ORIGINAL ARTICLE



Various *In Vitro* Bioactivities of Secondary Metabolites Isolated from the Sponge *Hyrtios aff. Erectus* from the Red Sea Coast of Egypt

Mısır'ın Kızıl Deniz Kıyısındaki Sünger *Hyrtios aff. Erectus*'tan İzole Edilen Farklı Sekonder Metabolitlerin Biyoaktiviteleri

Asmaa NABIL-ADAM1*, Mondamed A. SHREADAH1, Nondamed M. Abd EL MONEAM2, Nondamed Nondamed M. Abd EL

¹Marine Biotechnology and Natural Products Lab (MBNP), National Institute of Oceanography and Fisheries (NIOF), Alexandria, Egypt ²Alexandria University, Faculty of Science, Department of Biochemistry, Alexandria, Egypt

³Alexandria University, Faculty of Science, Department of Botany and Microbiology, Alexandria, Egypt

ABSTRACT |

Objectives: The present study revealed the presence of bioactive constituents in *Hyrtios aff. erectus* sponge (HES) extract collected from the Red Sea using skin and scuba diving.

Materials and Methods: Cytotoxicity was tested against hepatocellular carcinoma cell lines as a prescreening test.

Results: The HES extract had high contents of total phenolic compounds (0.061 mg/g), flavonoids (0.2839 mg/g), and carotenoids (1.976 mg/g). Moreover, the HES extract showed high antioxidant capacity with 93.0% and 99% at 1 mg using 2.2'-Diphenyl- α -picrylhydrazyl and 2.2'-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid), respectively. Cytotoxic activity against cancerous cell lines showed that the HES extract could inhibit cell growth effectively with IC $_{50}$ =47.5 µg/mL. Furthermore, anticancer activity using protein tyrosine kinase and sphingosine kinase 1 inhibitor screening assays resulted in 71.66% and 85.21% inhibition activity, respectively. The anti-inflammatory assays showed that the inhibition activity against cyclooxygenase (COX $_1$), COX $_2$, interleukin-6, and tumor necrosis factor- α was 71.82%, 81.13%, 80.89%, and 59.74%, respectively. At the same time, the anti-Alzheimer results using acetylcholine inhibition assay showed high activity at 1 mg with 83.51%. Additionally, the antiviral activity using the reverse transcriptase inhibition assay was 91.70%.

Conclusion: This marine sponge isolated from the Red Sea showed tremendous activity against many diseases and it is considered an excellent source for bioactive pharmaceutical compounds.

Key words: Red Sea, cytotoxic, antioxidant, anti-Alzheimer, anticancer, anti-inflammatory, antiviral

OZ

Amaç: Bu çalışma normal ve tüplü dalışlar ile Kızıl Deniz'den toplanan *Hyrtios aff. erectus* süngerindeki (HES) biyoaktif bileşiklerin varlığını göstermeyi amaçlamıştır.

Gereç ve Yöntemler: Sitotoksisite hepatosellüler karsinoma hücre hatlarında ön izleme testleriyle belirlenmiştir.

Bulgular: HES ekstresi yüksek derecede fenolik bileşikler, (0,061 mg/g), flavonoidler (0,2839 mg/g) ve karotenoidleri (1,976 mg/g) içermektedir. Ayrıca, 1 mg HES ekstresi 2'-difenil- α -pikrilhidrazil ve 2,2'-azino-bis (3-etilbenzotiyazolin-6-sulfonik asit) ile sırasıyla %93,0 ve %99 ile yüksek antioksidan kapasite göstermiştir. HES ekstresi kanseröz hücre hatlarına karşı sitotoksik bulunmuştur ve hücre büyümesini inhibe edebilmektedir (IC₅₀=47,5 μg/mL). Dahası, protein tirozin kinaz ve sfingosin kinaz 1 inhibitör izleme testleri kullanılarak belirlenen antikanser aktivitesi, sırasıyla %71,66 ve %85,21 inhibitör aktiviteyle sonuçlanmıştır. Antienflamatuvar testler siklooksijenaz 1 (COX₁), siklooksijenaz 2 (COX₂), interlökin-6 ve tümör nekroz faktör- α inhibitor aktivitesinin sırasıyla %71,82, %81,13, %80,89 ve %59,74 olduğunu göstermiştir. Aynı zamanda, asetil kolin esteraz inhibisyon teti kıllanılarak elde edilen anti-Alzheimer sonuçlar 1 mg dozda yüksek aktiviteyi (%83,51) belirlemiştir. Ek olarak, geri transkriptaz inhibisyon yöntemi kullanılarak bakılan antiviral aktivitesi %91,70'dir.

Sonuç: Kızıl Deniz'den elde edilen bu deniz süngeri birçok hastalığa karşı büyük bir aktivite göstermiştir ve biyoaktif farmasötik bileşikler için mükemmel bir kaynak kabul edilebilir.

Anahtar kelimeler: Kızıl Deniz, sitotoksisite, antioksidan, anti-Alzheimer, antikanser, anti-enflamatuvar, antiviral

INTRODUCTION

Marine habitats contain a broad range of different organisms having a variety of biochemical and physiological characteristics and ability to adapt to their environment. Marine organisms such as sponges, tunicates, fishes, soft corals, nudibranchs, sea hares, mollusks, echinoderms, bryozoans, prawns, shells, sea slugs, and marine microorganisms are sources of bioactive compounds. Marine sponges belonging to the phylum Porifera (Metazoa), evolutionarily the oldest animals, are the single best source of marine natural products. Very recently, marine sponges of the Red Sea have been recognized as a rich source of bioactive secondary metabolites. A great number of biologically active compounds with potential antitumor, anticancer, antimicrotubule, antiproliferative, cytotoxic, and photoprotective, as well as antibiotic and antifouling properties have been isolated.

The main objective of the present study was to investigate the tremendous activities of sponge secondary metabolites collected from the Red Sea as antioxidant, cytotoxic, anti-Alzheimer, anticancer, anti-inflammatory, and antiviral agents.

MATERIALS AND METHODS

Area of study

The Red Sea (Figure 1a) comprises a wide range of tropical marine habitats, many of which are internationally recognized for their conservation, scientific, economic, or recreational value.¹⁻⁸ It attracts many human activities, which in turn impact its environment⁹⁻¹⁸ and are likely to affect biological life and disturb the Red Sea's natural ecosystems.¹⁹⁻²⁶

Sampling, identification, and prescreening bioassays of the sponge Hyrtios aff. erectus

Hyrtios aff. erectus samples were collected from Hurghada on the Egyptian Red Sea coastline during spring 2014 (Figure 1b). The samples were collected using skin and scuba diving, processed, washed with freshwater, and transferred directly to the laboratory in sterile polyethylene bags under reduced temperature (0°C). Identification of the sponge species was kindly performed by Dr. Nicole Voogd, at the Naturalis Biodiversity Center, Department of Marine Zoology, RA Leiden, the Netherlands. The voucher specimen is incorporated in the collections of the Zoological Museum of the University of Amsterdam under registration number RMNH POR.8633.

Chemicals and solvents

Potassium ferricyanide, ferric chloride, NaOH, chloroform, glacial acetic acid, ferric chloride solution, H_2SO_4 , Folin-Ciocalteu reagent, vanillin, methanol, HCl, n-hexane, H_2O_2 , HNO $_3$, Se standard, Mn standard, β -carotene, catechin, (+)-quercetin, sodium nitrite, aluminum chloride, and gallic acid were purchased from Sigma Aldrich.

Instruments

Atomic absorption spectrophotometry [AAS and Graphite Furnace Atomizer (GFA) Shimadzu] and GC-MS (Thermo, USA) were applied.

Preliminary bioactive screening of Hyrtios aff. erectus sponge extract (HES)

The ethyl acetate extract of Hyrtios aff. erectus was subjected to different chemical tests for the detection of different phytoconstituents, i.e. tannins, phlobatannins, saponins, alkaloids, flavonoids, quinines, coumarin, terpenoids, and cardiac glycosides.²⁷

Quantitative chemotaxonomy profiling

Determination of total phenolic content in HES extract

Total phenolic compounds in Hyrtios aff. erectus extract were determined as described by Taga et al.²⁸

Determination of total flavonoid content in HES extract
Total flavonoid content was determined by a colorimetric method reported by Zhishen et al.²⁹

Determination of total tannins in HES extract

Tannins (proanthocyanidins) were determined according to the method described by Sun et al.³⁰

Determination of total carotenoids in HES extracts

Total carotenoid content was measured according to Thaipong et al. 27

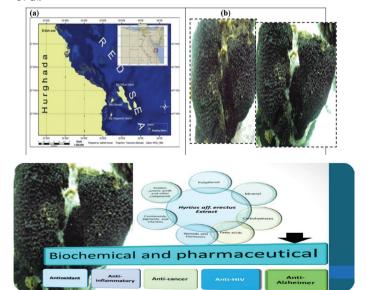


Figure 1. Location of sampling stations at the Red Sea (a) and sponge sample *Hyrtios aff. erectus* RMNH POR.8633 sample (b) and (c) represent the all steps of the present study (c) The schematic diagram of sponge extract constituents and multimedicinal effect

Preparation and extraction for mineral and metal assessment (Fe, Zn, Co, Mn, Cu, and Se) of HES extract

A 0.5 g dried sample of HES marine extract was digested using 5 mL of concentrated HNO $_3$, the mixture was heated using a hot plate for 1 h and when semidried 5 mL of concentrated HNO $_3$ and 2 mL of H $_2$ O $_2$ were added and it was kept on the hot plate for 1 h. The semidried cooled residue was filtered with the help of Whatman filter paper and the residue volume was made up to 25 mL with 2N HNO $_3$. Analysis was carried out using an AAS (GFA Shimadzu atomic absorption spectrophotometer AA–6800) according to the Official Methods of Analysis 31 for the determination of Fe, Zn, Co, Mn, Cu, and Se.

Elemental analysis of HES extract

The total carbon and hydrogen contents of marine HES extract were determined using a CHNO Elemental Analyzer.

Prescreening bioassays using in vitro cytotoxicity with cell lines

Different concentrations of HES extract (µg/mL) from all samples were tested for each cell line. Samples were dissolved in dimethyl sulfoxide (DMSO) and further diluted with cell culture medium. The final DMSO concentration used was 1% of total volume of the medium in all treatments, including the control group. Cells with no treatment were examined as negative and positive controls.³²

Primary screening assay

2.2'-Diphenyl- α -picrylhydrazyl radical scavenging effect of HES extract

A 2.2'-Diphenyl- α -picrylhydrazyl (DPPH) radical scavenging assay of the total *Hyrtios aff. erectus* extract was performed using a modified previously established methodology by Blois³³ and Amarowicz et al.³⁴ The scavenging ratio of the DPPH assay was calculated as follows:

% scavenging=[(A control-A sample)]/A control×100

2.2'-Azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) radical scavenging activity assay of HES extract

The 2.2'-Azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) (ABTS+) free radical decolorization assay was developed according to Chkraborty et al.³⁵ The percentage scavenging of ABTS+ was calculated by the following formula:

Scavenging activity (%)=(A₂-A₂)/A₂×100

 $\rm A_x$ and $\rm A_o$ were the absorbance at 734 nm of samples with and without extract, respectively.

Specialized screening assays

Acetylcholinesterase inhibition (AChEI) assay of HES extract Inhibition of acetylcholinesterase inhibition (AChE) by Hyrtios aff. erectus extract was evaluated as described by Moyo et al.³⁶ Percentage inhibition by extracts was calculated using the following equation:

Inhibition (%)=(1-sample reaction rate)/(blank reaction rate)×100

Determination of protein tyrosine kinase inhibitory activity of HES extract

Sample preparation

The dimethylsulfoxide (DMSO) sample solution of the appropriate extract was diluted with $\rm H_2O$ (1:1 v/v) to yield corresponding sample solutions (1 mg/mL). Tyrosine kinase (TK) inhibitory activity was determined using a commercial test kit (TK assay kit, nonradioactively, Takara Cat. #MK410). Protein tyrosine kinase (PTK) activity of the samples was calculated based on the prepared standard curve. The color intensity is stable for 1 h after addition of stop solution at room temperature in a light room.

% inhibition=
$$\frac{[Initial\ activity-inhibitor]}{Initial\ activity}$$

Determination of sphingosine kinase 1 inhibitor screening assay (SHK1) of HES extract

Sphingosine kinase inhibitory activity of the crude extract was determined by using the colorimetric sphingosine kinase 1 (SK1) inhibitor screening assay kit from Cayman. The plate was covered and the fluorescence was measured using an excitation wavelength between 530 and 540 nm and an emission wavelength between 580 and 590 nm.

$$\% \ inhibition = \frac{[Initial \ activity-inhibitor]}{Initial \ activity}$$

Determination of cyclooxygenase 1 (COX_1) and cyclooxygenase 2 (COX_2) inhibitor screening assay of HES extract

The COX inhibitory activity of the crude extract was determined using the colorimetric COX (ovine) inhibitor screening assay kit from Cayman. The absorbance was measured at 590 nm using a plate reader.

$$\% \ inhibition = \frac{[Initial\ activity-inhibitor]}{Initial\ activity}$$

Determination of tumor necrosis factor alpha (TNF- α) assay of HES extract

The TNF- α inhibitory activity of the crude extract was determined using a KOMA BIOTECH colorimetric kit. The absorbance was measured at 450 nm.

Determination of interleukin 6 (IL-6) assays of HES extract

The IL-6 inhibitory activity of the crude extract was also determined using a KOMA BIOTECH colorimetric kit. The absorbance was also measured at 450 nm.

Determination of reverse transcriptase (RT) enzyme inhibitor screening assay of HES extract

The RT inhibitory activity of the crude extract against a purified recombinant, human immunodeficiency virus (HIV-1)-RT, was determined using a Roche colorimetric kit. The assay was performed according to Fonteh et al.³⁷ with HIV-1 protease enzyme and the substrate, which is a synthetic peptide that

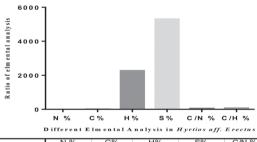
contains a cleavage site Tyr-Pro for HIV protease, as well as two covalently modified amino acids for the detection of cleavage. Acetyl pepstatin was used as a positive control for HIV-1 PR inhibition. The blank treatment consists of an assay buffer with only the substrate; untreated control of enzyme and substrate was also included. The absorbance was measured at 450 nm.

Statistical analysis

All results were analyzed by ANOVA using Prism.

RESULTS AND DISCUSSION

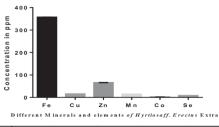
The secondary metabolites isolated from HES extract showed high contents of sulfur compounds (Figure 2). The mineral results showed high iron and zinc contents (Figure 3), in addition to polyphenol contents, which reflected high tannins and flavonoids. The crude extract of the sponge showed also high carotenoids contents (Figure 4). The bioactive profiling and diversity of natural compounds produced by sponge showed the presence of certain chemical classes of steroids, chromones, guinones, alkaloids, fatty acids; diketopiperazine, steroid, lactone, quinolone, anthraquinone, trisindole, phenol, and dihydropyridine benzoic acid derivatives; terpenoids; macrolactam; ethers; carboxylic acid; and terpenes, which are responsible for antioxidant, anti-inflammatory, antimicrobial, anti-HIV, anticancer, or antitumor activity. The quinolone derivatives are responsible for anti-HIV activity, fatty acid esters and fatty acids are responsible for anti-inflammatory activity, and pentaketides and alkaloids are responsible for neuroprotective activity.38 The present study revealed that this sponge has cytotoxicity against hepatocellular carcinoma (Table 1) (Figure 5). This finding agrees well with other research



	N %	С%	Н%	S%	C/N %	C/H %
Mean	3.490	7.680	2269	5301	48.73	74.25
Std. Deviation	0.01000	0.1200	0.06006	0.01001	0.2750	0.05508

Figure 2. The elemental analysis result of Hyrtios aff. erectus

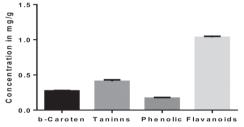
Std: Standard



	Fe	Cu	Zn	Mn	Co	Se
	356.8		65.55		1.990	7.886
Std. Deviation	0.2050	0.06506	0.4500	0.005774	0.01000	0.01445

Figure 3. The mineral profiling for *Hyrtios aff. erectus* Std: Standard

papers.^{4-8,39} Cytotoxic activity is considered the first parameter in screening for anticancer agents,⁴⁻⁸ while the cytotoxicity assay needs to be followed by other experiments to confirm their potential activity as an anticancer agent and to determine the mechanism. It has been reported that cell death can be induced through three mechanisms: apoptosis, autophagy, and oncosis.⁴⁰ In the present study the anticancer activity was determined through two different experimental models using TK and SK1 as anticancer targets.⁴¹ Sphingolipid-metabolizing enzymes have an important role in controlling the balance of the cellular levels of some important bioactive lipids, for example proliferative compound as well as apoptotic and ceramide compounds in addition to sphingosine 1-phosphate.⁴²



Different Phytochemical paramters of Hyrtiosaff. Erectus Extract

	b-Caroten	Taninns	Phenolic	Flavanoids
	0.2733	0.4100	0.1700	1.035
Std. Deviation	0.005773	0.02000	0.01000	0.01500

Figure 4. The polyphenol and carotenoid profiling for *Hyrtios aff. erectus* sponge extract

Std: Standard

Table 1. Inhibitory activities of *Hyrtios aff. erectus* sponge extract against hepatocellular carcinoma cells

Sample concentration (µg)	Viability %
50.00	47.83
25.00	69.17
12.50	80.24
6.25	93.62
3.125	97.89
1.56	100
0.00	100.00

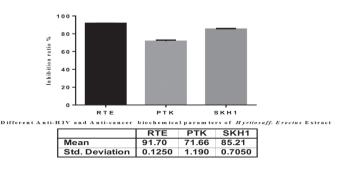


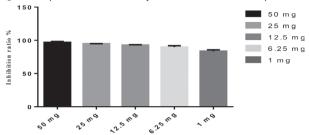
Figure 5. The anti-human immunodeficiency virus and anticancer profiling of Hyrtios aff. erectus sponge extract

Std: Standard, PTK: Protein tyrosine kinase, SKH: Sphingosine kinase RTE: Reverse transcriptase enzyme

The discovery of new chemotherapeutic resistance is an urgent and important challenge in oncology. Increased level of SK1 is considered a poor prognosis, and overexpression of SK1 means resistance to chemotherapeutics. Sphingosine kinase is involved in the development of different cancers and in chemotherapeutic resistance to drugs. Thus, SK1 represents an important target for anticancer drug therapy. Receptor tyrosine kinases are cell surface transmembrane proteins responsible for intracellular signal transduction. They are expressed in several cell types and, after activation by growth factor binding, trigger a series of intracellular pathways, leading to a wide variety of cell responses such as differentiation, proliferation, migration, invasion, angiogenesis, and survival. The overexpression of protein kinase members is associated with cancers and tumor cells. Therefore, tyrosine kinases are pivotal targets in drug therapy for cancer. The flavonoids, which are remarkably nontoxic⁴³ and could inhibit PTK and SKH activity, appear to have promising bioactivity as anticancer agents and are worthy of further investigation.⁴⁴ Phenolic compounds, especially flavonoids, exhibit anti-inflammatory and anticancer effects by inhibiting PTKs through several mechanisms. The first one is as an antioxidant and as being competitive inhibitors for the ATP binding sites on a variety of kinase enzymes. 45,46 Agullo et al.47 reported that the effectiveness of flavonoids depends mainly on the position, number, and substitution of the hydroxyl group of the β -ring. The saturation of the C_2 - C_2 bond is also an important factor that affects flavonoids inhibition of phosphatidylinositol 3-kinase. This can be easily found in marine natural products as more rings and chiral centers are there compared to synthetic compounds and drugs. Moreover, marine natural products provide molecules with larger molecular weight than synthetic compounds. While on average natural products contain fewer nitrogen, sulfur, and halogen atoms, they have higher ratios of these constituents compared to synthetic compounds and drugs.⁴⁸ Another explanation is that the pp60src gene product is a PTK, the activity of which has been shown to be inhibited by phenolic compounds, especially flavonoids.49 In the present study, the total polyphenolic assay, i.e. total phenolic and flavonoids, showed that the Hyrtios aff. erectus extract had high polyphenolic contents (Figure 4). Flavonoids are naturally occurring polyphenolic compounds that are present in a variety of natural products, and are the most abundant antioxidants in the human diet.50-52 While there has been a major focus on their antioxidant properties, there is an emerging view that flavonoids and their in vivo metabolites do not act only as conventional hydrogendonating antioxidants, but also to modulate cell function through actions at protein kinase and lipid kinase signaling pathways (PTK and SHK). These findings are in agreement with many other previous studies.53 In fact, flavonoids, and their metabolites, have been reported to act at PI 3-kinase, Akt/ protein kinase B, tyrosine kinases, protein kinase C, and mitogen-activated protein kinase signaling cascades. Inhibitory or stimulatory actions at these pathways are likely to affect cellular function profoundly by altering the phosphorylation state of target molecules and by modulating gene expression.53

An understanding of the mechanism of action of flavonoids. either as antioxidants or as modulators of cell signaling, is key to evaluating the potency of biomolecules as inhibitors of oxidative stress in general and in neurodegeneration.⁵⁴ The flavonoid compounds are characterized by their inhibitory effect on tyrosine kinase. Accordingly, the Hyrtios aff. erectus extract revealed the highest inhibition activity in PTK and SHK assays. Saponara et al.55 reported that the activity of pp60src gene product, which is a PTK, has been shown to be inhibited by flavonoids. Two major types of HIV have been identified so far, HIV-1 and HIV-2. HIV-1 is the cause of the worldwide epidemic and is most commonly referred to as HIV. The basic biological processes in the HIV-1 life cycle are now well established, and natural compounds targeting specific steps in this life cycle can be found.⁵⁶ HIV RT inhibitors include nucleotide RT inhibitors and non-nucleotide RT inhibitors. Most clinical anti-HIV drugs are HIV RT.57 In the last decade (2002-2011), 132 anti-HIV natural products were obtained from marine organisms. Of the anti-HIV bioactive marine natural products, before or after 2002, more than half were derived from marine sponges.⁵⁷ The present study indicated that the highest activities were from Hyrtios aff. erectus by 91.7, in agreement with previous studies.⁵⁸ Moreover, Simmons et al.59 concluded that sessile marine organisms (sponge and seaweeds) contain substances capable of potent biological activity, which has also been demonstrated against different types of cancer and HIV/Acquired Immune Deficiency syndrome. Restoring acetylcholine levels by inhibiting AChE has become the primary treatment for the cognitive deficits of Alzheimer disease (AD).60,61 The inhibition of AChE is beneficial not only to the enhancement of cholinergic transmission in the brain, but also to reduce the aggregation of β -amyloid and the formation of the neurotoxic fibrils in AD. In recent decades, researchers have attempted to develop new AChE inhibitors, especially the so-called "multifunctional AChE inhibitors" with additional efficacy in vascular dementia treatment.60 There have been plenty of phytochemicals found to be effective in inhibiting AChE, which mainly consist of alkaloids, cannabinoids, curcumins, stilbenes, and flavonoids.60 Among them, flavonoids have attracted more and more interest for their high inhibitory activity and low toxicity.⁶² Moreover, their diverse activities such as antioxidation, inhibition of advanced glycation products, and cardio-cerebrovascular protection give them extra advantages as potential multifunctional therapeutic agents for aging-related diseases.⁶³ The anti-Alzheimer results of the present study (Figure 6) using different extracts showed the highest inhibition ratio by Hyrtios aff. erectus, which produces secondary metabolites for defense against other microorganisms and these secondary metabolites serve as a source of bioactive compounds for use in human therapies as they thrive in harsh oceanic climates. 52,53,64 Many studies confirm the high activity of secondary metabolites isolated from marine Hyrtios aff. erectus including alkaloids, esters, fatty acids, glycosides, ketones, lipids, macrolides, alcohols, peptides, peroxides, polyketides, quinones, steroids, sterols, terpenes, and terpenoids. 52,53,65 Chronic inflammation is thought to play crucial roles in the pathogenesis of various

diseases. Several types of drugs are used to treat inflammatory disorders, but they cause adverse side effects. Natural products offer a great hope for the discovery of bioactive lead compounds.



	50 mg	25 mg	12.5 mg	6.25 mg	1 mg
Mean	96.99	95.02	92.89	89.82	83.51
Std. Deviation	1.365	0.02828	0.5515	2.001	2.135

Figure 6. The acetylcholinesterase inhibitor activity of *Hyrtios aff. erectus* extract

Std: Standard

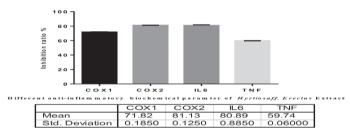


Figure 7. The anti-inflammatory profiling of *Hyrtios aff. erectus* extract Std: Standard, COX: Cyclooxygenase, IL-6: Interleukin-6, TNF: Tumor necrosis factor- α

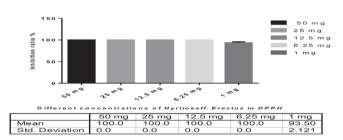
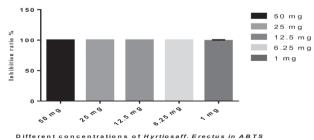


Figure 8. The total antioxidant capacity using scavenging (%) of 2,2'-Diphenyl- α -picrylhydrazyl assay

Std: Standard



	50 mg	25 mg	12.5 mg	6.25 mg	1 mg		
	100.0	100.0	100.0	100.0	99.00		
Std. Deviation	0.0	0.0	0.0	0.0	1.414		

Figure 9. The total antioxidant capacity using scavenging (%) 2,2'-Azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) assay

Std: Standard

These compounds can be developed into drugs for treatment of inflammatory disorders. The biological and chemical diversity of marine habitats constitutes a sizeable reservoir of novel compounds. Some of them, like sesquiterpenoids, diterpenes, steroids, polysaccharides, alkaloids, fatty acids, proteins, and other chemical compounds, isolated from marine organisms are found to exhibit anti-inflammatory activity 66 in agreement with our results as shown in Figure 7. Recently different compounds from sponge have been shown to have potent anti-inflammatory activity. 52,53 The natural products from marine sponge with different structures such as diterpenes, alkaloids, sulfated polysaccharides, and polyphenols inhibit different types of pro-inflammatory biomarkers, IL-6, TNF, NF- κ B, IL-1 β , COX, and COX, through different pathways:

- 1. The antioxidant effect by inhibition of the production of ROS compounds, which stimulate the pro-inflammatory biomarkers.⁶⁷
- 2. The direct effect by inhibition of prostaglandin and in sequence inhibition of the NF- κB cascaded stimuli also the TNF and IL-6. 68

When the human body faces a lot of stress, ROS are produced as a result.69 Deficiency of antioxidant agents leads to different degenerative diseases, 69 for example cardiovascular, Alzheimer, and various inflammatory diseases. 70 Consumption of antioxidants from natural sources reduces oxidative stress. Many studies showed that flavonoids and phenolic constituents have attributed to the antioxidant activities of natural compounds. Furthermore, many studies cited that minerals, for example Cu, Zn, Mg, Mn, and Se, played a significant role as antioxidants.71 Additionally, dietary antioxidants including tocopherols, carotenoids, and ascorbic acid have been investigated.72 Although many synthetic antioxidants have been shown to remediate oxidative stress, their lack of availability, high cost, and side effects remain the main challenge in dealing with oxidative stress, making the need to discover new antioxidant agents urgent. The sponge extract exhibits a potent antioxidant effect as shown in Figures 8 and 9, and the marine extract contains a variety of bioactive compounds known by their effect as antioxidants such as polyphenol (tannins, phenolic compounds, and flavonoids), carotenoids, and different minerals (Cu, Fe, Se, Zn, and Mn).

CONCLUSION

The secondary metabolites isolated from the sponge *Hyrtios aff. erectus* collected from the Red Sea in Egypt have been confirmed to have multimedicinal effects as anticancer, antiviral, anti-inflammation, and anti-Alzheimer agents. Further investigations should be performed to purify the pure compounds.

Conflict of interest: The authors declare that there is no conflict of interest.

REFERENCES

 Hegazy MF, Mohamed TA, Elshamy AI, Hassanien AA, Abdel-Azim NS, Shreadah MA, Abdelgawad II, Elkady EM, Pare PW. A new steroid

- from the Red Sea Soft Coral *Lobophytum Lobophytum*. Nat Prod Res. 2016;30:340-344.
- 2. Hegazy MF, Gamal-Eldeen AM, Mohamed TA, Alhammady MA, Hassanien AA, Shreadah MA, Abdelgawad II, Elkady EM, Pare PW. New Cytotoxic Constituents from the Red Sea Soft Coral *Nephthea* Sp., Nat Prod Res. 2016;30:1266-1272.
- Aboul-Ela H, Shreadah MA, Abdel-Monem NM, Yakout GA, Van Soest R. Isolation, cytotoxic activity and phylogenetic analysis of Bacillus sp. bacteria associated with the red sea sponge *Amphimedon ochracea*. Adv Biosci Biotechnol. 2012;3:815-823.
- AbdelMonein NM, Yacout GA, Aboul-Ela HM, Shreadah MA. Hepatoprotective activity of chitosan nanocarriers loaded with the ethyl acetate extract of Astenotrophomonas sp. bacteria associated with the Red Sea sponge Amphimedon Ochracea In CCl₄ induced hepatotoxicity in rats. Adv Biosci Biotechnol. 2017;8:27-50.
- Abd El Moneam NM, Al-Assar SA, Shreadah MA, Nabil-Adam A. Isolation, Identification and Molecular Screening of Psudomance Sp Metabolic pathways NRPs and PKS associated with the Red Sea sponge, Hyrtios aff. Erectus, Egypt. J Pure Appl Microbio. 2017;11:1299-1311.
- Abd El Moneam NM, Shreadah MA, Al-Assar SA, Nabil-Adam A. Protective role of antioxidants capacity of Hyrtios aff. Erectus sponge extract against mixture of persistent organic pollutants (POPs)-induced hepatic toxicity in mice liver: biomarkers and ultrastructural study Environ Sci Pollut Res. 2017:1-12.
- Shreadah MA, Abd El Moneam NM, El-Assar SA, Asmaa Nabil-Adam A. Metabolomics and Pharmacological Screening of Aspergillus versicolor Isolated from Hyrtios Erectus Red Sea Sponge; Egypt. Current Bioactive Compounds. 2019.
- 8. Shreadah MA, Abd El Moneam NM, Al-Assar SA, Nabil-Adam A. The Ameliorative Role of a Marine Sponge Extract against Mixture of Persistent Organic Pollutants induced Changes in Hematological Parameters in Mice. Expert Opinion Environ Biol. 2017;6.
- Shriadah MA, Okbah MA, El-Deek MS. Trace metals in the water columns of the Red Sea and the Gulf of Aqaba, Egypt. Water Air and Soil Pollut. 2004;153:115-124.
- Shreadah MA, Said TO, El Zokm GM, Masoud MS. Physico-Chemical Characterititics of the Surficial Sediments along the Egyptian Red Sea Coasts. Egyptian J Aqu Res. 2008;34:16-34.
- Shreadah MA, Said TO, Abd El Ghani SA, Ahmed AM. Alkyllead and Alkyltin Species in different fishes collected from the Suez Gulf, Egypt. Egyptian J Aqu Res. 2008;34:64-73.
- Shreadah MA, Masoud MS, Said TO, El Zokm GM. Application of IR, X-Ray, TGA and DTA to determine the mineral composition of the Sediments and study of reaction kinetics along the Egyptian Red Sea Coasts. Egyptian J Aqu Res. 2008;34:83-95.
- Shreadah MA, Said TO, Abdel Ghani SA, Ahmed AM. Distribution of Different Organotinand Organolead Compounds in Sediment of Suez Gulf. J Environ Protect. 2011;2:545-554.
- Okbah MA, Shata MA, Shriadah MA. Gochemical forms of trace metals in mangrove sediments-Red Sea (Egypt). Chem Ecol. 2005;21:23-36.
- 15. Abdel Ghani SA, Shobier AH, Said TO, Shreadah MA. Organotin compounds in Egyptian Mediterranean sediments. Egyptian J Aqu Res. 2010;36:221-229.

- Masoud MS, Said TO, El- Zokm GM, Shreadah MA. Speciation of Fe, Mn and Zn in Surficial Sediments from the Egyptian Red Sea Coasts. Chem Spec Bioavailab. 2010;22:257-269.
- Masoud MS, Said TO, El-Zokm GM, Shreadah MA. Assessment of Heavy Metals Contamination in Surface Sediments of the Egyptian Red Sea Coasts. Aust J Basic Appl Sci. 2012;6:44-58.
- Said TO, Shreadah MA, AbdelGhani SA, Ahmed AM. Alkyltin and alkayllead compounds in coastal water of Suez Gulf, Egypt. Egyptian J Aqu Res. 2010;36:33-42.
- Fahmy MA, Shriadah MA, AbulSoud A, Abdel Rahman SM, Shindy M. Hydrography and Chemical Characteristics of the Coastal Water along the Gulf of Suez. Egyptian J Aquatic Res. 2005;31:1-14.
- Fahmy MA, Abdel Fattah LM, Abdel-Halim AM, Abdel Nabi MA, Abo-El-Khair EM, Ahdy HH, Hemeilly A, Abu El-Soud A, Shreadah MA. Evaluations of the Coastal Water Quality of the Egyptian Red Sea during 2011-2013. J Environ Protect. 2016;7:1810-1834.
- Abdel-Halim AM, Aboel-Khair EM, Fahmy MA, Shreadah MA. Environmental Assessment on the Aqaba Gulf Coastal waters, Egypt. Egyptian J Aqu Res. 2007;33:1-14.
- 22. Abdel-Halim AM, Aly-Eldeen MA, Abdel Fattah LM, Fahmy MA, Abo-El-Khair EM, Khaled AM, El-Soud AA, Shreadah MA. Environmental studies on the Aqaba Gulf coastal waters during 2011-2013. J Environ Protect. 2016;7:1411-1437.
- Abo-el-Khair EM, Abdel Halim AM, Shriadah MA, Fahmy MA. Environmental Conditions of the Suez Gulf and the Red Sea Coastal Waters, Egypt. In: Ozhan E, ed. Proceedings of the 8th International Conference on the Mediterranean Coastal Environment. MEDCOAST 2007. 13-17 November 2007. Alexandria Egypt. 2007:517-526.
- 24. Abo-El khair EM, Abdel Halim AM, Fahmy MA, Shreadah MA. Environmental Impact Assessment of Northern Red Sea Regions during 2005 2007. Egyptian J Aqu Res. 2008;34:20-30.
- Abo-El-Khair EA, Abdel Fattah LM, Abdel-Halim AM, Abdel Nabi MA, Fahmy MA, Ahdy HH, Hemeilly A, Abu El-Soud A, Shreadah MA. Assessment of the hydrochemical characteristics for the coastal waters of the Suez Gulf during 2011-2013. J Environ Protect. 2016;7:1497-1521.
- Gurguess SM, Shreadah MA, Fahmy MA, Aboul El Kheir EM, Abdel Halim A. Assessment of Water Quality in the Red Sea using in Situ Measurements and Remote Sensing Data. Egyptian J Aqu Res. 2009;35:1-13.
- Thaipong K, Boonprakob U, Crosby K, Cisneros-Zevallos L, Byrne DH. Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruit extracts. J Food Compos Anal. 2006;19:669-675.
- Taga MS, Miller EE, Pratt DE. Chia seeds as a source of natural lipid antioxidants. J Am Chem Soc.1984;61:928-931.
- 29. Zhishen J, Mengcheng T, Jianming W. The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. Food Chem. 1999;64:555-559.
- Sun B, Richardo-Da-Silvia JM, Spranger I. Critical factors of vanillin assay for catechins and proanthocyanidins. J Agric Food Chem. 1998;46:4267-4274.
- AOAC. Official Methods analysis of association of official analytical chemists 15th End., Association of official analytical chemists. Washington DC, USA; 1990.

- Kosanic M, Rankovic B, Stanojkovic T. Biological activities of two macroalgae from Adriatic coast of Montenegro. Saudi J Biol Sci. 2015;22:390-397.
- Blois MS. Antioxidant Determinations by the Use of a Stable Free Radical. Nature. 1958;181:1199-1200.
- 34. Amarowicz R, Naczk M, Zadernowski R, Shahidi F. Antioxidant activity of condensed tannins of beach pea, Canola hulls, evening primrose, and faba bean. J Food Lipids. 2000;7:195-205.
- Chkraborty K, Lipton AP, Paul Raj R, Vijayan KK. Antibacterial labdane diterpenoids of Ulva fasciata Delile from southwestern coast of the Indian Peninsula. Food Chem. 2010;119:1399-1408.
- Moyo SJ, Aboud S, Kasubi M, Lyamuya EF, Maselle SY. Antimicrobial resistance among producers and non-producers of extended spectrum beta- lactamases in urinary isolates at a tertiary Hospital in Tanzania. BMC Res Notes. 2010;3:348.
- 37. Fonteh PN, Keter FK. Meyer D. New bis(thiosemicarbazonate) gold (III) complexes inhibit HIV replication at cytostatic concentrations: potential for incorporation into virostatic cocktails. J Inorg Biochem. 2011;105:1173-1180.
- Thomas TR, Kavlekar DP, LokaBharathi PA. Marine Drugs from Sponge-Microbe Association--A Review. Mar Drugs. 2010;8:1417-1468.
- Abdelmohsen UR, Grkovic T, Balasubramanian S, Kamel MS, Quinn RJ, Hentschel U. Elicitation of secondary metabolism in actinomycetes. Biotechnol Adv. 2015;33:798-811.
- 40. Elmore S. Apoptosis: A Review of Programmed Cell Death. Toxicol Pathol. 2007;35:495-516.
- Min J, Traynor D, Stegner AL, Zhang L, Hanigan MH, Alexander H, Alexander S. Sphingosine Kinase Regulates the Sensitivity of *Dictyostelium discoideum* Cells to the Anticancer Drug Cisplatin. Eukaryot Cell. 2005;4:178-189.
- 42. Ponnusamy S, Meyers-Needham M, Senkal CE, Saddoughi SA, Sentelle D, Selvam SP, Ogretmen B. Sphingolipids and cancer: ceramide and sphingosine-1-phosphate in the regulation of cell death and drug resistance. Future Oncol. 2010;6:1603-1624.
- 43. Rice-Evans CA, Miller NJ, Paganga G. Structure-antioxidant activity relationships of flavonoids and phenolic acids. Free Radic Biol Med. 1996;20:933-956.
- Shreadah MA, Abd El Moneam NM, El-Assar SA, Nabil-Adam A. Marine Algae of the Genus Gracilaria as a Multi Products Source for different Biotechnological and Medical Applications. Recent Pat Biotechnol. 2020.
- Santos CC, Salvadori MS, Mota VG, Costa LM, de Almeida AA, de Oliveira GA, Costa JP, de Sousa DP, de Freitas RM, de Almeida RN. Antinociceptive and Antioxidant Activities of Phytol *In Vivo* and *In Vitro* Models. Neurosci J. 2013;2013:949452.
- Guerra B, Issinger OG. Natural Compounds and Derivatives as Ser/ Thr Protein Kinase Modulators and Inhibitors. Pharmaceuticals (Basel). 2019:12.
- 47. Agullo G, Gamet-Payrastre L, Manenti S, Viala C, Rémésy C, Chap H, Payrastre B. Relationship between *flavonoid* structure and inhibition of phosphatidylinositol 3-kinase: a comparison with tyrosine kinase and protein kinase C inhibition. Biochem Pharmacol. 1997;53:1649-1657.
- 48. Lahlou M. The Success of Natural Products in Drug Discovery. Pharmacol Pharm. 2013;4:17-31.

- Chahar MK, Sharma N, Dobhal MP, Joshi YC. Flavonoids: A versatile source of anticancer drugs. Phoogn Rev. 2011;5:1-12.
- 50. Kumar S, Pandey AK. Chemistry and Biological Activities of Flavonoids: An Overview. Scientific World Journal. 2013;2013:162750.
- 51. Abd El Moneam NM, Shreadah MA, El-Assar SA, De Voogd NJ, Nabil-Adam A. Hepatoprotective effect of Red Sea sponge extract against the toxicity of a real-life mixture of persistent organic pollutants. Biotecnol Biotec Eq. 2018;32:734-743.
- Shreadah MA, El Moneam NMA, Al-Assar SA, Nabil-Adam A. Phytochemical and pharmacological screening of Sargassium vulgare from Suez Canal, Egypt. Food Sci Biotechnol. 2018;27:963-979.
- 53. Kasote DM, Katyare SS, Hegde MV, Bae H. Significance of Antioxidant Potential of Plants and its Relevance to Therapeutic Applications. Int J Biol Sci. 2015;11:982-991.
- Williams RJ, Spencer JP, Rice-Evans C. Flavonoids: antioxidants or signalling molecules? Free Radic Biol Med. 2004;36:838-849.
- Saponara S, Carosati E, Mugnai P, Sgaragli G, Fusi F. The flavonoid scaffold as a template for the design of modulators of the vascular Ca (v) 1.2 channels. Br J Pharmacol. 2011;164:1684-1697.
- Neagu IA, Olejarz J, Freeman M, Rosenbloom DIS, Nowak MA, Hill AL. Life cycle synchronization is a viral drug resistance mechanism. PLoS Comput Biol.2018;14:e1005947.
- Carroll AR, Copp BR, Davis RA, Keyzers RA, Prinsep MR. Marine natural products. Nat Prod Rep. 2019;36:122-173.
- 58. Abdel-Monem NM, Abdel-Azeem AM, El-Ashry el-SH, Ghareeb DA, Nabil-adam A. Pretreatment Hepatoprotective Effect of the Marine Fungus Derived from Sponge on Hepatic Toxicity Induced by Heavy Metals in Rats. Biomed Res Int. 2013;2013:510879.
- Simmons TL, Andrianasolo E, McPhail K, Flatt P, Gerwick WH. Marine natural products as anticancer drugs. Mol Cancer Ther. 2005;4:333-342.
- Mendiola-Precoma J, Berumen LC, Padilla K, Garcia-Alcocer G. Therapies for Prevention and Treatment of Alzheimer's Disease. Biomed Res Int. 2016;2016:2589276.
- 61. Howes MJ, Perry E. The role of phytochemicals in the treatment and prevention of dementia. Drugs Aging. 2011;28:439-468.
- 62. Uriarte-Pueyo I, Calvo MI. Flavonoids as acetylcholinesterase inhibitors. Curr Med Chem. 2011;18:5289-5302.
- 63. Gauthier S, Leuzy A, Racine E, Rosa-Neto P. Diagnosis and management of Alzheimer's disease: past, present and future ethical issues. Prog Neurobiol. 2013;110:102-113.
- 64. Nong E, Lee W, Merriam JE, Allikmets R, Tsang SH. Disease progression in autosomal dominant cone-rod dystrophy caused by a novel mutation (D100G) in the GUCA1A gene. Doc Ophthalmol. 2014;128:59-67.
- Mehbub MF, Lei J, Franco C, Zhang W. Marine Sponge Derived Natural Products between 2001 and 2010: Trends and Opportunities for Discovery of Bioactives. Mar Drugs. 2014;12:4539-4577.
- Odeleye T, White WL, Lu J. Extraction techniques and potential health benefits of bioactive compounds from marine molluscs: a review. Food Funct. 2019;10:2278-2289.
- 67. Birben E, Sahiner UM, Sackesen C, Erzurum S, Kalayci O. Oxidative stress and antioxidant defense. World Allergy Organ J. 2012;5:9-19.

- 68. Wojdasiewicz P, Poniatowski ŁA, Szukiewicz D. The role of inflammatory and anti-inflammatory cytokines in the pathogenesis of osteoarthritis. Mediators Inflamm. 2014;2014:561459.
- Lobo V, Patil A, Phatak A, Chandra N. Free radicals, antioxidants and functional foods: Impact on human health. Pharmacogn Rev. 2010;4:118-126.
- 70. Leonard WR, Snodgrass JJ, Robertson ML. Effects of brain evolution on human nutrition and metabolism. Annu Rev Nutr. 2007;27:311-327.
- 71. Arulselvan P, Fard MT, Tan WS, Gothai S, Fakurazi S, Norhaizan ME, Kumar SS. Role of Antioxidants and Natural Products in Inflammation. Oxidn Med Cell Longev. 2016;5276130.
- Pertuzatti PB, Sganzerla M, Jacques AC, Barcia MT, Zambiazi RC. Carotenoids, Tocopherols and Ascorbic Acid content in Yellow Passion Fruit (Passiflora Edulis) Grown Under Different Cultivation Systems. LWT- Food Sci Tech. 2015; 64:259-263.