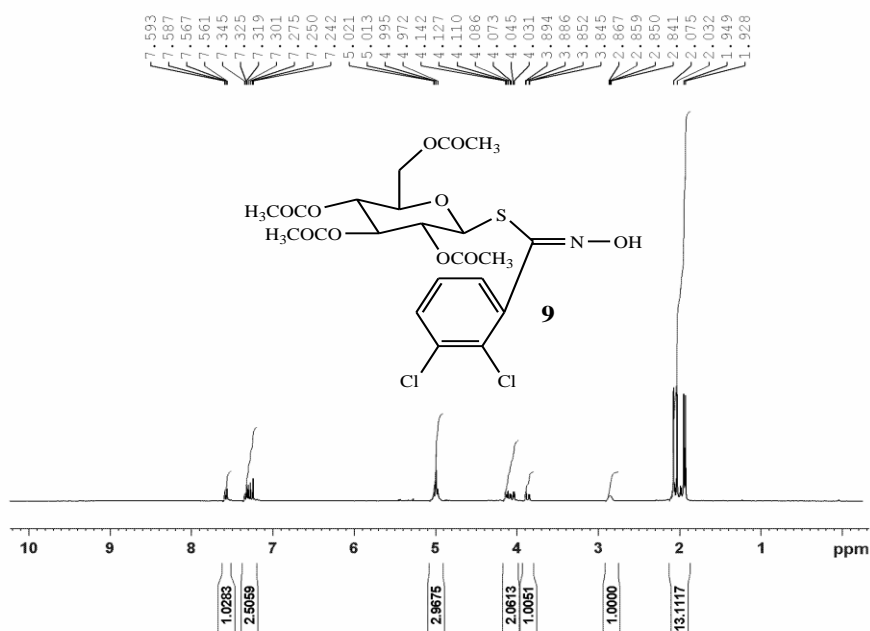
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TE         300.2 K
D1         1.00000000 sec
D11        0.03000000 sec
TDO        1

===== CHANNEL f1 =====
NUC1       13C
P1         9.00 usec
PL1        -1.00 dB
SFO1       75.4745111 MHz

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2     77.00 usec
PL2        -1.00 dB
PL12       17.00 dB
PL13       17.00 dB
SFO2       300.1317000 MHz
SI         16384
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h1-qv-01-31

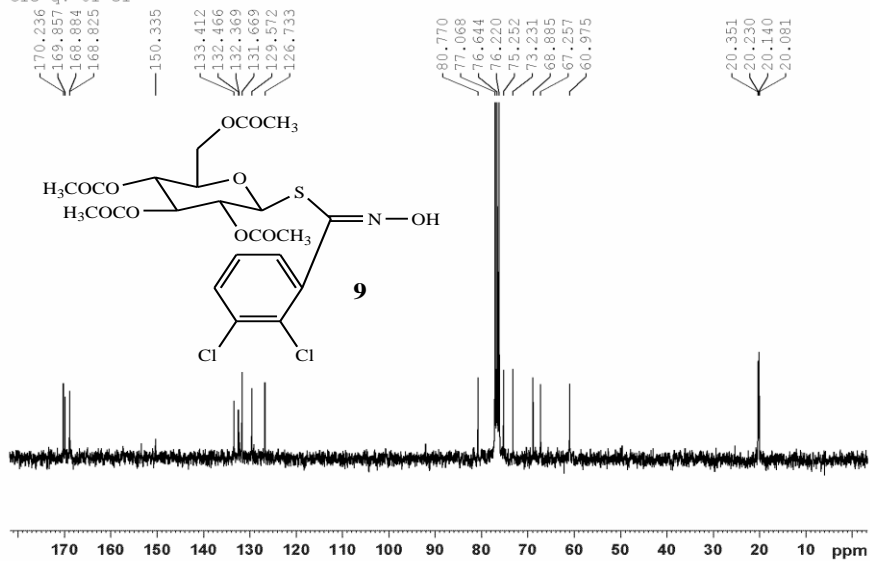


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C13-qv-01-31

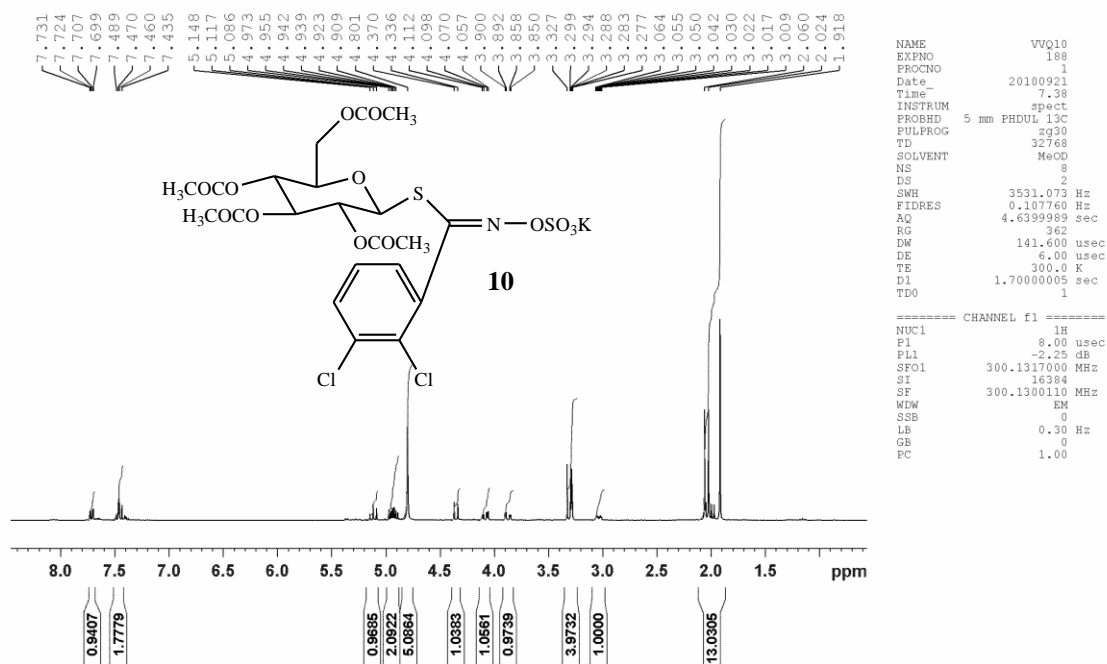


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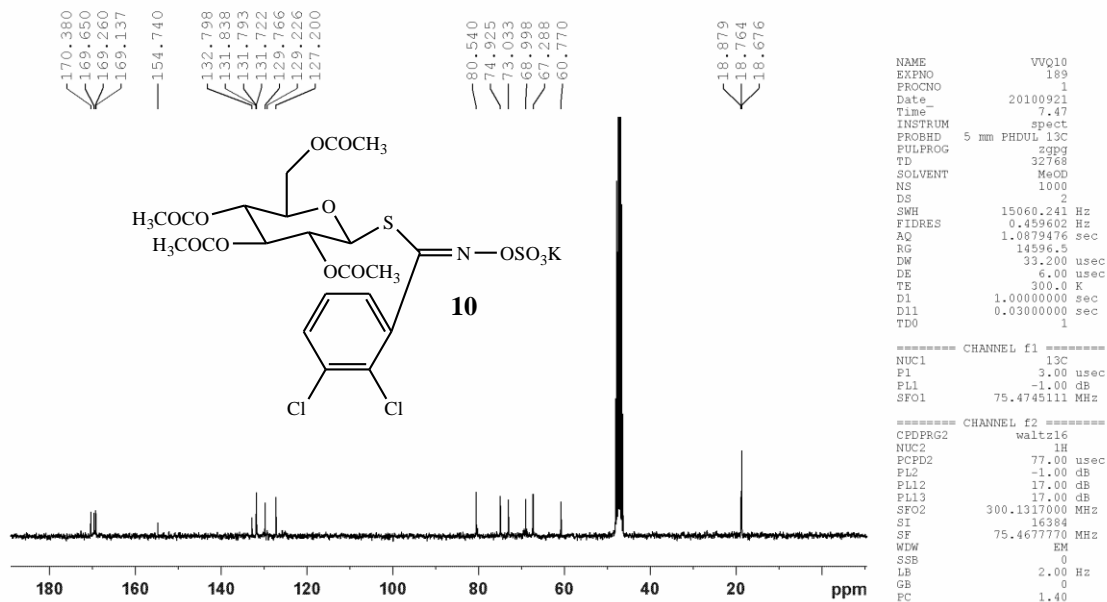
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FIDRES        0.459602 Hz
AQ            1.0879476 sec
RG            14596.5
DW            33.200 usec
DE            6.00 usec
TE            300.0 K
D1            1.00000000 sec
D11           0.03000000 sec
TD0           1
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NUC1          13C
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PL1           -1.00 dB
SFO1          75.4745111 MHz
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NUC2          1H
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PL12          17.00 dB
PL13          17.00 dB
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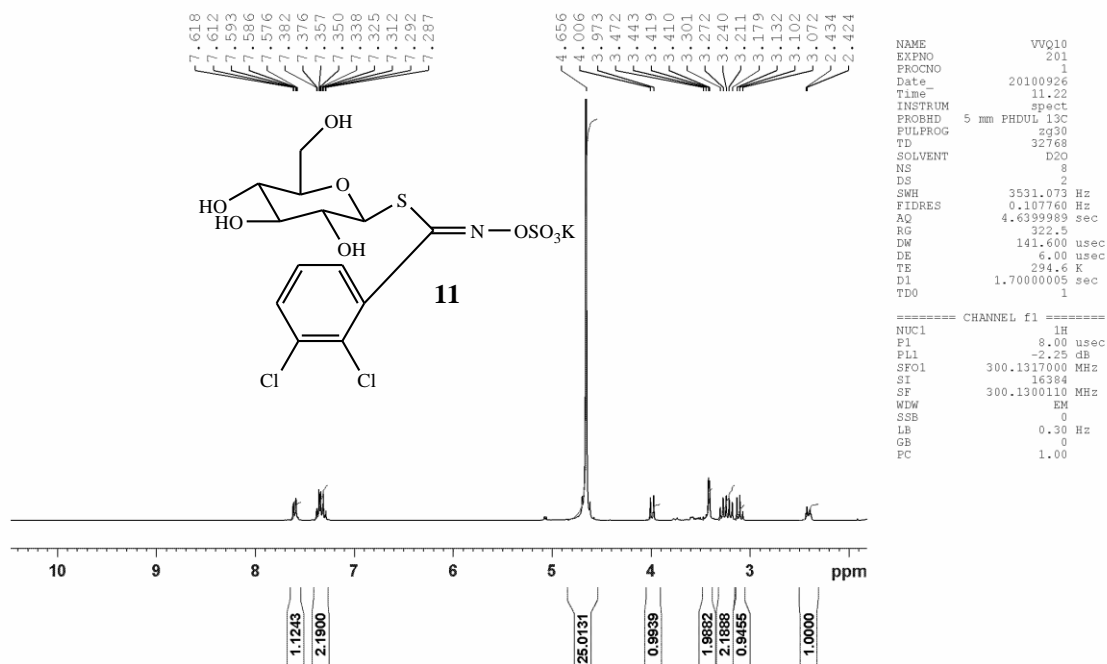

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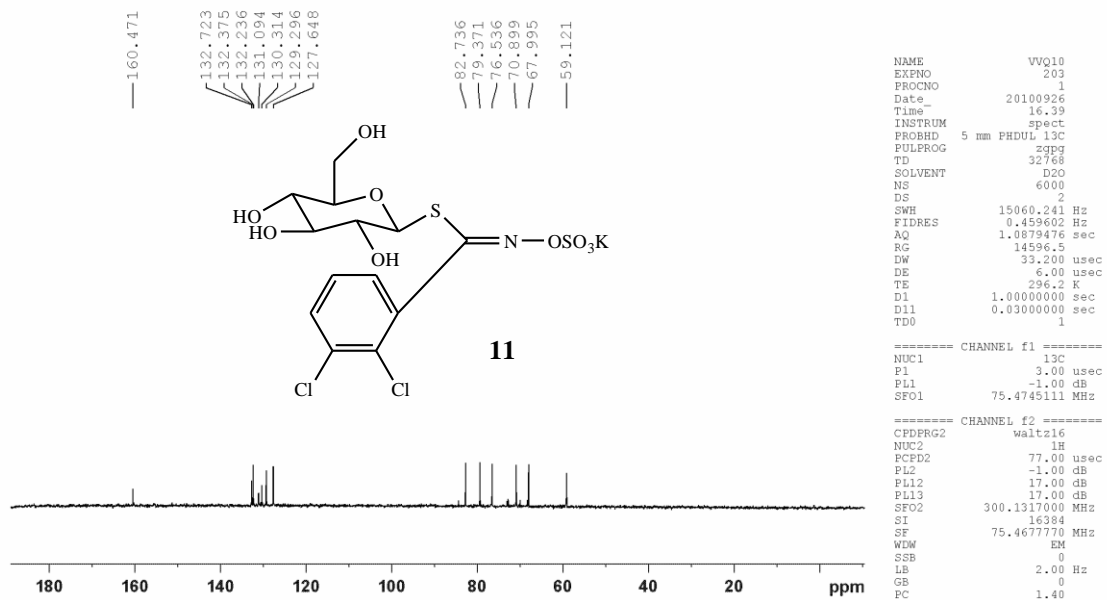
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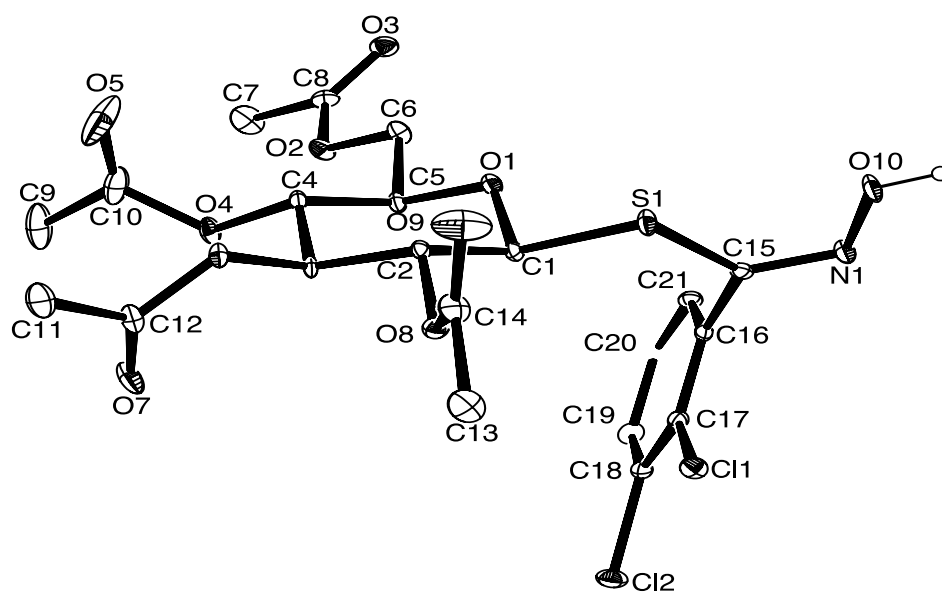


H1-QV-01-33-01



C13-QV-01-33





ORTEP plot of Compound **9**.

