

Rec. Nat. Prod. 14:4 (2020) 307-311

records of natural products

# Two New Compounds from the Deep-Sea-Derived Fungus Aspergillus sp. YPGA8

Meng-Hua Pan<sup>®1</sup>, Zhi-Yong Tian<sup>®1</sup>, Hui Yang<sup>®1\*</sup>, Wei Xu <sup>®2</sup>, Chen Pan<sup>®1</sup>, Zhong-Bin Cheng<sup>®1, 3\*</sup> and Qin Li <sup>®1,3\*</sup>

<sup>1</sup> School of Pharmacy, Henan University, Kaifeng, Henan, 475004, China
<sup>2</sup> Key Laboratory of Marine Biogenetic Resources, Third Institute of Oceanography, Ministry of Natural Resources, Xiamen 361005, China
<sup>3</sup> Eucommia Ulmoides Cultivation and Utilization of Henan Engineering Laboratory, Kaifeng 475004, China

(Received November 23, 2019; Revised December 09, 2019; Accepted December 11, 2019)

**Abstract:** Chemical examination of a fraction of the EtOAc extract of a marine-derived fungus *Aspergillus* sp. YPGA8 resulted in the isolation of two new compounds, namely aspertriols A–B (1–2). Compounds 1–2 possess an identical 2,3,4-trihydroxybutoxy moiety, which is rarely found in natural products. Their structures were determined by extensive analyses of spectroscopic data (1D and 2D NMR, HRESIMS). The bioassay study revealed that compounds 1 and 2 were inactive toward  $\alpha$ -glucosidase.

**Keywords:** *Aspergillus* sp.; aspertriols A–B; isolation; identification. © 2020 ACG Publications. All rights reserved.

## 1. Plant Source

In our continuous efforts to characterize bioactive natural products from deep-sea-derived *Aspergillus* strains [1, 2], chemical examination of the fermentation broth of a deep-sea derived fungus *Aspergillus* sp. YPGA8 led to the isolation of two new compounds 1-2 (Figure 1).

Fungus YPGA8 was isolated from a deep-sea sediment sample that was collected at a depth of 4993 m in the Yap Trench of West Pacific Ocean. The strain was identified as *Aspergillus* sp. based on microscopic examination and by internal transcribed spacer (ITS) sequencing. The ITS sequence has been deposited in GenBank (http://www.ncbi.nlm.nih.gov) with accession number MG835905. The strain YPGA8 (MCCC 3A01017) was deposited at the Marine Culture Collection of China.

The article was published by ACG Publications

http://www.acgpubs.org/journal/records-of-natural-products July-August 2020 EISSN:1307-6167 DOI: http://doi.org/10.25135/rnp.162.19.11.1487

<sup>\*</sup> Corresponding author: E- Mail:<u>10200097@vip.henu.edu.cn (H. Yang)</u>, <u>czb360@126.com (Z-B. Cheng)</u>, <u>liqin6006@163.com</u> (Q. Li).

## 2. Previous Studies

Marine-derived fungi have been recognized as a new source for producing secondary metabolites possessing fascinating structures and versatile biological activities. Many metabolites have been used in pharmaceuticals to exploit lead compounds or have become attractive targets for medicinal or synthetic chemists. The *Aspergillus* species, frequently isolated from marine marine organisms or sediments, have been proved to be highly prolific, previous chemical study of marine-derived *Aspergillus* strains resulted in many structurally unique and bioactive metabolites, including alkaloids [3, 4], cyclopeptides [5], butenolide derivatives [6], nortritterpenoids [7], nitrobenzoyl sesquiterpenoids [8], diphenyl ethers [9], and pentaketides [10], some exhibited pronounced biological activities, such cytotoxic, antibacterial, lipid-lowering, and  $\alpha$ -glucosidase inhibitory effects.

#### 3. Present Study

The fermentation was carried out in 10 Fernbach flasks ( $10 \times 500 \text{ mL}$ ), each containing 60 g of rice. Distilled H<sub>2</sub>O (100 mL) was added to each flask, and the contents were soaked overnight before autoclaving at 15 psi for 30 min. After cooling to room temperature, each flask was inoculated with 5.0 mL of the spore inoculum and incubated at 25 °C for 25 days. The fermented material was extracted with EtOAc for three times ( $3 \times 5000 \text{ mL}$ ). After evaporation under vacuum, the extract (1.0 g) was chromatographed over ODS silica gel column chromatography (CC) (MeOH/H<sub>2</sub>O: 20:80 $\rightarrow$ 100:0) to give ten fractions (F1 to F10). Fraction F3 was again purified on an ODS silica gel CC eluted with MeOH/H<sub>2</sub>O (40:60 $\rightarrow$ 70:30) as a mobile phase to afford four fractions (F3c1–F3c4). F3c2 was subjected to RP-HPLC with a mobile phase of MeOH/H<sub>2</sub>O (45:55, 2 mL/min) to yield **1** (2.6 mg) and **2** (1.7 mg).

Aspertriol A (1): colorless oil;  $[\alpha]^{25}_{D}$  –4 (*c* 0.2, MeOH); UV (MeOH)  $\lambda_{max}$  223, 294 nm; <sup>1</sup>H and <sup>13</sup>C NMR data, see Table 1; HRESIMS *m*/*z* 287.1132 [M + HCOO]<sup>-</sup> (calcd. for C<sub>13</sub>H<sub>19</sub>O<sub>7</sub>, 287.1131).

Aspertriol B (2): colorless oil;  $[\alpha]^{25}_{D}$  –12 (*c* 0.1, MeOH); UV (MeOH)  $\lambda_{max}$  222, 295 nm <sup>1</sup>H and <sup>13</sup>C NMR data, see Table 1; HRESIMS *m*/*z* 265.1057 [M + Na]<sup>+</sup> (calcd. for C<sub>12</sub>H<sub>18</sub>O<sub>5</sub>Na, 265.1052).

 $\alpha$ -Glucosidase Assay: The  $\alpha$ -glucosidase inhibitory effect was assessed as described in the literature [1, 11].



Figure 1. Structures of compounds 1 and 2 from Aspergillus sp. YPGA8.

Compund **1** had a molecular formula of  $C_{12}H_{18}O_5$ , as established by the HRESIMS and NMR data (Table 1), requiring four degrees of unsaturation. The <sup>1</sup>H NMR and HSQC spectra showed six olefinic protons ( $\delta_H$  5.70, 6.00, 6.39, 6.78, 7.08, 7.25), an olefinic methyl ( $\delta_H$  1.82), two oxygenated methylenes ( $\delta_H$  4.23, 4.01; 3.54, 3.37,), and two oxygenated methines ( $\delta_H$  3.59, 3.37), while the <sup>13</sup>C NMR resonances exhibited six olefinic carbons for three double bonds ( $\delta_C$  116.9, 121.8, 126.6, 135.8, 138.1, 139.1), one carbonyl carbon for an ester group ( $\delta_C$  166.0). All the four degrees of unsaturation

were fully covered by three double bonds and a carbonyl carbon, suggesting compound **1** was acyclic. The <sup>1</sup>H-<sup>1</sup>H COSY relationship (Figure 2) from H-2 ( $\delta_c$  5.70) to H-8 ( $\delta_H$  1.82) established the spin system from C-2 ( $\delta_c$  116.9) to C-8 ( $\delta_c$  18.5) (unit A), in which three disubstituted double bonds were found to be located at C-2/C-3, C-4/C-5, C-6/C-7. Additional COSY correlations from H<sub>2</sub>-1' ( $\delta_H$  4.23, 4.01) to H<sub>2</sub>-4' ( $\delta_H$  3.37, 3.54) established a 2,3,4-trihydroxybutoxy moiety (unit B). The coupling constant between H-2' and H-3' (6.8 Hz) of compound **1** was identical with those of reported analogues erythrin and montagnetol [12], suggesting that the 2,3,4-trihydroxybutoxyl moiety in **1** had a 2*S*\*, 3 *R*\*-configuration. The above two framents (units A and B) were further connected by HMBC correlations from H-2 ( $\delta_H$  5.70), H-3 ( $\delta_H$  7.25), H<sub>2</sub>-1' ( $\delta_H$  4.23, 4.01) to the ester carbon C-1 ( $\delta_C$  166.0). The configurations of the double bonds at  $\Delta^2$ ,  $\Delta^4$ , and  $\Delta^6$  were assigned to be *Z*, *Z*, and *E* configurations by the coupling constants  $J_{2,3}$  (11.3 Hz),  $J_{4,5}$  (11.5 Hz), and  $J_{6,7}$  (13.9 Hz), respectively, which were also supported by the NOESY correlations of H-2 ( $\delta_H$  6.39), H-6 ( $\delta_H$  6.78)/H-8 ( $\delta_H$  1.82) (Figure 1). Thus, the structure of **1** was determined as depicted and was named aspertriol A.

**Table 1.** <sup>1</sup>H (400 Hz) and <sup>13</sup>C NMR (100 Hz) Data of **1** and **2** in DMSO- $d_6$  ( $\delta$  in ppm)

No.	1		2	
	$\delta_{\rm H}$ (mult., <i>J</i> in Hz)	$\delta_{C}$	$\delta_{\rm H}$ (mult., J in Hz)	$\delta_{\mathrm{C}}$
1		166.0		166.4
2	5.70, d (11.3)	116.9	5.94, d (15.1)	120.0
3	7.25, dd (11.8, 11.3)	139.1	7.28, dd (15.1, 11.6)	144.8
4	7.08, dd (11.8, 11.5)	121.8	6.34, dd (14.4, 11.6)	127.6
5	6.39, dd (11.5, 11.1)	138.1	6.70, dd (14.4,11.4)	141.3
6	6.78, dd (13.9, 11.1)	126.6	6.21, dd (14.6, 11.4)	131.3
7	6.00, dq (13.9, 6.8)	135.8	6.01, dd (14.6, 7.0)	135.2
8	1.82, d (6.8)	18.5	1.80, d (7.0)	18.3
1′	4.23, dd (11.4, 2.4) 4.01, dd (11.3, 7.1)	66.1	4.24, dd (11.0, 2.0) 4.01, dd (10.9, 7.2)	66.2
2'	3.59, ddd (7.1, 6.9, 2.4)	69.4	3.58, ddd (7.2, 6.9, 2.4)	69.5
3'	3.37, m	72.4	3.38, m	72.3
4′	3.54, m 3.37, m	63.1	3.55, m 3.38, m	63.1





Figure 2.  $^{1}H^{-1}H COSY (-)$ , HMBC ( $\checkmark$ ), and NOE ( $\checkmark$ - $\checkmark$ ) correlations of 1 and 2.

Compound **2**, a colorless oil, had a molecular formula of  $C_{12}H_{18}O_5$  as determined by the HRESIMS and NMR data. The <sup>1</sup>H NMR, <sup>13</sup>C NMR, and HSQC spectra exhibited signals for three disubstituted double bonds ( $\delta_H$  5.94, 6.01, 6.21, 6.34, 6.70, 7.28;  $\delta_C$  120.0, 127.6, 131.3, 135.2, 141.3, 144.8), an olefinic methyl ( $\delta_H$  1.80;  $\delta_C$  18.3), an ester carbon ( $\delta_C$  166.4), two oxygenated methylenes ( $\delta_H$  3.38, 3.55, 4.01, 4.24;  $\delta_C$  66.2, 63.1), and two oxygenated methines ( $\delta_H$  3.58, 3.38;  $\delta_C$  69.5, 72.3). The aforementioned information was almost identical with that of **1**. The different chemical shifts and coupling constants of the double bonds suggested that **2** was an olefinic isomer of **1**, the structure of **2** was further established by detailed analyses of the 2D NMR data (Figure 1). The coupling constants  $J_{2,3}$  (15.1),  $J_{4,5}$  (14.4), and  $J_{6,7}$  (14.6) defined the double bonds  $\Delta^2$ ,  $\Delta^4$ , and  $\Delta^6$  all to be *E* configurations, which were also supported by the NOESY correlations of H-4 ( $\delta_H$  6.34)/H-2 ( $\delta_H$  5.94), H-6 ( $\delta_H$  6.21)/H-4, and H<sub>3</sub>-8 ( $\delta_H$  1.80)/H-6 (Figure 2). The NMR data of C-1' to C-4' and the coupling constant between 2' and H-3' (6.9 Hz) implied that the 2,3,4-trihydroxybutoxy moiety in **2** was the same as that of **1**. Thus, the structure of compound **2** was determined as depicted and was given the trivial name aspertriol B.

Compounds 1 and 2 were screened for their inhibitory activities against  $\alpha$ -glucosidase. As results, compounds 1 and 2 exhibited inhibition rate less than 30% at the concentration of 200  $\mu$ M, while the positive control acarbose exhibited an IC<sub>50</sub> values of 190.2  $\mu$ M.

#### Acknowledgments

This work was supported by the grants from National Key R&D Program of China (2017YFD0600702-2), China Ocean Mineral Resources R&D Association (COMRA) Program (DY135-B2-01), China Postdoctoral Science Foundation (2018M630815), and First-Class Discipline Construction Project of Henan University (2018YLZDCG03).

#### **Supporting Information**

Supporting Information accompanies this paper on <u>http://www.acgpubs.org/journal/records-of-natural-products</u>

#### ORCID 💿

Meng-Hua Pan: <u>0000-0002-8219-2522</u> Zhi-Yong Tian: <u>0000-0001-6913-235X</u> Hui Yang: <u>0000-0001-5250-4647</u> Wei Xu: <u>0000-0002-3265-7475</u> Chen Pan: <u>0000-0002-0261-9363</u> Zhong-Bin Cheng: <u>0000-0003-0942-6422</u> Qin Li: <u>0000-0001-8295-6230</u>

### References

- [1] Z. Cheng, Y. Li, W. Liu, L. Liu, J. Liu, W. Yuan, Z. Luo, W. Xu and Q. Li (2019). Butenolide derivatives with α-glucosidase inhibitions from the deep-sea-derived fungus Aspergillus terreus YPGA10, Mar. Drugs 17, 332 (9 pages).
- [2] Y. L. Li, W. Liu, W. Xu, X. Zeng, Z. B. Cheng and Q. Li (2020). Aspterrics A and B, new sesquiterpenes from deep sea-derived fungus *Aspergillus terreus* YPGA10, *Rec. Nat. Prod.* 14, 18-22.
- [3] K. Nakanishi, M. Doi, Y. Usami, T. Amagata, K. Minoura, R. Tanaka, A. Numata and T. Yamada (2013). Antheolorins A-F, novel cytotoxic metabolites from a sea urchin-derived *Aspergillus versicolor*, *Tetrahedron* **69**, 4617-4623.
- [4] X. Liang, X. Zhang, X. Lu, Z. Zheng, X. Ma and S. Qi (2019). Diketopiperazine-type alkaloids from a deep-sea-derived *Aspergillus puniceus* fungus and their effects on liver X receptor α, *J Nat. Prod.* 82, 1558-1564.

- [5] J. Peng, H. Gao, X. Zhang, S. Wang, C. Wu, Q. Gu, P. Guo, T. Zhu and D. Li (2014). Psychrophilins E-H and versicotide C, cyclic peptides from the marine-derived fungus *Aspergillus versicolor* ZLN-60, *J. Nat. Prod.* **77**, 2218-2223.
- [6] Y. Sun, J. Liu, L. Li, C. Gong, S. Wang, F. Yang, H. Hua and H. Lin (2018). New butenolide derivatives from the marine sponge-derived fungus *Aspergillus terreus*, *Bioorg. Med. Chem. Lett.* **28**, 315-318.
- [7] F.D. Kong, X.L. Huang, Q.Y. Ma, Q.Y. Xie, P. Wang, P.W. Chen, L.M. Zhou, J.Z. Yuan, H.F. Dai, D.Q. Luo and Y.X. Zhao (2018). Helvolic acid derivatives with antibacterial activities against *Streptococcus agalactiae* from the marine-derived fungus *Aspergillus fumigatus* HNMF0047, *J. Nat. Prod.* 81, 1869-1876.
- [8] Y. Tan, B. Yang, X. Lin, X. Luo, X. Pang, L. Tang, Y. Liu, X. Li and X. Zhou (2018). Nitrobenzoyl sesquiterpenoids with cytotoxic activities from a marine-derived Aspergillus ochraceus fungus, J. Nat. Prod. 81, 92-97.
- [9] Y. Wu, Y. Chen, X. Huang, Y. Pan, Z. Liu, T. Yan, W. Cao and Z. She (2018). α-Glucosidase inhibitors: diphenyl ethers and phenolic bisabolane sesquiterpenoids from the mangrove endophytic fungus *Aspergillus flavus* QQSG-3, *Mar. Drugs* 16, 307.
- [10] Q. Wang, X.Q. Yang, C.P. Miao, L.H. Xu, Z.T. Ding, Y.B. Yang and L.X. Zhao (2018). A new pair of pentaketide diastereoisomers from *Aspergillus melleus* YIM PHI001, *Rec. Nat. Prod.* 12, 216-221.
- [11] S. Jibril, H. Mohd Sirat and N. Basar (2017). Bioassay-guided isolation of antioxidants and α-glucosidase inhibitors from the root of *Cassia sieberiana* D.C. (Fabaceae), *Rec. Nat. Prod.* **11**, 406-410.
- [12] J.-F. Basset, C. Leslie, D. Hamprecht, A. J. P. White and A. G. M. Barrett (2010). Studies on the resorcylates: biomimetic total syntheses of (+)-montagnetol and (+)-erythrin, *Tetrahedron Lett.* **51**, 783-785.

A C G publications

311