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Supporting information for article:

Anaerobic fixed-target serial crystallography
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Table S1 LIC primers used in the cloning of the codon-optimised VioC into pNIC28-Bsa4 vector.

| Oligo name | Sequence (5'-3') | $\operatorname{Tm}\left({ }^{\circ} \mathrm{C}\right)$ |
| :--- | :--- | :--- |
| LIC_VioC fw | TACTTCCAATCCATGATGACCGAAAGT | 85.5 |
|  | CCGACCACACATCATGG |  |
| LIC_VioC rev | TATCCACCTTTACTGGCGCTGACCCAG | 83.3 |
|  | AACACGGC |  |

Table S2 Data collection and refinement statistics. Absorbed doses were calculated using RADDOSE-3D (Bury et al., 2018).

| Datasets | VioC:Fe:2OG: <br> Arg (PDB ID: <br> 6Y0N) | VioC:Fe:SIN: <br> (3S)-OH-Arg <br> (PDB ID: <br> 6Y12) | AlkB:Fe:2OG: <br> T-1meA-T(PDB <br> ID: 6Y0Q) | AlkB:Fe:2OG <br> (PDB ID: <br> 6YPV) | IPNS:Fe:ACV <br> (PDB ID: <br> 6Y0O) |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Data Collection |  |  |  |  |  |
| Beamline (Wavelength, $\AA$ ) and detector | DLS I24 (0.9686), Pilatus3 6M |  |  |  |  |
| Data Processing | DIALS (integration), cctbx.xfel (merging) |  |  |  |  |
| Diffraction weighted dose / kGy | 36 | 36 | 68 | 68 | 38 |
| Number of images used | 12318 | 16594 | 11778 | 4640 | 9449 |
| Space group | C2 | C2 | P 1 | P1 | $\mathrm{P} 2_{1} 2_{1} 2_{1}$ |
| Cell <br> dimensions |  |  |  |  |  |
| $a, b, c$ ( $\AA$ ) | 82.0, 66.6, 63.6 | 82.4, 67.1, 64.0 | 36.4, 39.1, 40.8 | $\begin{aligned} & 37.0,39.1, \\ & 40.9 \end{aligned}$ | $\begin{array}{\|l} 41.9,75.8 \\ 102.0 \end{array}$ |
| $\alpha, \beta, \gamma\left({ }^{\circ}\right)$ | 90, 110.2, 90 | 90, 110.2, 90 | 79.0, 78.0, 66.9 | $\begin{aligned} & 78.1,76.0, \\ & 66.5 \end{aligned}$ | 90, 90, 90 |
| No. of | 1 | 1 | 1 | 1 | 1 |


| molecules/ASU |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| No. reflections | 27077 (2700) | 36065 (3574) | 20346 (1028) | 11994 (593) | 16880 (600) |
| Resolution (A) | $\begin{aligned} & 59.69-1.86 \text { (1.93- } \\ & 1.86) \end{aligned}$ | $\begin{aligned} & 50.69-1.70(1.76- \\ & 1.70) \end{aligned}$ | $\begin{aligned} & 35.67-1.75(1.81- \\ & 1.75) \end{aligned}$ | $\begin{aligned} & 39.35-2.10 \\ & (2.13-2.10) \end{aligned}$ | $\begin{aligned} & 60.85-2.20 \\ & (2.24-2.20) \end{aligned}$ |
| $\mathrm{R}_{\text {split }}$ | 0.232 (0.820) | 0.215 (0.841) | 0.178 (0.789) | 0.333 (0.617) | 0.235 (1.004) |
| <I> | 74992 (6733) | 59601 (4048) | 6663 (660) | 11298 (1965) | 65916 (16988) |
| $<\mathrm{I} / \sigma(\mathrm{I})>$ | 28.8 (2.3) | 34.2 (2.3) | 37.2 (2.8) | 48.4 (8.795) | 15.0 (1.4) |
| CC-1/2 | 0.94 (0.28) | 0.95 (0.26) | 0.98 (0.26) | 0.87 (0.41) | 0.96 (0.18) |
| Completeness (\%) | 100 (100) | 99.9 (99.4) | 100 (100) | 100 (99.9) | 97.7 (99.8) |
| Multiplicity | 86.89 (10.86) | 62.41(10.54) | 57.57 (10.60) | 25.8 (12.92) | 130.22 (7.96) |
| Wilson B value $\left(\AA^{2}\right)$ | 22.73 | 19.93 | 18.39 | 23.04 | 24.74 |
| Refinement | PHENIX |  |  |  |  |
| $\mathrm{R}_{\text {work }} / \mathrm{R}_{\text {free }}$ | 0.1886/0.2283 | 0.1819 /0.2103 | 0.1622/0.1914 | 0.2195/0.2506 | 0.2088/0.2474 |
| No. atoms | 2677 | 2787 | 1738 | 1593 | 2761 |
| - Enzyme | 2515 | 2574 | 1548 | 1510 | 2614 |
| - Ligand | 11 | 9 | 54 | 11 | 36 |
| Water | 151 | 204 | 136 | 72 | 112 |
| Average Bfactors | 33.26 | 26.87 | 24.24 | 26.83 | 30.09 |
| - Enzyme | 32.91 | 26.08 | 22.75 | 26.44 | 29.94 |
| - Ligand | 53.12 | 23.44 | 38.27 | 33.70 | 40.68 |
| - Water | 37.73 | 36.93 | 35.65 | 33.98 | 30.21 |
| R.m.s <br> deviations |  |  |  |  |  |
| $\begin{aligned} & -\quad \text { Bond } \\ & \text { lengths }(\AA) \end{aligned}$ | 0.003 | 0.004 | 0.004 | 0.004 | 0.002 |
| $\begin{aligned} & -\quad \text { Bond } \\ & \text { angles }\left({ }^{\circ}\right) \end{aligned}$ | 0.62 | 0.71 | 0.68 | 0.54 | 0.49 |



Figure S1 Fluorescence measurements. (a) Exemplary raw and smoothened accumulated optical spectra taken at the beginning ( $\mathrm{t}_{0}$, magenta) and the end of the kinetic series ( $\mathrm{t}=265 \mathrm{~min}$, black) for the "5" type of setup indicated in Fig. 2(d). (b) Exemplary plot showing decay of the fluorescence signal for the " 3 " type of setup indicated in Fig. 2(d) plotted as the absolute difference between the number of counts at 615 nm with 550 nm (dots) fitted to a single exponential decay model (solid line), from which a $\mathrm{t}_{0.5}=13.7$ minutes was obtained.
(a)

(c)


(f)


Figure S2 Structural comparison of the anaerobic and $\mathrm{O}_{2}$ exposed VioC structures. (a) Ribbons representation and (b) close-up view of the active site from the VioC:Fe:2OG:Arg complex structure reported in this study (PDB: 6 Y 0 N ). The N - and C-termini are labelled as N and C , respectively. Alpha-helices and beta-strands are shown in gold and cyan, respectively. The arginine substrate, 2OG cosubstrate and coordinating residues are shown in sticks. Panel (b) shows the composite omit map to $1.86 \AA$ resolution displayed at $0.7 \sigma$ contour level, carved around the arginine substrate and 2OG. (c) Ribbons representation and (d) close-up view of the active site of VioC:Fe:succinate:(3S)-hydroxy-1Arg complex after dioxygen exposure (PDB: 6Y12). Panel (d) shows the composite omit map to 1.70 $\AA$ resolution displayed at $0.7 \sigma$ contour level, carved around the ( $3 S$ )-hydoxy-l-Arg and succinate. (e) Superimposition of the cryogenic (magenta, and salmon, PDB: 6ALM) and serial room temperature (cyan, and yellow, PDB: 6Y0N, this study) VioC:Fe:2OG:Arg complex structures reveals small apparent differences in the bidentate coordination of the metal by 20G, as well as a change in the conformation of the arginine substrate (including the $\alpha$-amino group), possibly reflecting the flexible nature of the VioC active site (Mitchell et al., 2017). (f) Superimposition of the cryogenic (magenta
and salmon, PDB: 2WBP) and serial room temperature (cyan and yellow) VioC:Fe:succinate:(3S)-hydroxy-l-Arg complex (PDB: 6Y12, this study) structures reveals small apparent differences in the conformation of (3S)-hydroxy-l-Arg, whereas the binding of succinate is consistent with that in the reported cryogenic structure (Helmetag et al., 2009; Mitchell et al., 2017).

