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Supporting information for article:

Crystal engineering of an adenine–decavanadate molecular device towards label-free chemical sensing and biological screening

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I. Development and optimization of a synthesis method for compound 1

Details of all experiments have been tabulated here. For simplification, our method for the synthesis of adeninium decavanadate (reported method in the manuscript) has been referred to as "**method A**".

- ,	Solvothermal	condition	experiments:
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Test	Variation	Result	Description
code			
011	Method A at room temp*	green precipitation	-
012	Method A under reflux conditions	green precipitation	-
013- 015	Method A at 100, 90 & 80°C solvothermal conditions	mixture of green precipitations and gel	By decreasing the reaction temperature percentage of the gel product was increased. At 80 °C the sole product was a green metallogel which is the subject of another ongoing research in our group.
016	Method A + at 120°C solvothermal conditions	same as main reaction	Yield of the reaction didn't increase in comparison with 110 °C

* Adenine was dissolved in hot deionized water after which the adenine solution was cooled to RT. The BTC and VOSO₄ were both added and mixed for 4 hours at RT.

As described in **method A**, upon completion of the hydrothermal reaction at 110 °C for 72h and then 4°C/h cooling rate to RT, the resulting solution contained yellow crystals of adeninium decavanadate. In brief, according to the above table, by altering the temperature parameter of the solvothermal conditions, we didn't obtain adeninium decavanadate: $[AdH]_6[V_{10}O_{28}].4(H_2O)$.

- 1,3,5-benzenetricarboxylic acid	extra reagent experiments:
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Test code	Variation	Result	Description
021	Method A without BTC	orange solution	Slow evaporation of the resulted solution didn't give any crystals even until very small

027- 028	Method A (BTC=0.1 & 0.05 mmol)	Same as main reaction (less crystals)	By increasing the BTC concentrations yield of the reaction was dropped drastically.
025- 026	Method A (BTC=0.2 & 0.15 mmol)	same as main reaction + white precipitations	By increasing the BTC concentration the yields of the reactions were equal in comparison with the main reaction but some white precipitates appeared.
024	021 + reactants' concentrations×3	orange solution	same as 021 test results
023	021 + reactants' concentrations×2	orange solution	same as 021 test results
022	021 + at 120°C solvothermal conditions	orange solution	-
			volumes remains clear (negligible colloidal orange precipitates were observed).

As described in **method A**, a small amount of colloidal green precipitates was formed shortly after addition of vanadyl sulfate (VOSO₄) that certainly confirms the presence of V^{+3} . Upon completion of the hydrothermal reaction, the resulting solution contained yellow crystals of adeninium decavanadate, alongside the green precipitate (small amount).

But when the reaction was carried out in the absence of BTC, only a clear yellow solution (no yellow crystals nor green precipitations) were obtained, the resulting solution was subjected to slow evaporation but no crystals were grown and only in the last few drops of solution a negligible amount of colloidal orange precipitates was obtained (test code 021).

Thus, it could be concluded that V^{+4} (vanadyl sulfate) in the presence of BTC, during a redox reaction under hydrothermal condition, leads to the formation of a green precipitate (containing V^{+3}) and adeninium decavanadate crystals (containing V^{+5}).

- NH₄VO₃ (vanadium cheaper salt) experiments:

Test	Variation	Result	Description
code			

031	Method A + salt NH ₄ VO ₃ was	green solution	At the beginning orange solution and at the
	used instead of VOSO4		end of the reaction green solution appeared.
032	031 without BTC	mixture of white	-
		and green	
		precipitations	
033	031 + 2 <ph<3**< th=""><th>colorless</th><th>colorless solution with small amounts of</th></ph<3**<>	colorless	colorless solution with small amounts of
		solution	white precipitates
034	NH ₄ VO ₃ + 2 <ph<3+ th="" without<=""><th>orange poor-</th><th>XRD pattern of obtained powder was</th></ph<3+>	orange poor-	XRD pattern of obtained powder was
	BTC+ without DMF +not under	quality crystals	different from adeninium decavanadate
	solvothermal conditions***	1 5 5	XRD pattern

*** NH₄VO₃ was used instead of VOSO₄ as cheaper reactant and the 2<pH<3 was adjusted by dilute HCl
*** Sánchez-Lara, E., Treviño, S., Sánchez Gaytán, B. L., Sánchez-Mora, E., Castro, M. E., Meléndez-Bustamante, F. J., ... & González-Vergara, E. (2018). *Frontiers in chemistry*, 6, 402.

- DMF key factor:

Test	Variation	Result	Description
code			
041	Method A without	Mixture of white	-
	DMF solvent	and green	
		precipitates.	

- pH key factor

Test	Variation	Result	Description
code			
051-	Method A (pH=6,	same as main	Yield of the reactions were almost equal to the
-55	6.3, 6.5, 6.8 & 7)	reaction	main reaction
056-	method A (pH= 5.8	Same as the main	Yield of the reactions were dropped drastically.
057	& 7.3)	reaction (less crystals)	

In summary, according to the results of the above tests, it appears that all the components VOSO₄, BTC, DMF, and hydrothermal conditions at 110°C for three days are critical for the production of the reported adeninium decavanadate compound.

II. Compound 1 bio-sensing feasibility

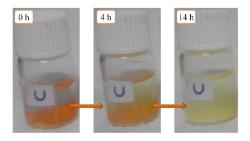


Figure S1: Bio-sensing feasibility of the compound 1 after (a) 0 h; (b) 4 h and (c) 14 h.

- Compound 1 optical assessments in solid-state:
- *Sample preparation:* Appropriate amount of crashed crystals (3 mg) of compound **1** transferred into an vessel, which then charged with 3ml of nucleic acid solutions (3M) in phosphate-buffered saline (PBS) solution, pH=7.4. The interaction between **1** and nucleic acids were monitored by UV-visible and fluorescence measurements and XRD patterns approved this changes.

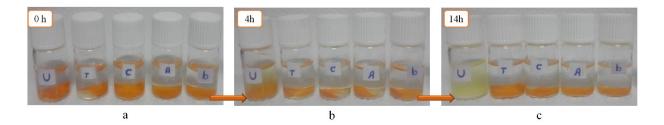


Figure S2: Crashed crystals of compound **1** incubated with diffident nucleic acids (U: Uracil, T: Thymine, C: Cytosine, G: Guanine, A: Adenine and B: blank), after (a) 0 h; (b) 4 h and (c) 14 h.

III. Compound 1 optical assessments in solution form:

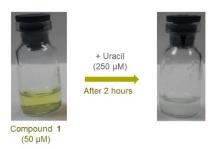


Figure S3: Feasibility of the bio-sensing of compound 1 in solution form.

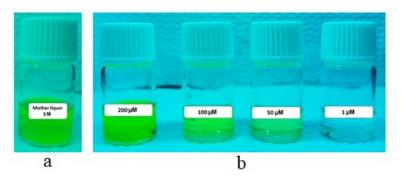


Figure S4: (a) Mother liquor of compound 1 (3 M), (b) Compound 1 samples to find optimum concentration through optical assessments

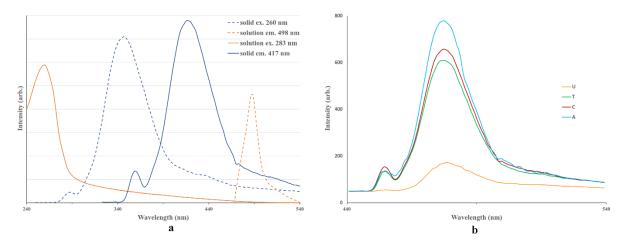


Figure S5: a) Fluorescent spectrum of compound 1 in solid state and solution form; and b) Fluorescent intensities of compound 1 solution in the presence of nucleobases (U, T, C and A)

IV. Biological assays

- Sample preparation for biological assay: Mother liquor of compound 1 (3M) for UV-Visible and fluorescent data collection was prepared by DMSO solvent after 3 hours stirring at ambient temperature.
- Sample preparation to find optimum concentration: Then to find optimum concentration for optical biological assessments, diluted samples of 1 (1, 50, 100, 150, 200 μM) were prepared over adding proper amount of fresh phosphate-buffered saline (PBS) solution (pH=7.4) to the mother liquor.
- Sample preparation for Nucleic acid sensing study: For biosensing comparison between solid and solution forms' of compound **1**, Nucleic acid solutions (250 μ M) were incubated with **1** (50 μ M) for 4 hours at RT in total 3ml PBS. Similar to solid-state optical assessments, the interaction between **1** and the nucleic acids were monitored by fluorescent optical measurements at 283 nm excitation wavelength (see section 3.3.1) (Fig. S5b).
- Sample preparation for Hydrolysis rate study: Hydrolysis rate of the compound 1 containing 50 μM (see section 3.2.2) have been evaluated vs time (from fresh and after 2, 6, 9, 12 and 24 hours after preparation).
- *Cell culture:* Human mammary carcinoma MDA-MB-231 and MCF7 cell lines were purchased from National Cell Bank of Iran (Pasteur Institute, Iran). Both cell lines were fed with RPMI-1640 (Gibco). This culture medium was supplemented with 10% FBS (Gibco) and 1% antibiotics (100 U/ml penicillin and 100µg/ml streptomycin). Cells were maintained at 37°C in a humidified incubator containing 5% CO₂ and were passaged using trypsin/EDTA (Sigma-Aldrich) and phosphate-buffered saline (PBS) solution.
- Anticancer study: The cytotoxicity of the compound 1 in solution was evaluated by 3-(4,5- dimethylthialzol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. MDA-MB-231 and MCF7 cells were seeded into 96-well plates at a density of 1.5x10⁴ per well in 200 μl of media and grown overnight. The cells were incubated with various concentrations (3.125/6.25/12.5/25/50/100/200μM) of compound 1 for 24 h at 37°C under 5% CO₂. Following this incubation, cells were incubated in medium containing

0.5 mg/ml of MTT (Sigma-Aldrich) for 4 h. The medium was discarded, and the precipitated formazan violet crystals were dissolved in 100 μ L of DMSO to solubilize the formazan. After shaking the plate for 20 min, the absorbance of the sample was measured at 570nm using an ELISA reader (Model wave xs2 Biotek, USA). The absorbance of dissolved formazan in the visible region correlates with the number of intact active cells.

Cell permeability: Internalization of the compound **1** was visualized using the fluorescence microscope (Nikon, TS-100) equipped with an appropriate filter set. MDA-MB-231 and MCF7 cells (1×10^4 cells/well) were seeded in 6-well plates (or 60mm^2 plate) overnight before experiments. Compound **1** were added into the incubation medium at concentration of 50µM for 1h incubation in 5% CO₂ at 37°C. Both cells were washed twice with phosphate-buffered saline (PBS). Microscopic images in the green channel for detection of the compound **1** and in the bright-field were obtained by fluorescence microscopy.

Nitrogen atom	Angle	Degree (°)
N8	C7-N8-C9	124.2(2)
N18	C17-N18-C19	122.2(2)
N28	C27-N28-C29	121.6(2)
N4	C3-N4-C5	106.59(19)
N14	C13-N14-C15	106.52(19)
N24	C23-N24-C25	106.31(19)
N6	C5-N6-C7	112.4(2)
N16	C15-N16-C17	112.3(2)
N26	C25-N26-C27	111.7(2)
N2	C1-N2-C3	103.81(19)
N12	C11-N12-C13	104.13(19)
N22	C21-N22-C23	103.96(18)

Table S1. Internal angles (°) at the N atoms of adeniniums (AdH⁺1, AdH⁺2, AdH⁺3)

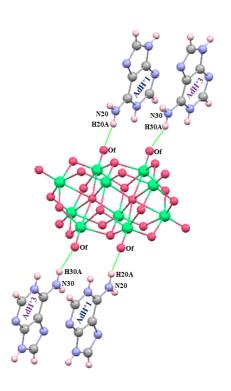


Figure S6: Decavanadate and adeniniums interactions