

RESEARCH PAPER

Efficacy of Clove Oil, 2-Phenoxyethanol and Benzocaine on European Catfish, Silurus glanis Linnaeus 1758

Kaya Gökçek¹, Fatih Öğretmen², Mahir Kanyilmaz³

¹ Mustafa Kemal University, Agriculture Faculty, Animal Sciences Department Antakya, Hatay, Turkey.

² Muğla Sıtkı Koçman University, Faculty of Fisheries and Aquaculture, Aquaculture Department, Muğla, Turkey.

³Mediterranean Fisheries Research, Production and Training Institute, Döşemealtı, Antalya, Turkey.

* Corresponding Author: Tel.: +90.534 4150716;	Received 30 June 2016
E-mail: kayagokcek@yahoo.com	Accepted 17 August 2016

Abstract

The efficacy of clove oil, 2-phenoxyethanol and benzocaine was tested on captive-bred European catfish (wels), Silurus glanis L., 1758 in this study. According to effectiveness criteria, the most optimal implementation results were obtained from 100 μ L⁻¹ (induction 69.00±6.48 s and recovery 253.86±14.78 s) clove oil, 1000 μ L⁻¹ (induction 88.71±8.50 s and recovery 201.43 ± 12.19 s) PE, and $100 \ \mu$ L⁻¹ (induction 103.00 ± 6.90 s and recovery 215.71 ± 19.99 s) benzocaine. The concentration of anesthetics agent used directly affected the onset of each physiological respond (p<0.05). The relationship between doses and induction times was inverse exponential, in spite of that an exponential relationship was confirmed between doses and recovery times for all anesthetics. In conclusion, all these three anesthetics can be effectively used for wels.

Keywords: European catfish, clove oil, 2-phenoxyethanol, benzocaine, anesthesia.

Introduction

The anesthetics are commonly used in aquatic culture activities such as classification, breeding, weighing, broodstock management and drug applications against diseases. Anesthetics cause reduction of fish activity and the total loss of consciousness is the last step of treatment. Furthermore, apnea and low oxygen saturation level in the blood may be caused due to over dose (Tytler and Hawkins, 1981; Gomulka et al., 2015).

Clove oil, eugenol, benzocaine (ethyl-paminobenzoate), 2-phenoxyethanol (ethylene glycol monophenyl ether) and MS-222 (tricaine methane sulphonate) are the commonly used anesthetics by aquaculture farms and researchers (Velisek et al., 2006, 2011). Although clove oil, eugenol, benzocaine and 2-phenoxyethanol are generally used for non-food fish and researches, MS-222 is the only licensed product for cultured fish species in USA and EU (Öğretmen and Gökçek, 2013). On the other hand, although Aqui-S is still illegal for USA and many other countries, it has been approved for use in food fish production in some southern hemisphere countries (Zahl et al., 2012). Recently, a new agent which is called Sedanol was practically started to be used in Turkey.

An appropriate anesthetic should possess some

features such as being inexpensive, accessible, nontoxic etc. (Treves-Brown, 2000). Each fish species responses to the same anesthetics at considerably varied concentrations, thus determination of the lowest effective doses of different matters is very important (Pawar et al., 2011). Also, recovery duration and survival ratio are substantial points on choosing an effective species specific anesthetic (Gilderus and Marking, 1987; Burka et al., 1997).

The European catfish or wels, S. glanis L., 1758, is a carnivorous and fast growing species. It is commercially produced more than a hundred years in especially central Europe due to high value as food. Although reproduction and hatchery technics, yield efficiency in polyculture and biology were already studied, the effect of commercially used anesthetics on European catfish is still obscure. Recently, intensive culture in cages started and production ratio is expected to be increased in the near future in Turkey. Due to increasing interest on intensive culture Wels and inconclusive information about of anesthetics, the main aim of this study was to reveal the effective doses of three different anesthetics (clove oil, benzocaine and 2-phenoxyethanol) that could be effectively used in European catfish under controlled conditions.

© Published by Central Fisheries Research Institute (CFRI) Trabzon, Turkey in cooperation with Japan International Cooperation Agency (JICA), Japan

Material and Methods

Fish

Wels were obtained from a commercial fish farm in Kahramanmaraş, Turkey (Tuğçe Ltd.). Fish were transferred to the hatchery of company for acclimation 2 weeks before the study. Six-month old individuals (n=105, average length 15.6 ± 1.5 cm and weight $44,6\pm6.7$ g) were not fed the day before study.

Anesthetic Agents

In this study, clove oil (Aromax, Hungary), benzocaine (Himedia, India) and 2-phenoxyethanol (PE)(Sigma Aldrich Chemic, Germany) were tested as the anesthetic agents. Doses were prepared just before the trials. Clove oil and benzocaine were diluted in ethanol (94%) (1:9) due to low water solubility. In a similar way, PE was also diluted in ethanol (1:1) in a falcon tube before implementation to prevent small droplets in the water (Öğretmen and Gökçek, 2013).

Experimental Design

Five concentrations of three anesthetic agents were tested on Wels to find out the effective dose. The minimum and maximum concentrations were chosen according to previously published studies (Weber *et al.*, 2009; Gökçek and Öğretmen, 2011). The concentrations were adjusted for clove oil and benzocaine 25, 50, 75, 100 and 125 μ L/L, and for PE 250, 500, 750, 1000 and 1250 μ L/L. Each replicate contains seven individuals and all concentrations were tested in triplicate.

During the implementation, fish were netted and transferred to the 20 L aquarium containing different concentration of anesthetic solutions. The induction and recovery times for all agents were measured by a stopwatch under the same conditions. Fish were immediately transferred to the anesthetic free water just after they reached the last anesthesia stage. The recovery stage was recorded after transferring the fish to the aerated clean water in 20 L aquarium. Also, water quality parameters such as dissolved oxygen, temperature and pH were monitored by oxygen meter (Hanna HI-9146) and a pH meter (Hanna) in aquariums and detected as 8.01±0.93 mg/L, 20°C and

8.63±0.22, respectively.

Four consecutive stages for induction and three stages for recovery were specified according to Theinpoint and Niemegeers (1965). Some modifications were put to use based on behavioral response of Wels (Table 1).

Statistics

The relationship between dosage and induction/recovery times were stated by Regression analyses. Shapiro-Wilk test for normality and Bartlett's test for homogeneity of variance was used. One-way analyzes of variance (ANOVA) was used to determine the differences in time of occurrence of physiological responses to the different doses of the same anesthetic agent compare the means and Duncan test was employed to analyze differences between means. Significance of differences was tested at P<0.05 level. All statistical processes were analyzed by the SPSS (SPSS Systems for Windows, Version 15.0).

Results

Physiological responses to the different doses of anesthetic agents were given in Table 2. Induction times significantly decreased with increasing concentrations in all agents, except the lowest concentrations (P<0.05). All the lowest concentrations were not efficient enough for fish to get the stages I3 and I4. On the other hand, recovery times increased with increasing concentrations of anesthetics (P<0.05).

A significant relation was observed between different concentrations and induction/recovery times for all tested agents (P<0.05). While scatter plots yielded an inverse exponential relationship for induction, the scatter plots showed exponential relationships in recovery (Figure 1). The equations of induction times (in seconds) to reach I4 stage of anesthesia and concentrations (c) of anesthetics were I4= 458.99e^{-0.019c} (R²=0.9759) for clove oil, I4=310.42e^{-0.001c} (R²=0.9368) for PE and I4=240.37e^{-0.009c} (R²=0.9482) for benzocaine. The regression equations established for recovery time and concentrations were R3=48.02e^{0.017c} (R²=0.9549) for clove oil, R3=49.05e^{0.002c} (R²=0.9478) for PE and

Table 1. Behavioral stages of anesthesia in Wels, S. glanis (modified from Theinpoint and Niemegeers, 1965)

Induction stages

Recovery stages

R 2: Regular breathing. Reaction to strong stimuli. Irregular balance

I 1: Respiratory disorder: Operculum excessively comes open

I 2: Loss of balance: partial inhibition of reactions to external stimuli

I 3: Total loss of equilibrium: Fish still react to strong stimuli

I 4: Total loss of reflexes and movement: Fish lay on the bottom of the tank

R 1: Start of movement. Fish still lay on bottom of the tank

R 3: Total recovery of equilibrium. Reaction to slight stimuli. Normal swimming

R3=54.80 $e^{0.014c}$ (R²=0.9506) for benzocaine. No mortality was observed after treatments.

Discussions

There are several biological and environmental factors which effects to the efficiency of an anesthetic agent such as specie, size, weight, age, tissue fat content, water temperature and pH, etc. (Iversen *et al.*, 2003). Furthermore, the efficacy of an anesthetic is directly affected by metabolic rate (Burka *et al.*, 1997; Ross and Ross, 1999).

In this study, induction times significantly decreased with increasing of concentrations of clove oil, 2-phenoxyethanol and benzocaine (P<0.05). Similar results were gained by previous studies in teleost fish (Hseu et al., 1998; Mylonas et al., 2005; Gullian and Villanueva, 2009; Weber at al., 2009; Heo and Shin, 2010; Gökçek and Öğretmen, 2011; Öğretmen and Gökçek, 2013; Öğretmen et al., 2014). On the other hand, recovery times increased with increasing concentrations of three anesthetic agents in wels (P<0.05). Prolonged recovery has been reported in Persian sturgeon, Acipencer persicus (Bagheri and Imanpour, 2011), Russian sturgeon, Α. gueldenstaedtii (Akbulut et al., 2011), Himri barbell, Carasobarbus luteus (Gökçek and Öğretmen, 2011), African catfish, Clarias gariepinus (Öğretmen and Gökçek, 2013), and Shabut, Barbus grypus (Öğretmen et al., 2014). However, decreasing recovery times with increasing concentrations of clove oil and 2-phenoxyethanol was reported for Sea bass and Sea bream by Mylonas *et al.* (2005). The authors explained this unexpected situation by that the fish did not contact with the highest concentrations of anesthetic agents for long which allows recovered fast (Pawar *et al.*, 2011).

The effectiveness criteria for an anesthetic agent is to supply a complete induction in 180 s and recovery in 300 s for teleosts (Marking and Meyer, 1985). In the present study, the application of 75 and 100 μ l L⁻¹ clove oil, 500, 750 and 1000 μ l L⁻¹ PE, and 50, 75 and 100 μ l L⁻¹ benzocaine resulted adequate induction (<180 s), and recovery (<300 s). Although highest concentrations achieved shorter induction time for wels, aforementioned concentrations were presented a good margin of safety when compare against the efficacy criteria above. Fish wasn't fully sedated by the lowest doses of all three agents.

In conclusion, the present study reveals that clove oil, PE and benzocaine can be safely used as sedatives in culture of wels. Although the recovery time was slightly longer than the other, $100 \ \mu l \ L^{-1}$ clove oil is the most effective agent for wels due to not only smaller cost for fish farms but also a lesser polluting matter for the environment. However, Gomulka *et al.*, 2015 emphasized that this agent's therapeutic index is low. On the other hand, it is clear that there is obviously a need to regulate of laws for using of anesthetics in the countries except EU and USA.

Table 2. Physiological responding time	s (s) for clove oil, 2-phenoxyethanol a	nd benzocaine. Data are presented as mean±sd.
--	---	---

Clove oil		Concentrations (µl L ⁻¹)					
Stages	25	50	75	100	125		
I1	24.57 ± 3.87^{b}	18.86±2.79 ^b	16.86±4.30 ^{ab}	$13.14{\pm}1.68^{a}$	$10.29{\pm}2.18^{a}$		
I 2	38.00 ± 4.32^{b}	34.85 ± 9.48^{ab}	33.57±5.91 ^{ab}	31.14±2.85 ^{ab}	27.86±5.15 ^a		
I 3	-	79.14±14.87 ^b	58.71±6.40 ^b	39.43±4.69 ^a	37.14±5.21 ^a		
I 4	-	$189.00{\pm}6.43^{d}$	110.57±6.48°	69.00 ± 6.48^{b}	47.42±4.65ª		
R 1	-	32.43±4.69 ^b	19.29±5.41ª	38.43 ± 7.70^{b}	104.67±7.05°		
R 2	19.29±2.56ª	130.00±26.48 ^b	112.57±10.31 ^b	139.00±21.16 ^b	247.33±16.17°		
R 3	69.86±11.04 ^a	139.86±9.67 ^b	170.43±7.98°	253.86±14.78 ^d	475.33±12.27 ^e		
PE		Concentrations (μ l L ⁻¹)					
Stages	250	500	750	1000	1250		
I1	43.71±4.03 ^b	32.29 ± 7.57^{b}	17.71±2.75 ^a	16.14±3.02 ^a	14.14±4.60 ^a		
I 2	66.29±5.94°	46.00 ± 9.88^{b}	33.29±5.65 ^b	26.29±4.07 ^{ab}	21.86±5.08ª		
I 3	-	87.86±14.89 ^b	75.57±16.45 ^b	47.43 ± 4.96^{a}	38.29±8.90 ^a		
I 4	-	174.86 ± 10.59^{d}	124.00±11.20°	88.71 ± 8.50^{b}	71.71±6.78 ^a		
R 1	-	20.00±7.33ª	27.71±3.45 ^a	32.43±6.48ª	106.57±35.26 ^b		
R 2	-	44.00±11.46 ^a	64.14±13.67 ^a	150.29±25.79 ^b	264.71±45.80 ^b		
R 3	-	107.29±8.26 ^a	142.43±20.93 ^b	201.43±12.19°	322.71 ± 7.43^{d}		
Benzocaine		Concentrations (mg L ⁻¹)					
Stages	25	50	75	100	125		
I 1	37.29±5.19°	32.29±4.39 ^{bc}	28.14±2.34 ^b	18.00±4.43ª	11.57±2.30 ^a		
I 2	-	44.00 ± 8.37^{a}	39.29±6.10 ^a	36.29±3.99ª	31.57±3.95 ^a		
I 3	-	115.14±27.69 ^b	107.71±22.82 ^b	$81.43{\pm}12.9^{ab}$	$65.14{\pm}15.89^{a}$		
I 4	-	156.14±8.75 ^d	126.43±8.00°	103.00±6.90 ^b	81.71 ± 6.90^{a}		
R 1	-	13.00±2.77 ^a	14.28±3.04 ^a	16.86 ± 2.34^{a}	17.57±6.88 ^a		
R 2	-	83.71±9.70 ^a	135.84±9.26 ^b	191.43±29.17°	258.86±43.83 ^d		
R 3	-	120.57±4.86 ^a	149.29±9.48 ^b	215.71±19.99°	352.14±12.20 ^d		

In all lines, means with different superscripts are significantly different from each other (P<0.05).

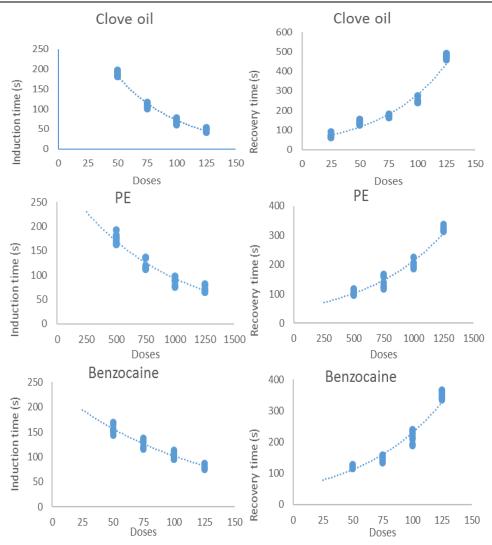


Figure 1. Induction and recovery times (s) relation to anesthetic doses for Wels (n=7 for each trial).

Acknowledgements

Special thanks to owner and staffs of Tuğçe Ltd. for their complimentary helps during the experiment.

References

- Akbulut., B., Çavdar, Y., Çakmak, E., Aksungur, N., 2011. Use of clove oil to anesthetize larvae of Russian Sturgeon (*Acipencer gueldenstadtii*). Jour. Appl. Ichthy. 27, 618-621. DOI: 10.1111/j.1439-0426.2010.01653.x
- Bagheri, T., Imanpour, M. R., 2011. The efficacy, physiological responses and hematology of Persian sturgeon, *Acipencer percicus*, to clove oil as an anesthetic agent. Turkish Jour. Fish. Aqua. Sci. 11, 477-483. DOI: 10.4194/1303-2712-v11_20
- Burka, J.F., Hammell, K.L., Horsberg, T.F., Johnson, G.R., Rainnie, D.J. and Speare, D.J., 1997. Drugs in salmonid aquaculture. J Vet Pharmacol Ther. 20, 333-349. DOI: 10.1046/j.1365-2885.1997.00094.x
- Gilderus. P.A. and Marking. L.L., 1987. Comparative

efficacy of 16 anaesthetic chemicals on rainbow trout. N. Am. J. Fish Manag. 7, 288-292.

- Gullian, M. and Villanueva. J., 2009. Efficacy of tricaine methanesulphonate and clove oil as anaesthetics for juvenile cobia *Rachycentron canadum*. Aquaculture Res. 40, 852-860. DOI: 10.1111/j.1365-2109.2009.02180.x
- Heo. G.J. and Shin. G., 2010. Efficacy of benzocaine as an anaesthetic for Crucian carp (*Carassius carassius*). Vet Anaesth Analg. 37, 132-135. DOI: 10.1111/j.1467-2995.2009.00510.x
- Hseu, J.R., Yeh, S.L., Chu, Y.T. and Ting, Y.Y., 1998. Comparison of efficacy of five anaesthetic goldlined sea bream, Sparus sarba. Acta Zool, Taiwan, 9, 35-41.
- Iversen. M., Finstad. B., Mac Kinley. R.S. and Eliiassen. R.S., 2003. The efficacy of metomidate, clove oil, Aqui-S and Benzoak as anaesthetics in atlanti salmon (*Salmo salar* L.) smolts, and their potential stressreducing capacity. Aquaculture. 221, 549-566. DOI:10.1016/S0044-8486(03)00111-X
- Gomulka, E., Czerniak, E., Dagowski, J., Luczynski, M., Szczerbowski, A., Szkudlarek, M., 2015. Effects of propofol and carbondioxide on acid-base balance in

Siberian sturgeon. Pol.Jour. Vet. Sci. 2, 267-272. DOI 10.1515/pjvs-2015-0035

- Gomulka, P., Dagowski, J., Wlasow, T., Szczepkowski, M., Czerniak, E., Ziomek, E., Szcerbowski, A., Luczynski, M., Szkudlarek, M., 2015. Haematological and biochemical blood profile in Russian sturgeon following propofol and eugenol anesthesia. Turk. J. Fish. Aqua. Sci. 15, 13-17. DOI: 10.4194/1303-2712v15_1_02
- Gökçek, K., Öğretmen, F., 2011. Comparative efficacy of three anesthetic agents in Himri Barbel, *Carasobarbus luteus* (Heckel, 1843) under controlled conditions. Jour. Ani. Vet. Sci. 10, 3350-3355. DOI: 10.3923/javaa.2011.3350.3355
- Marking. L.L. and Meyer. E.P., 1985. Are better anesthetics needed in fisheries? Fisheries. 10, 2-5.
- Mylonas. C.C., Cardinaletti. G., Sigelaki. I. and Polzonetti-Magni. A., 2005. Comparative efficacy of clove oil and 2-phenoxyethanol as anaesthetics in aquaculture of European sea bass (*Dicentrarchus labrax*) and gilthead sea bream (*Sparus auratus*) at different temperatures. Aquaculture. 246, 467-481. DOI:10.1016/j.aquaculture.2005.02.046
- Öğretmen, F., Gökçek, K., 2013. Comperative efficacy of three anesthetic agents on juvenile African catfish, *Clarias gariepinus* (Burchell, 1822). Turk. J. Fish. Aqau. Sci. 13, 51-56. DOI: 10.4194/1303-2712v13_1_07
- Öğretmen, F., Gölbaşı, S., Inanan, B. E., Kizak, V., Kayım, M., 2014. Use of clove oil and eugenol to anesthetize fingerling Shabut *Barbus grypus*. North American Journal of Aquaculture. 76, 9-13. DOI: 10.1080/15222055.2013.824942
- Pawar. H.B., Sanaye. S.V., Sreepada. R.A., Harish. V., Suryavanshi. U. and Ansari. Z.A., 2011. Comparative efficacy of four anaesthetic agents in the yellow seahorse, *Hippocampus kuda* (Bleeker, 1852).

Aquaculture. DOI:10.1016/j.aquaculture.2010.12.007

- Ross. L.G., Ross. B., 1999. Anesthetics and sedative techniques fro aquatic animals. Blackwell Sci, Oxford. pp.159.
- Theinpoint. D., Niemegeers. C.J.E., 1965. 7464- a new potenet anesthetşic in fish. International Zoo. Year book 5, 202-205.
- Treves-Brown. K.M., 2000. Anaesthetics in applied fish pharmacology. Kluwer Academic Publishers, Dordrecht, Netherlands. pp.206-217.
- Tytler P., Hawkins. A.D. 1981. Vivisection, anesthetics and minor surgery. In: Hawkins AD (ed.) Aquarium Sys. Academic Press, New York, pp.247-278.
- Velisek. J., Wlasow. T., Gomulka. P., Svobodava. Z., Novotny. L., Ziomek. A., 2006. Effects of clove oil anaesthesia on catfish (*Siluris glanis* L.). Acta Vet Brno. 75, 99-106. http://dx.doi.org/10.2754/avb200675010099
- Velisek. J., Stara. A., Li Z.H., Silovska. S., Turek. J., 2011. Comparison of effects of four anaesthetics on blood chemical profiles and oxidative stress biomarkers in rainbow trout. Aquaculture. 310, 369-375. DOI:10.1016/j.aquaculture.2010.11.010
- Weber. R.A., Peleterio. J.B., Garcia Martin. L.O., Aldegunde. M., 2009. The efficacy of 2phenoxyethanol, metomidate, clove oil and MS-222 as anaesthetic agents in the Senegalese sole (*Solea* senegalensis Kaup, 1858). Aquaculture. 288, 147-150. DOI:10.1016/j.aquaculture.2008.11.024
- Zahl, I., H., Samuelsen, O., Kiessling, A., 2012. Anesthesia of farmed fish: implications for welfare. Fish Physiology and Biochemistry. 38, 201-218. DOI: 10.1007/s10695-011-9565-1
- Zar. J.H.,1999. Biostatistical Analysis, 4th ed. New Jersey, Prentice Hall. p929.

311,155-161.