



1 **The cooperative application of oyster shell and biochar efficiently enhanced**
2 **in-situ remediation of cadmium contaminated soil around intensive industry**

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27

Abstract

28 Biochar has been widely used for the in-situ remediation in the cadmium (Cd)
29 contaminated soil, while the high-cost of biochar limited its application in farmland.
30 In this study, we firstly investigated the possibility of cooperative application of
31 oyster shell and biochar to enhance Cd immobilization efficiency and reduce the cost
32 in field experiments under rice-oilseed rape rotation. Treatments were comprised of:
33 rice planting without amendments (R-PA0); followed with 15000 kg/ha biochar
34 (R-PA1); followed with 15000 kg/ha oyster shell (R-PA2); followed with 7500 kg/ha
35 biochar and 7500 kg/ha oyster shell (R-PA3); rice-oilseed rape rotation without
36 amendments (RT-PA0); rotation with 15000 kg/ha biochar (RT-PA1); rotation with
37 15000 kg/ha oyster shell (RT-PA2); rotation with 7500 kg/ha biochar and 7500 kg/ha
38 oyster shell (RT-PA3). Results revealed that HOAc-extractable Cd was significantly
39 decreased by 38.46% in R-PA2. Cd contents in brown rice and oilseed were reduced
40 by 29.67% in R-PA3 and 19.74% in RT-PA3 compared with control. Meanwhile,
41 Hazard Quotient of brown rice and oilseed significantly decreased in RT-PA3. The
42 Olsen-P in R-PA3 and RT-PA3 was markedly increased by 187.46% and 184.73%,
43 respectively. In addition, activities of urease, catalase, and β -galactosidase in RT-PA3
44 were significantly increased by 268.88%, 30.44% and 245.28%, respectively.
45 Furthermore, the joint application of biochar with oyster shell significantly decreased
46 the cost of soil remediation at least 9600 RMB/ha. These results demonstrated that the
47 joint utilization of biochar with oyster shell might be an economical and effective
48 pathway to achieve in-situ remediation of Cd contaminated farmland.



49 **Keywords:** Biochar; Oyster shell; Rice-oilseed rape rotation; In-situ remediation;
50 Enzyme activities; Cadmium

51 **1. Introduction**

52 Heavy metals have been considered as hazardous materials for human health,
53 and among which cadmium (Cd) is one of the most toxic heavy metals (Yang et al.,
54 2021). The excessive intake of Cd could cause serious damages to bones, thyroid, and
55 kidneys (Ma et al., 2021a). According to the latest national survey on the status of soil
56 environmental quality in China, Cd has ranked as the highest contaminants (7%)
57 among all heavy metals (Mou et al., 2020). In southwest China, a large amount of
58 farmlands were contaminated with Cd owing to the intensive industrialization (Chen
59 et al., 2018a). In addition, soil acidification also aggravated the bioavailability and
60 solubility of Cd, thus enhancing Cd uptake by crops (Feng et al., 2020). Therefore, the
61 development of cost-effective and eco-friendly remediation technologies is crucial for
62 food safety and soil quality.

63 In recent years, in-situ immobilization as an effective technology has raised wide
64 attentions in the remediation of Cd contaminated farmlands, which can reduce the Cd
65 uptake by plants without delaying agricultural production (Palansooriya et al., 2020;
66 Wang et al., 2021). In recent years, biochar derived from bio-wastes is widely
67 recommended as a soil amendment (Zong et al., 2021). Amounts of nutrients (such as
68 C, N, P, K, and Mg etc.) in biochar could improve soil fertility and promote plant
69 growth (Lu et al., 2015). Moreover, biochar has a large surface area and plenty of
70 functional groups, which are reactive to immobilize heavy metals, including Cd, Pb,



71 and Ni (Wang et al., 2021). However, the high price of biochar limited its large
72 application. In addition, the soil pH regulation by biochar in acid fields was not very
73 significant, while the soil pH values were negatively related to Cd availability (Liu et
74 al., 2018). Therefore, it is vital to decrease the remediation cost of biochar without
75 reducing immobilization efficiency of Cd. Oyster shell is a low-cost and largely
76 available bio-waste product from oyster farming (Li et al., 2020). Previous studies
77 found that oyster shell as a promising slow-release alkaloid has outstanding effects on
78 pH adjustment and Cd immobilization in soils (Chen et al., 2018b; Peng et al., 2020).
79 In this sense, we think that the joint use of biochar and oyster shell might be a
80 low-cost and effective pathway to decrease Cd uptake by crops and improve soil
81 biochemical quality in acidic fields. However, there was little known about the joint
82 effects of biochar and oyster shell on the in-situ remediation in Cd contaminated soil.

83 Rice and oilseed rape were the main food and economic crops in southwest
84 China, and rice-oilseed rape rotation is the dominant production model (Liu et al.,
85 2014). Previous studies mainly focused on the effects of amendments on reducing the
86 Cd uptake by rice, while the remediation efficiency of passivators under the rice-rape
87 rotation was little known. Based on above opinions, a field experiment under
88 rice-oilseed rape rotation was designed: (1) to investigate the cooperative effects of
89 biochar and oyster shell on Cd immobilization; (2) to evaluate the effects of biochar
90 and oyster shell on decreasing human health risk of Cd; (3) to reveal the effects of
91 biochar and oyster shell on soil biochemical properties including pH, CEC, total
92 organic carbon, organic matter, Olsen-P, Olsen-K, Alkeline-N, and the activities of



93 soil enzymes, so as to estimate the pearson correlation analysis (PCA) model of main
94 parameters in the moderately polluted farmland.

95 **2. Material and methods**

96 **2.1. Experimental site and soil properties**

97 A field trial was conducted during 2019-2020 in a rice-oilseed rape rotation
98 cultivated site where the soil was moderately contaminated by Cd. The field site was
99 located in a dominant agricultural cultivation region round industrial parks in
100 Chengdu plain, Sichuan province, China (104°18'N, 31°81'E). This region belongs to
101 a subtropical monsoon humid climate with an average temperature of 16.1 °C and
102 annual rainfall of about 1000 mm. The main properties of the topsoil (0 - 20 cm)
103 collected from the site in 2019 and 2020 were shown in Table S1.

104 **2.2. Characteristics of experimental materials**

105 Biochar was purchased from Zhenjiang Zedi agricultural and biological Co., Ltd.,
106 which was produced from rice straw in a reactor with N₂ and 500 °C for about 4 h.
107 Oyster shell was purchased from Fujian Mata Co., Ltd (< 0.3 mm mesh). The surface
108 structures of biochar and oyster shell were analyzed by Scanning Electron Microscope
109 (SEM, JSM-7500F). The functional groups of biochar and oyster shell were measured
110 by Fourier Transform Infrared Spectra (Nicolet 6700). The seeds of rice “Yixiang
111 2115” and seeds of oilseed rape “Yiyou 15” were obtained from Rice Research
112 Institute, Sichuan Academy of Agricultural Science.

113 **2.3. Experimental setup**

114 The field experiment was conducted during 2019 - 2020 as following treatments:



- 115 ➤ R-PA0: Rice planting without passivators;
- 116 ➤ R-PA1: Rice planting with 15 t/ha biochar;
- 117 ➤ R-PA2: Rice planting with 15 t/ha oyster shell;
- 118 ➤ R-PA3: Rice planting with 7.5 t/ha biochar and 7.5 t/ha oyster shell;
- 119 ➤ RT-PA0: Rice-oilseed rape rotation without passivators;
- 120 ➤ RT-PA1: Rice-oilseed rape rotation with 15 t/ha biochar;
- 121 ➤ RT-PA2: Rice-oilseed rape rotation with 15 t/h oyster shell;
- 122 ➤ RT-PA3: Rice-oilseed rape rotation with 7.5 t/ha biochar and 7.5 t/ha oyster
- 123 shell.

124 The concentrations of biochar and oyster shell used in this study were referred to
125 previous reports (Ameloot et al., 2014). Each experimental plot was 56 m² (7 x 8 m)
126 and arranged in a randomized design with three replicates. Before rice planting, the
127 passivators were sufficiently mixed with topsoil. After the harvest of rice, the oilseed
128 rape was planted following the conventional tillage pattern without any passivator.

129 **2.4. Plant analysis**

130 The rice grain and rapeseed were dried and ground to powder. Then, 0.2 g
131 samples were digested with HNO₃:HClO₄:HF in a mixture of 5:4:3 (v/v) and the
132 mixture was then diluted to 10 mL with 1% HNO₃ (Wu et al., 2019b). The Cd
133 concentrations in the mixture were determined by AAS.

134 **2.5. Soil analysis**

135 Soil pH was determined by a pH meter (METTLER-S220) with a soil/water ratio
136 of 5 g/25 mL. The bioavailable Cd of soil was measured by the TCLP method (Xu et



137 al., 2020). Briefly, 2 g of soil sample was mixed with 40 mL 0.11 M acetic acid and
138 shaken at 25 °C, 150 rpm for 16 h. Then, the mixture was centrifuged for 5 min at
139 8000 rpm and the supernatant was collected to determine Cd content by atomic
140 absorption spectroscopy (AAS; VARIAN, SpecterAA-220Fs). Olsen-P, Olsen-K, and
141 Alkeline-N were measured according to the method ascribed by (Liu et al., 2017).
142 Soil TOC and OM were determined by the method ascribed by (Walz et al., 2017).

143 In addition, activities of soil enzyme were analyzed to reflect the biological
144 quality in this study. Dehydrogenase activity was evaluated by the production of
145 triphenylformazan (TPF) at OD_{492nm} and expressed as µg TPF/g soil/24 h (Benefield et
146 al., 1977). Acid phosphate activity was assayed by the *p*-nitrophenol (pNP) release at
147 OD_{400nm} and expressed as µg pNP/g soil/24 h (van Aarle and Plassard, 2010). Urease
148 activity was determined by the NH₄-complex at OD_{578nm} and expressed as µg
149 NH₄-N/g soil/24 h (Yan et al., 2013). Catalase activity was measured by back titration
150 of H₂O₂ added to soil with 0.1 M KMnO₄ and expressed as by mL 0.1 M KMnO₄/g
151 soil/h (Zhang et al., 2011). Invertase activity was assayed by the amount of glucose
152 production at OD_{508nm} and expressed as µg glucose/g soil/24 h (Wu et al., 2019b).
153 β-galactosidase activity was measured by the released 4-methylumbelliferone (MUF)
154 and expressed as µg MUF µmol/g soil/h (Martínez-Iñigo et al., 2009).

155 **2.6. Human risk assessment of Cd**

156 The health risks of Cd to adults and children in the crops were separately
157 assessed by the Hazard Quotient (HQ) according to the method introduced by
158 Environmental Protection Agency (EPA) in the US (Wei et al., 2020). HQ values were



159 calculated as the following formula:

$$160 \quad HQ = (EF \times ED \times C \times IR) / (BW \times AT \times RfD) \quad (1)$$

161 *EF* (Exposure Frequency): 365 days/year.

162 *ED* (Exposure Duration): 70 years for adult, 7 years for children.

163 *C*: Cd concentrations in the rice grain and oilseed (mg/kg).

164 *IR* (Ingestion Rate): For rice grain, 0.3892 kg/day for adult and 0.1984 kg/day for
165 children, respectively. For rape oil, 0.025 kg/day for adult and 0.0125 kg/days for
166 children, respectively.

167 *BW* (Body Weight): 62.71 kg for adult male, 55.1 kg for adult female and 25.6 kg
168 for children.

169 *AT* (Averaging Time): 25550 days for adult and 2555 days for children.

170 *RfD* (Reference of Dose): 0.001 mg/kg for Cd.

171 **2.7. Statistical analysis**

172 In this study, statistical significance was analyzed using SPSS 18.0 package, and
173 means values were considered to be different when $P < 0.05$ using least significant
174 difference (LSD). All statistics were performed using Origin 8.0 (USA).

175 **3. Results and discussion**

176 **3.1. Characteristics of soil and amendments**

177 The main characteristics of soil, biochar and oyster shell were shown in Table S1.
178 The soil in the field was acidic soil with pH values of 5.27 - 5.51. The biochar and
179 oyster shell used in the field study were alkaline materials and their pH values were
180 8.22 and 8.52, respectively. The OM of biochar (541.53 mg/kg) was significantly



181 higher than that of soil (39.32 mg/kg) and oyster shell (12.60 mg/kg). The carbon
182 percentage of biochar also reached 92.50%.

183 The surface of oyster shell (Figure 1a) was a regular filamentary layer with some
184 disordered deposition, which might be calcium compounds. The structure of biochar
185 (Figure 1b) was lamellar and polyporous, which was in favor of Cd absorption. In
186 addition, FTIR was operated to detect functional groups of oyster shell and biochar
187 (Figure 1c). The characteristic peaks of calcium carbonate in oyster shell were
188 observed at 1427 cm^{-1} and 879 cm^{-1} (Lu et al., 2021). Biochar showed obvious peaks
189 at 1089 cm^{-1} and 790 cm^{-1} , which were related to C-O, and C-H bending vibration,
190 respectively (Wu et al., 2019a). In addition, an obvious feature at 3436 cm^{-1}
191 corresponding to -OH was loaded on oyster shell and biochar (Lian et al., 2021).

192 **3.2. Analysis of soil Cd bioavailability**

193 To evaluate the effect of different amendments on Cd bioavailability, the
194 concentrations of AcOH-extractable Cd in soils were determined by TCLP method
195 (Halim, 2003). Figure 2 showed the variations of AcOH-extractable Cd with different
196 amendments in rice-oilseed rape rotation. Compared to rice planting, the
197 concentrations of AcOH-extractable Cd increased in the oilseed rape planting, which
198 was mainly related to the different irrigation methods. In this study, rice planting was
199 performed by flooding irrigation during the whole physiological period whereas
200 dry-land cultivation was used in oilseed rape planting. In general, flooding irrigation
201 is beneficial to reduce Cd bioavailability due to the precipitation of Cd compounds
202 under low-redox status ($E_h < 0\text{ mV}$) (Mou et al., 2020). In addition, treatments with



203 biochar and oyster shell resulted in the reduction of AcOH-extractable Cd in soils.
204 Compared to R-PA0, the AcOH-extractable Cd was significantly decreased by
205 20.79% and 40.59% in R-PA1 and R-PA2, respectively. Compared to RT-PA0, the
206 AcOH-extractable Cd was also reduced by 5.76% and 17.85% in RT-PA1 and RT-PA2,
207 respectively. Moreover, the Cd immobilization efficiency in R-PA3 and RT-PA3 was
208 higher than that in R-PA1 and RT-PA1. These results demonstrated that oyster shell
209 gave a better Cd immobilization than biochar and the addition of oyster shell could
210 strength the Cd immobilization capacity of biochar.

211 **3.3. Analysis of Cd contents in brown rice and oilseed**

212 As shown in Figure 3, the application of biochar and oyster shell significantly
213 reduced Cd contents in brown rice and oilseed. Treatments without amendments, the
214 contents of Cd in brown rice reached 0.88 mg/kg. Compared to control (R-PA0), the
215 Cd contents in brown rice was decreased by 20.88% in R-PA1 and 30.77% in R-PA2,
216 indicating that oyster shell has the superior of Cd immobilization capacity than
217 biochar. Obviously, the cooperative addition of biochar and oyster shell (R-PA3)
218 contributed to higher Cd reduction (29.67%) in brown rice than that in sole biochar
219 (R-PA1, 20.88%). In addition, Cd contents in oilseed were significantly reduced in the
220 RT-PA1 and RT-PA3, about 27.63% and 19.74% lower than that in RT-PA0,
221 respectively. The results indicated that biochar and oyster shell application could
222 efficiently decrease Cd accumulation in brown rice and oilseed.

223 **3.4. Health risk assessment of cadmium**

224 HQ values of Cd for brown rice and oilseed intake in different treatments were



225 presented in Figure 4. The order of HQ for consuming rice and oilseed was children >
226 adult female > adult male, indicating that children were more sensitive than adults
227 under Cd exposure. Without amendments (R-PA0), HQ values of consuming brown
228 rice for adult male, adult female and children reached 5.46, 6.21 and 6.82,
229 respectively. After the application of amendments, HQ values of brown rice intake
230 were significantly decreased by 20.87%, 31.11% and 29.76% in R-PA1, R-PA2 and
231 R-PA3, respectively. Although HQ values of oilseed were significantly lower than that
232 of rice grain, but the values also decreased by 17.27 - 28.14% in the oyster shell and
233 biochar treatments.

234 **3.5. Analysis of soil biochemical properties**

235 **3.5.1. Analysis of soil pH and CEC**

236 It was observed that soil pH was weakly increased by biochar, but significantly
237 increased by oyster shell (Figure 2a). After the oyster shell application, the soil pH
238 increased to neutral (6.9 - 7.3) from acidity (5.2 - 5.5). Compared with control
239 (R-PA0), the soil pH was increased by 1.8, 1.6, 1.4 and 1.7 points in R-PA2, R-PA3,
240 RT-PA2 and RT-PA3, respectively. The application of oyster shell slight increased the
241 CEC in the rice planting, while oyster shell and biochar had no significant effects on
242 CEC in the rice-oilseed rotation (Figure 2b).

243 **3.5.2. Analysis of soil nutrients**

244 It is important for in-situ remediation of Cd contaminated soil by bio-wastes
245 without inhibiting the soil available nutrients. To analyze the effects of amendments
246 on soil bioavailable nutrients, the contents of TOC, OM, Olsen-P, Olsen-K, and



247 Alkeline-N were determined during the rice-oilseed rape rotation (Table S2). Biochar
248 application slightly increased TOC and OM in rice planting and rice-oilseed rotation.
249 In rice planting, TOC and OM in R-PA3 were increased by 10.09% and 9.92%
250 compared with R-PA0, respectively. In rice-oilseed rape rotation, soil TOC in RT-PA1
251 was enhanced by 11.06% and 11.32% compared with RT-PA0, respectively. More
252 obviously, Olsen-P was significantly increased by the addition of oyster shell.
253 Compared with R-PA0, the Olsen-P in R-PA2 and R-PA3 significantly increased by
254 200.96% and 187.46%, respectively. Compared with RT-PA0, the Olsen-P in RT-PA2
255 and RT-PA3 significantly increased by 295.92% and 184.73%, respectively.

256 **3.5.3. Analysis of soil enzyme activities**

257 As shown in Figure 6, adding amendments variously changed the activities of
258 soil enzyme. In the rice planting, biochar application increased the dehydrogenase
259 activity, about 20.12% (R-PA1) and 25.49% (RT-PA1) higher than that of control
260 (R-PA0). However, oyster shell significantly increased the dehydrogenase activity in
261 rice-oilseed rotation, which was markedly increased by 59.75% and 53.39% in the
262 RT-PA2 and RT-PA3 compared with control, respectively. Urease activity was no
263 obvious variation in the biochar treatment whereas markedly enhanced in the oyster
264 shell treatment. Compared with RT-PA0, urease activity was significantly increased
265 by 268.88% in RT-PA3. However, oyster shell and biochar had no obvious impacts on
266 acid phosphate activity, except for a reduction of 43.30% in R-PA2. In addition, the
267 application of biochar has no negative effects on invertase activity, while oyster shell
268 slightly decreased the invertase activity on rice-oilseed rape rotation. In the RT-PA3



269 treatment, catalase activity was significantly increased by the application of biochar
270 and oyster shell. Moreover, β -galactosidase activity was significantly increased by
271 245.28% in RT-PA0 with the maximum of 12.29 $\mu\text{g MUF } \mu\text{mol/g soil/h}$.

272 **3.6. Analysis of correlation coefficient**

273 To analyze and confirm the relationship among different parameters, the Pearson
274 correlation analysis was used to experimental data. As shown in Figure 7a, Cd content
275 in brown rice was positively correlated to Cd bioavailability ($R^2 = 0.90$) but
276 negatively correlated to soil pH ($R^2 = -0.83$). Meanwhile, the activities of soil
277 enzymes except acid phosphate were positively connected to alkaline-N, Olsen-P,
278 Olsen-K, and TOC. Figure 7b showed a weak correlation between Cd uptake of rape
279 and Cd bioavailability. In addition, soil pH was positively correlated to Olsen-P and
280 β -galactosidase activity ($R^2 > 0.95$), which further demonstrated that alkaline
281 substances could increase Olsen-P content and β -galactosidase activity by adjusting
282 soil pH in acid fields.

283 **3.7. Cost approach for amendments**

284 Considering the large scale remediation of the contaminated agricultural soil, the
285 cost of amendments is a key parameter in the practical application. The market price
286 of biochar (> 1200 RMB/t) was much higher than that of oyster shell (500 RMB/t)
287 (detailed information see Supplementary Information). In this study, the dosage of
288 amendments was 12000 kg/ha. The market price for biochar amendment was at least
289 14400 RMB/ha, while the joint use of biochar and oyster shell significantly decreased
290 the cost of amendments at least 9600 RMB/h. Furthermore, the joint application of



291 biochar and oyster shell is more effective to improve soil biochemical properties
292 compared to biochar. Based on these consideration, the collaborative passivation of
293 biochar and oyster shell might be a economical pathway for the safe-use of Cd
294 contaminated soil.

295 **4. Discussion**

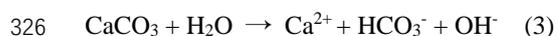
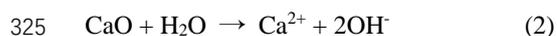
296 Rice and oilseed rape were the most important crops over the globe.
297 Simultaneously, rice and oilseed rape rotation was the main cultivated model in China.
298 However, the Cd contamination in agricultural lands, especially in acidic soils, has
299 severely threatened food safety production and human health. Cd accumulation in the
300 plants poses a great human health risk. Cd uptake by crops may result in kidney
301 damage and adverse effects on lung, cardiovascular, musculoskeletal systems (Wei et
302 al., 2020).

303 In-situ immobilization was an effective pathway to decrease Cd uptake by crops
304 by the application of amendments. Biochar is originated from bio-wastes, such as
305 straw, coconut shell and animal manure. Previous studies has revealed that biochar
306 has a great potential on Cd immobilization by surface absorption and co-precipitation.
307 However, the high price of biochar and weak Cd binding in acidic soil was limited its
308 large application in agricultural lands. In this study, our results showed that the
309 cooperative application of oyster shell and biochar could contribute to the reduction of
310 AcOH-extractable Cd in soils (Figure 2) and Cd uptake by crops (Figure 3).
311 Furthermore, the non-cancer risk description methodology of HQ was widely applied
312 to assess the possibility of health risk of Cd in different plants (Ma et al., 2021b). The



313 decreased HQ values demonstrated the human health risk of Cd decreased by the
314 application of oyster shell and biochar.

315 AcOH-extractable Cd has widely used to evaluate Cd bioavailability in soils, and
316 the reduction of Cd bioavailability was main mechanism of in-situ immobilization
317 (Liu et al., 2021). Soil pH was the main factor influencing the Cd bioavailability in
318 soils. It has been widely verified that soil pH determined solid-solution equilibria of
319 heavy metals in soils (Zhao and Masaihiko, 2007). Comparatively, soil pH in the
320 oyster shell treatment was higher than in biochar treatment (Figure 5). Although the
321 pH values of oyster shell (8.52) and biochar (8.22) were similar, oyster shell was
322 regarded as a low-release alkaloid in soils due to it is composed of CaO and CaCO₃.
323 The dissolution of CaO and CaCO₃ from oyster shell in water could produce hydroxyl
324 ion (OH⁻) as the following chemical reactions (Ok et al., 2010):



327 An increase in soil pH can cause an increase in the negative soil surface charge, which
328 easily causes an increased capacity of cationic metal adsorption (Ok et al., 2010). The
329 precipitants of metal oxy/hydroxides could be formed due to increased hydroxyl ions
330 (Bolan et al., 2014). In addition, previous studies also found that functional groups
331 such as -OH and C-O loaded on the surface of oyster shell and biochar can decrease
332 the Cd solubility by surface adsorption and precipitation (Ok et al., 2010; Tang et al.,
333 2020). Therefore, the cation exchange, surface complexation and co-precipitation
334 might be mechanisms for the Cd immobilization of biochar and oyster shell in acidic



335 filed.

336 Olsen-P, Olsen-K and Alkeline-N play an important role on soil biochemical
337 quality and plant growth. P fractions are mainly dependent on soil pH, soil mineralogy
338 and phosphate fertilizer application (Lee et al., 2008). Fe-P and Al-P are the
339 predominant forms in acidic soils while calcium bound-P is the predominant form in
340 alkaline soils (Dean, 1949). In acidic soils, the loosely bound phosphates are
341 converted into Fe-P and Al-P fractions gradually owing to the re-precipitation process.
342 Previous studies found that Olsen-P content reaches the maximum at neutral pH soils
343 (Lee et al., 2008). Our results showed that the addition of oyster shell markedly
344 increased the content of Olsen-P in soils, which might be resulted from the
345 enhancement of soil pH (Table S2). PCA analysis (Figure 7) further demonstrated that
346 Olsen-P was highly correlated with changes of soil pH ($R^2 > 0.99$). The contents of
347 Olsen-K and Alkeline-N also slightly increased with the application of biochar and
348 oyster shell, indicating an improvement of soil fertility.

349 Activities of soil enzyme have been widely used to reflect soil biological quality
350 (Lin et al., 2021). In this study, activities of dehydrogenase, urease, acid phosphate,
351 invertase, catalase and β -galactosidase were determined, and most of which were
352 increased by the application of biochar and oyster shell (Figure 6). Especially, the
353 increase of activities of dehydrogenase, urease, catalase and β -galactosidase was
354 obvious under the stimulation of biochar and oyster shell. The increase of soil enzyme
355 activities might be explained from the following aspects. The addition of oyster shell
356 increased the soil pH, which usually caused the enhancement of dehydrogenase and



357 urease activities (Wen et al., 2021). Abd El-Azeem et al reported that dehydrogenase
358 activity was positively correlated with soil pH (Abd El-Azeem et al., 2013). Oyster
359 shell could raise the urease activity, thus catalyzing the hydrolysis of urea to CO₂ and
360 NH₃ with an optimum pH around 7.4 (Lee et al., 2008). In addition, the porous
361 structure and rich nutrients of biochar and oyster shell can contribute to the growth of
362 soil microorganisms, and thus might increase the soil enzyme activities (Azadi and
363 Raiesi, 2021; Wu et al., 2019a). Moreover, the enhancement of enzyme activities in
364 biochar and oyster shell treatments could also be related to the decrease of Cd toxicity
365 in soils (Zhang et al., 2021). In conclusion, the cooperative application of biochar and
366 oyster shell was the most effective pathway in the in-situ remediation of Cd
367 contaminated farmlands.

368 **5. Conclusions**

369 The current study revealed the effects of oyster shell and biochar on Cd
370 bioavailability, Cd uptake by crops, and human health risk of Cd as well as soil
371 biochemical properties during rice-oilseed rape rotation. The application of oyster
372 shell showed an extraordinary potential to increase soil pH for a duration, which
373 significantly decreased the Cd bioavailability in soils. The cooperative application
374 significantly reduced Cd contents and human health risk of brown rice and oilseed. In
375 addition, biochar application increased OM and TOC, while the addition of oyster
376 shell was suitable to improve Olsen-P, Olsen-K, and Alkeline-N. Furthermore, the
377 activities of soil enzyme were markedly enhanced by the cooperative application of
378 oyster shell and biochar. In addition, Our results suggested that the joint application of



379 biochar and cheap oyster shell was a low-cost pathway to effectively reduce Cd
380 uptake of crops and improve soil biochemical properties.

381 **Acknowledgements**

382 This study was financially supported by Key Technologies Research and
383 Development Program (CN) (2018YFC1802605), the Science and Technology Project
384 of Sichuan Province (2022ZDYF0281) and Chengdu Science and Technology Project
385 (2021-YF05-00195-SN). The authors also wish to thank Professor Guanglei Cheng
386 and Hui Wang from Sichuan University for the technical assistance.

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523 **Figure captions:**

524 **Figure 1** SEM images of oyster shell (a) and biochar (b) and FTIR spectra (c) of
525 oyster shell and biochar.

526 **Figure 2** The effects of passivators on Cd bioavailability in soil. Dots represent the
527 value of each sample. Bars followed with different lowercase letters (a - c) and capital
528 letters (A, B) indicated significant ($p \leq 0.05$) difference among different treatments in
529 rice planting and oilseed rape planting according to the LSD test. Values represent
530 means \pm standard deviation.

531 **Figure 3** The effects of passivators on Cd contents in brown rice (a) and oilseed (b).
532 Dots represent the value of each sample. Bars with different lowercase letters
533 indicated significant ($p < 0.05$) difference among different treatments according to the
534 LSD test. Values represent means \pm standard deviation.

535 **Figure 4** The effects of different passivators on the HQ of grown rice and oilseed.
536 Mean with different lowercase letter indicated significant ($p < 0.05$) difference from
537 each other according to the LSD test. Values represent means \pm standard deviation.

538 **Figure 5** The effects of different passivators on soil pH (a) and CEC (b). Dots
539 represent the value of each sample. Bars followed with different lowercase letters (a -
540 c) and capital letters (A, B) indicated significant ($p \leq 0.05$) difference among different
541 treatments in rice planting and oilseed rape planting according to the LSD test. Values
542 represent means \pm standard deviation.

543 **Figure 6** The effects of different passivators on the activities of soil enzyme. Dots
544 represent the value of each sample. Bars followed with different lowercase letters (a -
545 c) and capital letters (A-C) indicated significant ($p \leq 0.05$) difference among different
546 treatments in rice planting and oilseed rape planting according to the LSD test. Values
547 represent means \pm standard deviation.

548 **Figure 7** The correlation of investigated parameters in rice planting (a) and
549 rice-oilseed rape rotation (b)

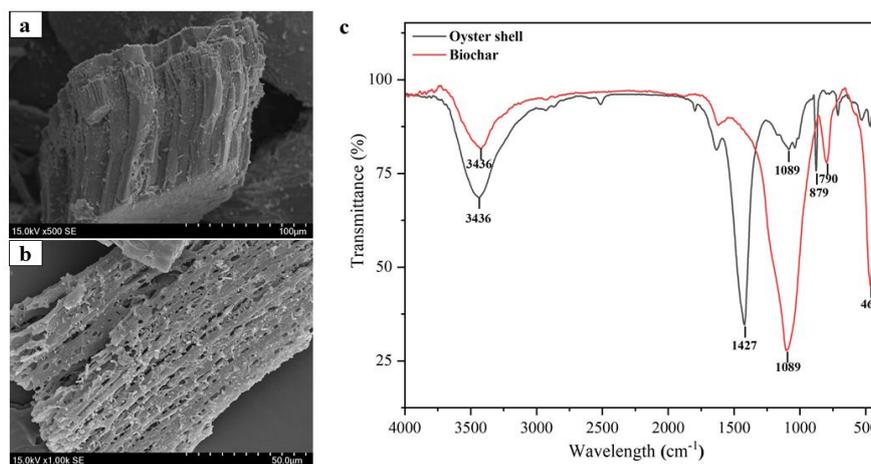


Figure 1 SEM images of oyster shell (a) and biochar (b) and FTIR spectra (c) of oyster shell and biochar.

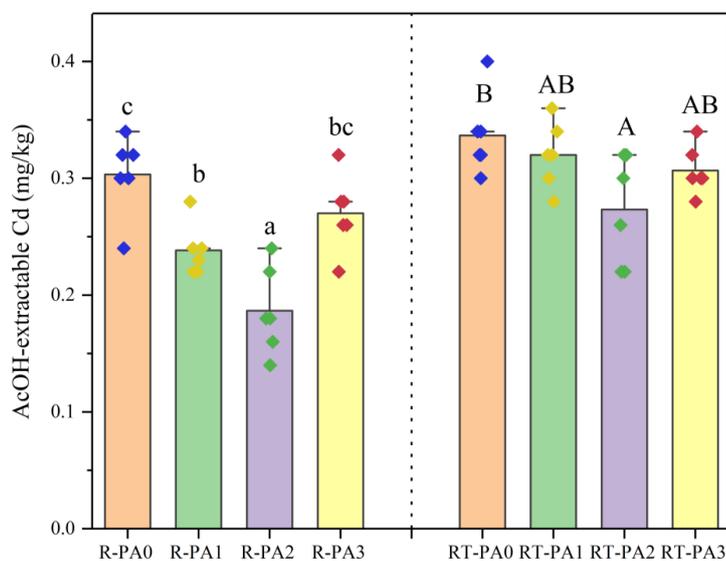


Figure 2 The effects of passivators on Cd bioavailability in soil. Dots represent the value of each sample. Bars followed with different lowercase letters (a - c) and capital letters (A, B) indicated significant ($p \leq 0.05$) difference among different treatments in rice planting and oilseed rape planting according to the LSD test. Values represent means \pm standard deviation.

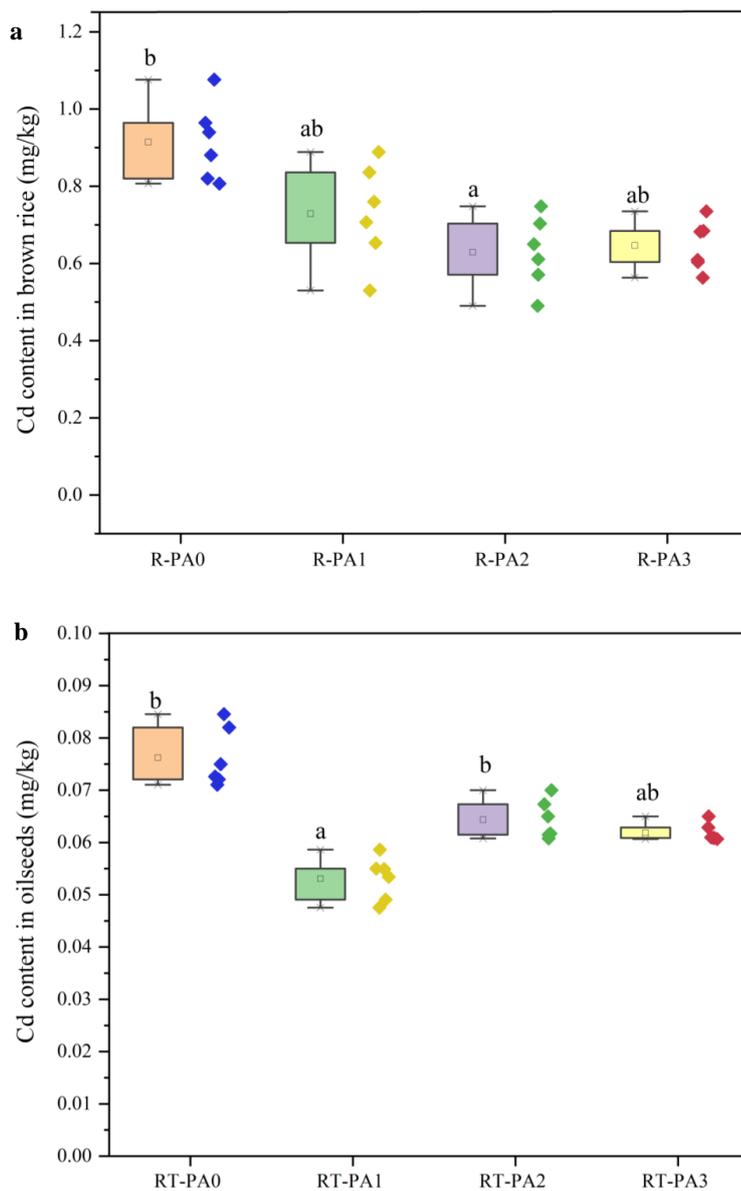


Figure 3 The effects of passivators on Cd contents in brown rice (a) and oilseed (b). Dots represent the value of each sample. Bars with different lowercase letters indicated significant ($p < 0.05$) difference among different treatments according to the LSD test. Values represent means \pm standard deviation.

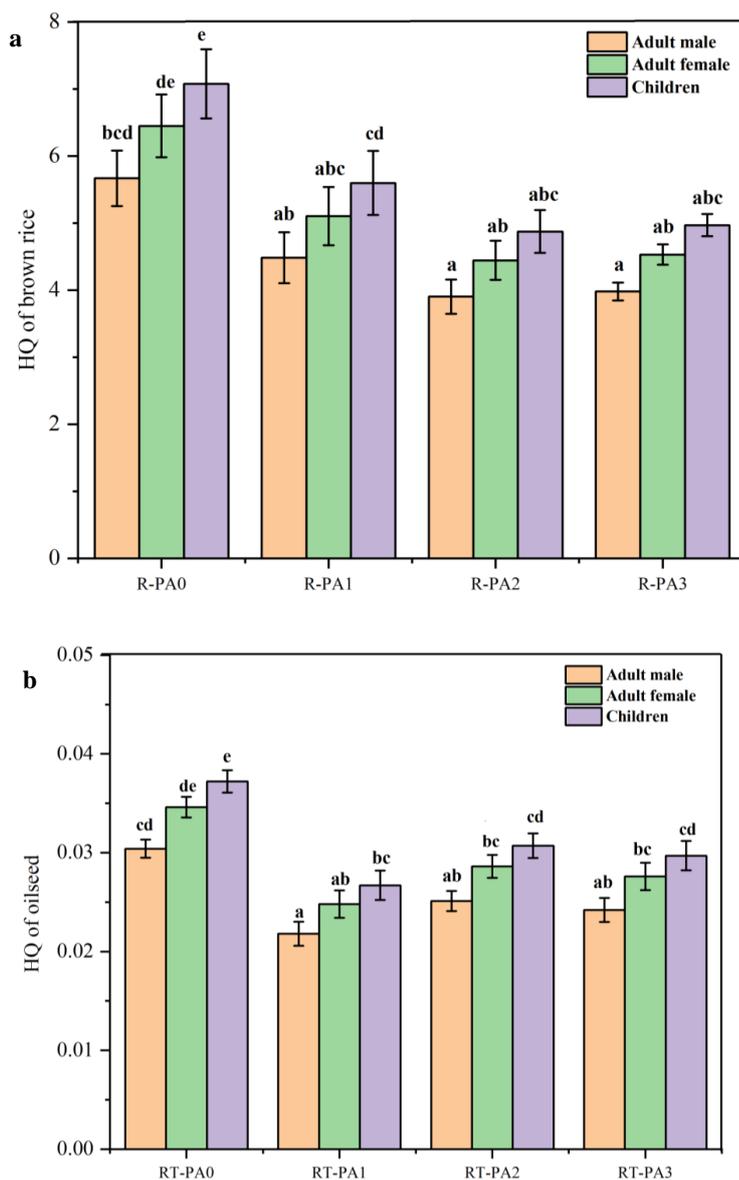


Figure 4 The effects of different passivators on the HQ of grown rice and oilseed. Mean with different lowercase letter indicated significant ($p < 0.05$) difference from each other according to the LSD test. Values represent means \pm standard deviation.

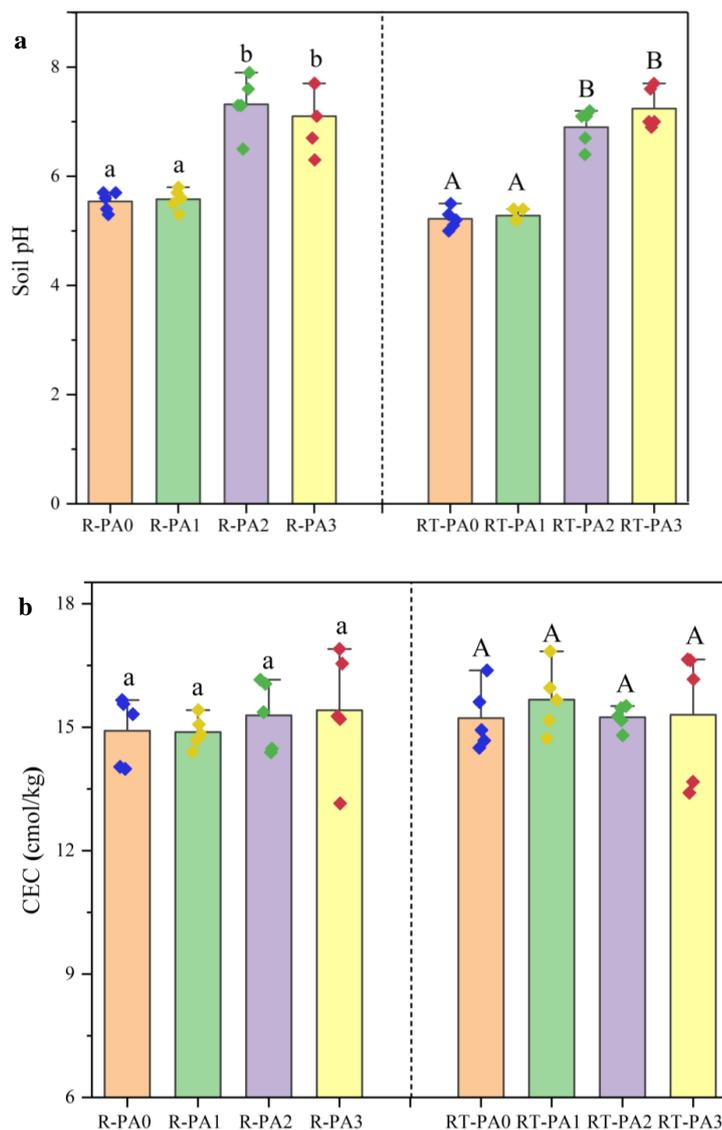


Figure 5 The effects of different passivators on soil pH (a) and CEC (b). Dots represent the value of each sample. Bars followed with different lowercase letters (a - c) and capital letters (A, B) indicated significant ($p \leq 0.05$) difference among different treatments in rice planting and oilseed rape planting according to the LSD test. Values represent means \pm standard deviation.

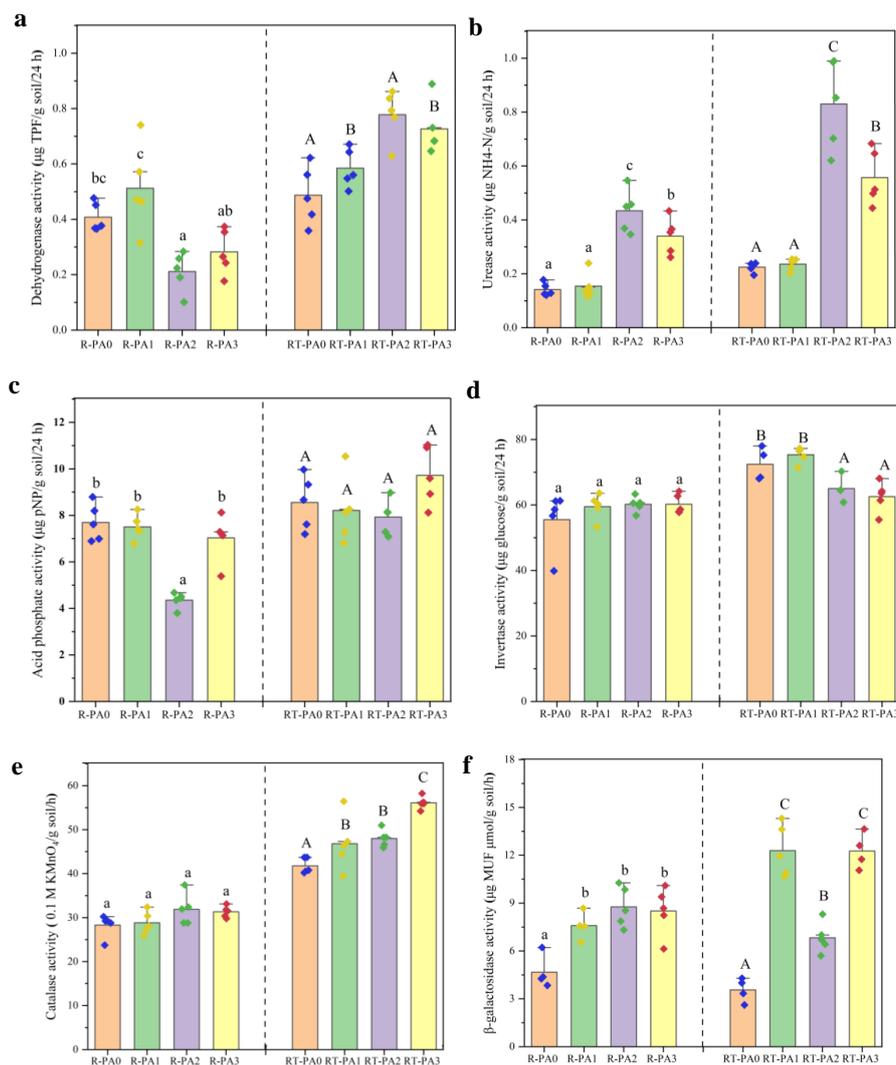


Figure 6 The effects of different passivators on the activities of soil enzyme. Dots represent the value of each sample. Bars followed with different lowercase letters (a - c) and capital letters (A-C) indicated significant ($p \leq 0.05$) difference among different treatments in rice planting and oilseed rape planting according to the LSD test. Values represent means \pm standard deviation.

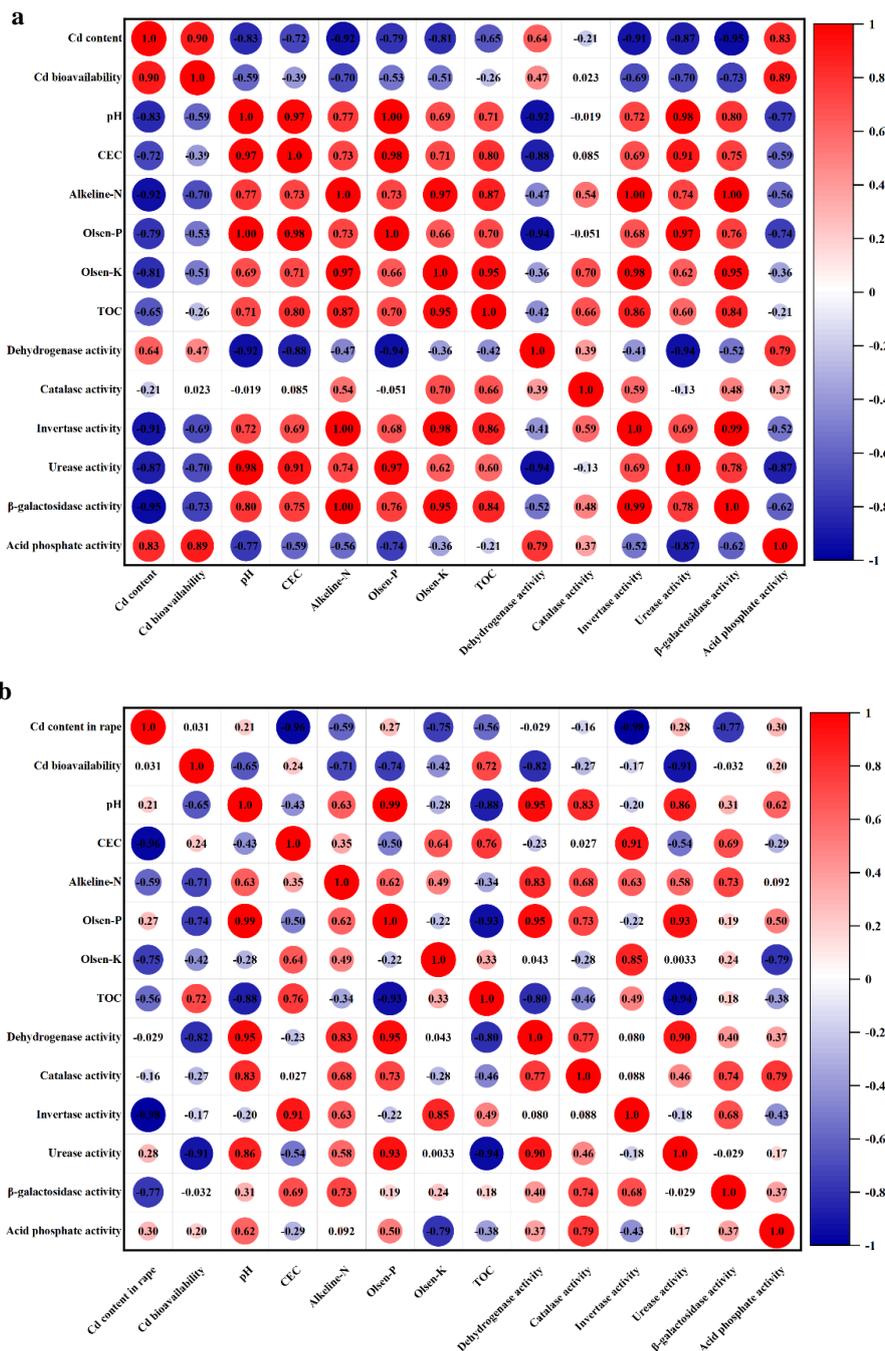


Figure 7 The correlation of investigated parameters in rice planting (a) and rice-oilseed rape rotation (b).



Code/Data availability

The data referring to this paper was all presented in the Supplemental file.

Author contribution

Bin Wu: Investigation, Writing Original Draft, Supervision

Jia Li: Writing - Review & Editing

Mingping Sheng: Investigation

He Peng: Investigation, Visualization

Dinghua Peng: Investigation, Data Curation

Heng Xu: Conceptualization, Resources, Funding acquisition

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.