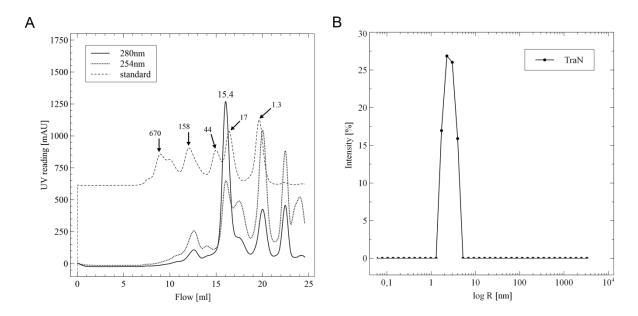
## Acta Crystallographica Section D

Volume 70 (2014)

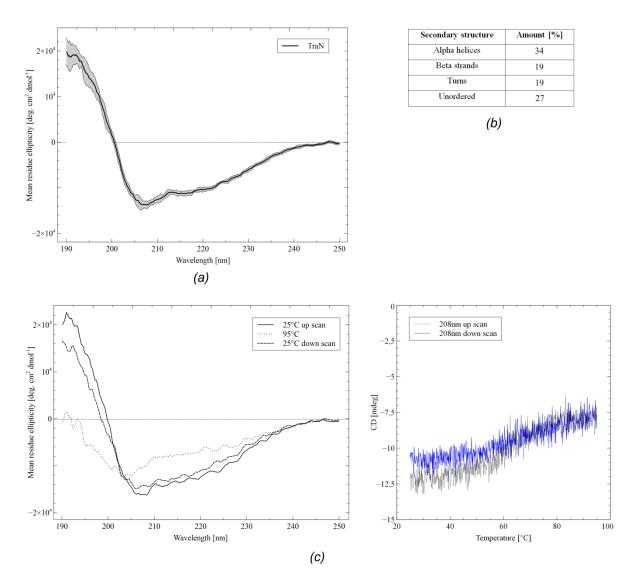
Supporting information for article:

Structure of the double-stranded DNA-binding type IV secretion protein TraN from *Enterococcus* 

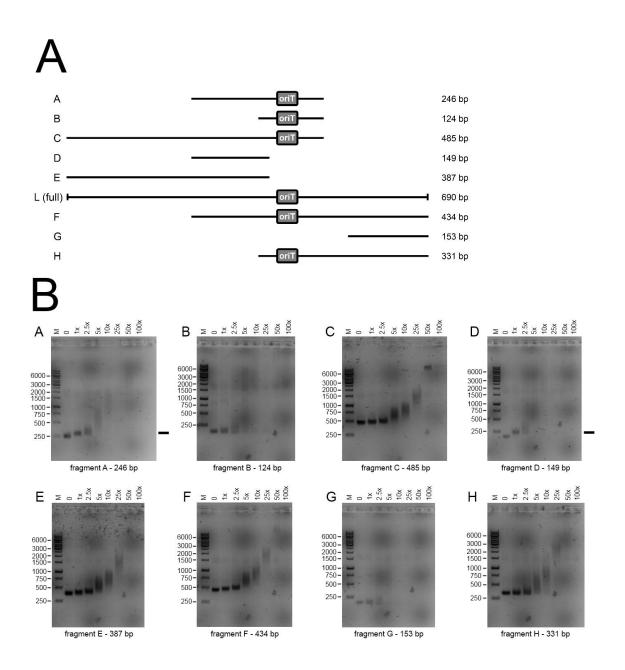
Nikolaus Gössweiner-Mohr, Markus Eder, Gerhard Hofer, Christian Fercher, Karsten Arends, Ruth Birner-Gruenberger, Elisabeth Grohmann and Walter Keller



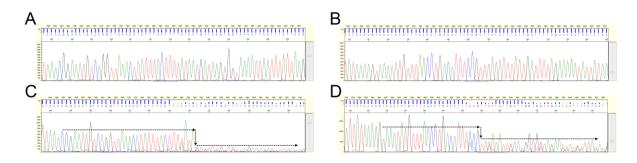
**Figure S1** Biophysical characterization of TraN. **A:** TraN elutes as a single peak from the Superdex 200 size exclusion column; 280 nm (black) and 254 nm (dotted) readings are shown; a standard is shown with its molecular weight (discontinued). **B:** In the mono-dispersity analysis via DLS, TraN appears as a narrow peak.



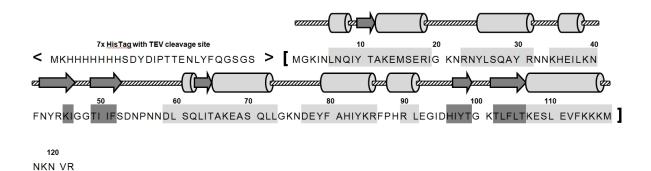
**Figure S2** CD analysis of TraN. **A:** TraN is folded in solution; the black curve represents the average of 10 individual wavelength scans; the standard deviation is displayed as a grey area. **B:** Secondary structure content of TraN; the NRMSD is 0.018. **C:** TraN unfolding and refolding characteristics. The CD spectra are shown at 25 and 95 °C and after cooling to 25 °C (left panel). The temperature scan (up- and down-scan) is shown in the right panel.



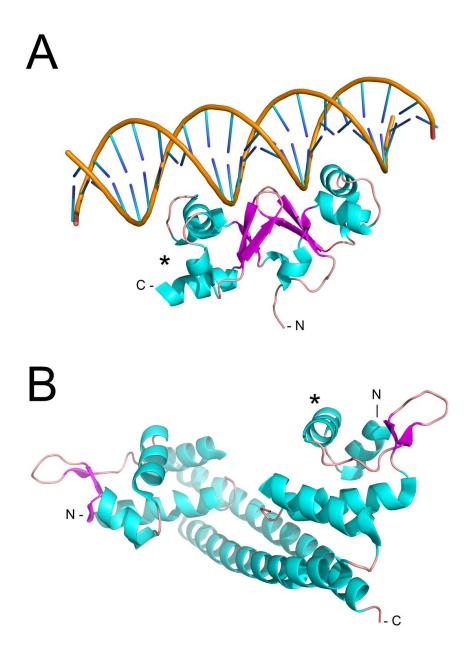
**Figure S3** The putative TraN binding region. **A:** The *oriT* upstream/downstream fragments used in EMSA experiments. The pIP501 *oriT* sequence is indicated; the length of the fragments is given. **B:** TraN specifically shifts fragments A and D already at an equimolar protein :: DNA ratio; all fragments are co-operatively shifted at higher TraN concentrations; lane M: molecular weight standard (GeneRuler 1 kb DNA Ladder, ThermoFisher Scientific, Waltham, Massachusetts, USA).



**Figure S4** Exemplary data set for identification of the TraN preferred binding site. Sequencing chromatograms of fragment L forward strand (A, C) and reverse strand (B, D) with no (A, B), respectively 15 minutes (C, D) T4 exonuclease digestion. Dashed lines were added to highlight the decrease of the signal intesity. The sequencing data were analyzed with free Sequence Scanner v1.0 software from Applied Biosystems.



**Figure S5** The TraN aa sequence and the TraN construct. Secondary structure features predicted by PsiPred are marked in light grey (helices) and dark grey (beta sheets); the 7xHis-tag was lost during crystallization; 5 residues on the C-terminus are not visible in the crystal structure; square brackets highlight the sequence present in the X-ray structure; secondary structure elements found in the X-ray model are shown as light grey tubes (helices) and dark grey arrows (beta sheets).



**Figure S6** Proposed binding mode of TraN to dsDNA and comparison to a MerR dimer. **A:** Proposed binding mode of TraN to random ds B-DNA. Both molecules are shown in cartoon representation with secondary structure elements highlighted (helices in cyan, strands in purple). The 3D model of the DNA was generated with the DNA structure modelling server 3D-DART (van Dijk and Bonvin, 2009) and docked manually to the TraN model in PyMOL. **B:** Cartoon representation of the MerR dimer structure (PDB: 3GPV) with secondary structure elements highlighted (helices in cyan, strands in purple). The two MerR molecules form the quaternary structure via a two-helix coiled-coil motif. The two DNA interaction motifs (the TraN-like half sites) are placed on the opposite sides of the dimer structure with the primary DNA interaction helices in parallel, as in the TraN structure. An asterisk (\*) marks the two structurally corresponding HTH domains of TraN (**A**) and MerR (**B**), respectively.

Туре	Gram	Species	System	Protein	10	20	30	40	D	50	60	70	80	90	100	110	120	130
α	+	Enterococcus faecalis	pIP501	TraN														
	+			crystal														
	+	Enterococcus faecalis	pRE25	Orf38														
	+	Enterococcus faecalis	pAM-beta1	OrfB														
	+	Enterococcus faecium	pVEF3	P52														
	+	Enterococcus italicus	DSM15952 (genomic)	HMPREF9088_2233						ľ					1			
	+	Enterococcus faecalis	7L76 (genomic)	ENT03500														

Figure S7 Results of the secondary structure based search for TraN-like proteins. Secondary structure (Psipred) and trans-membrane motif (HMMTOP, TMPRED) prediction for G+ TraN-like proteins from conjugative plasmids, transposons, ICEs and GIs; alpha helices (blue) and beta strands (red) are highlighted.

Appendix A. Supplemental Table

Table S1	Validating the structural similarity of TraN to its	self and related structures.

A

TraN - CTD	PDB code	Residues aligned	Sequence identity [%]	Secondary structure similarity [%]	Superfamily reliability [%]	Fold reliability [%]	
TraN N-terminal domain		4 - 56	28.8	80.8	97.0	99.1	
Excisionase (Xis) Tn916 - Enterococcus faecalis	1Y6U	9 - 64	12.5	87.5	89.3	98.5	
Excisionase Klebsiella pneumoniae	2KVV	5 - 70	10.3	91.4	63.3	93.5	
Excisionase (Xis)	1RH6	1 - 50	12.5	75.0	84.3	98.0	
bacteriophage Lambda Excisionase (Xis)	1PM6	1 - 53	11.8	70.6	58.8	91.5	
bacteriophage HK022 Transcriptional regulator	3GPV	20 - 72	5.7	81.1	35.3	68.4	
MerR family - Bacillus thuringiensis							

## B

TraN - CTD	PDB code	Number of aligned residues	Number of gaps	Sequence identity [%]	Core RMSD achieved [Å]	
TraN		51	3	31.4	1.78	
N-terminal domain			U U	0111	1110	
Excisionase (Xis)	1Y6U	53	3	13.2	2.78	
Tn916 - Enterococcus faecalis	1100	55	U U		0	
Excisionase	2KVV	53	2	7.6	2.84	
Klebsiella pneumoniae	211					
Excisionase (Xis)	1RH6	44	2	11.4	1.75	
bacteriophage Lambda			_			
Excisionase (Xis)	1PM6	46	3	13.0	1.98	
bacteriophage HK022						
Transcriptional regulator	3GPV	50	3	8.0	2.42	
MerR family - Bacillus thuringiensis						

CTD - C-terminal domain; Alignments were performed with MATRAS (A) and SUPERPOSE in Coot (B)